Investigation of Different Synthetic Approaches towards Methyl 2,6-Dideoxy-α-D-*arabino*- and -α-D-*lyxo*-hexopyranosides

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New synthetic routes for making available different methyl 2,6-dideoxyhexo-pyranosides are reported. Starting with methyl 6-deoxy-2,3-O-isopropylidene- α -D-manno- and $-\alpha$ -D-talo-pyranoside the respective methyl 2,6-dideoxy- α -D- α -D-talo-pyranosides were obtained. The investigation of different synthetic approaches, including the selective monoalkylation of vicinal diols, using tin-ether intermediates, and the radical induced mono-deoxygenation of cyclic thionocarbonates, is described. In the talo series an unusual intramolecular acetal migration was observed. The migration product was used in the synthesis of some methyl 2,6-dideoxy- α -D-lyxo-hexopyranosides.

Owing to their occurrence in Nature we were interested in the synthesis of methyl 2,6-dideoxyhexopyranosides.¹ They are found as the carbohydrate aglycones in a variety of antibiotic compounds. 2a,b Many of those antibiotics show antitumor activity and are active against gram positive bacteria but they are also very toxic. Thus their synthesis could provide useful material for further studies of biological and medical interest. Our aim was to develop short synthetic sequences towards the sugar aglycones starting with a common chiral precursor. Under these investigations methyl 6-deoxy-2,3-O-isopropylidine-α-D-mannopyranoside 1 has been found to be a useful starting material (Fig. 1). Different synthetic approaches from 1, yielding the methyl glycosides of five naturally occurring carbohydrate antibiotics, are now described.

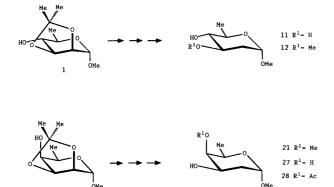


Fig. 1.

Results and discussion

In order to synthesize methyl 2,6-dideoxy-α-D-arabinohexopyranosides, namely the methyl glycosides of D-olivose 11^{3a-c,4a} and D-oleandrose 12,^{4a,b} the benzyl ether 3 was prepared.⁵ Using 3, two different synthetic routes were investigated. The first one made use of the activation of the equatorial OH-3 by the dibutyltin oxide method.⁶ After optimization of the reaction conditions the 3-O-methyl ether 4 and 3-O-benzyl ether 5 were obtained (95% and 86%, respectively) (Fig. 2). Preparation of the intermediate dibutyltin ether in methanol led in all cases to only partial transformation in the following alkylation reaction, but unchanged starting diol could be recovered. The best results were obtained when preparing the tin ether intermediate in toluene with azeotropic removal of water and with activation of the alkylation reaction with tetrabutylammonium iodide.⁷ It is interesting to note that no methylation at OH-2 was observed, whereas the 2,4-di-O-benzyl ether 6 (10%) was obtained in the benz-

Fig. 2.

ylation reaction under similiar conditions. An attempt to use bis(tributyltin) oxide instead of dibutyltin oxide did not improve the regioselectivity of the benzylation. The subsequent deoxygenation at C-2 for 4 and 5 could be achieved via the xanthogenates 7 and 8 (91 % and 83 %), followed by radical deoxygenation using tributyltin hydride (Bu₃SnH) to yield the 2,6-dideoxy-sugars 9 and 10 (71% and 66%). Because of their toxicity, the use of tin compounds in the synthesis of substances with biological and medicinal applications is rather inappropriate. In those cases tris (trimethylsilyl)silane (TTMSS)8 can be used as a substitute as exemplified by the synthesis of the 2,6-dideoxy-sugar 9 (84%). Finally, methyl α -D-olivoside 11 and methyl α -D-oleandroside 12 were obtained after debenzylation of 9 and 10 under standard hydrogenation conditions (90% yield each).

Another route towards 2,6-dideoxygenated sugars might be via cyclic 2,3-O-thionocarbonates by applying radical induced mono-deoxygenation. This was attempted, preparing the 2,3-thionocarbonate 13¹⁰ (81%) from diol 3. However, treating 13 with 3 equivalents of Bu₃SnH in refluxing toluene yielded two main products. The regioselectivity of this reaction was low (4:3), giving the 2,6-dideoxy sugar 14 (41%) and the 3,6-dideoxygenated isomer 15 (30%). Although 14 could be transformed into the 3-O-methyl ether 9 (85%) by methylation with MeI-NaH, this route yields the antibiotic sugar 12 in lower yield than the method described above.

To obtain the D-lyxo isomers of methyl 2,6-dideoxy-hexopyranosides, 1 was transformed into its epimer 2 using the oxidation/reduction sequence as described by Eis et al.¹¹ Methylation or benzylation at OH-4 and hydrolysis of the isopropylidene protecting group yielded the diols 16 and 17 (67% and 87%) from the alcohol 2. Different routes to achieve deoxygenation at C-2 using 16 and 17 were investigated (Fig. 3). Aspinall et al.¹² obtained a mixture of isomeric benzyl ethers when treating the benzyl α -L-enantiomer of 17 with

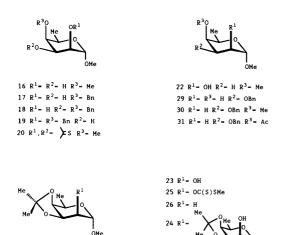


Fig. 3.

bis(tributyltin) oxide followed by benzyl bromide. A similiar attempt to activate 17 with dibutyltin oxide resulted in the formation of the 3,4-di-O-benzyl ether 18 (55%) and the 2,4-di-O-benzyl ether 19 (36%). Even the high total yield did not make this route very attractive, owing to a tedious separation procedure and a moderate yield of the desired 3,4-di-O-benzyl ether 18. It was therefore decided to proceed via the cyclic 2,3-thionocarbonate 20 which could be synthesized by using thiocarbonyldiimidazole (TCDI) in refuxing THF (73%). As in the manno series regioselectivity in the reaction of 20 with Bu₃SnH in refluxing toluene was low (3:2) but favored the 2,6-dideoxygenated sugar, thus giving methyl α-D-chromoside A 21^{13a-c} through a short synthesis. The yields were, however, only moderate (37% for 21 and 26% for 22) and therefore a more expedient approach towards 2,6-dideoxy-a-d-lyxo-hexopyranosides seemed desirable.

It has been described 14 that direct isopropylidenation of methyl 6-deoxy-α-L-talopyranoside yields the 3.4-O-isopropylidene compound as the major product. On isopropylidenation of the α-D-enantiomer two products were obtained. The major product proved to be the 3,4-O-isopropylidene compound 23 (60%), while compound 2 was obtained in 21% yield. The formation of 2 prompted us to investigate the possibility of a more direct synthesis of 23 via an acetal migration. 15 Such a migration indeed took place when 2 was treated with concentrated sulfuric acid in acetone under anhydrous conditions. In addition to 23 (74%) small amounts of 2 (8%) and of the disaccharide 24 (6%) were obtained. Thus, an easy access to an OH-2 talopyranoside was established. Deoxygenation of 23 was performed via the xanthogenate 25 (88%) and successive radical deoxygenation with TMSS yielded the 2,6-dideoxy-sugar 26 (79%). The subsequent hydrolysis of the isopropylidene protecting group was performed under mild conditions to avoid the formation of a methyl β-glycoside. The best results were obtained using methanolic HCl which gave methyl α-D-olioside 27^{13,16} in 81% yield. To obtain the methyl α-D-glycosides of chromose A 21 and chromose D 28. 16,17 the diol 27 was benzylated regioselectively at OH-3 (83%) using activation via an intermediate dibutyltin ether and tetrabutylammonium iodide as a catalyst. 13a The benzyl ether 29 was then methylated or acylated to yield the fully protected compounds 30 (79%) and 31 (86%) respectively. Both sugars could be debenzylated under standard hydrogenation conditions to yield methyl α-D-chromoside A 21 (88%) and methyl α-D-chromoside D 28 (86%).

The reported investigations show the synthetic potential of a carbohydrate-derived synthon, namely methyl 6-deoxy-2,3-O-isopropylidene- α -D-mannopyranoside 1, allowing the preparation of the five target compounds 11, 12, 21, 27 and 28. In contrast with the rather individual routes reported for the synthesis of these sugar aglycones, broad access to dideoxygenated hexopyranosides was established by using a common chiral precursor.

Experimental

General methods. For experimental details see Ref. 5. All compounds not described in detail show spectroscopic properties in full agreement with the assigned structures. In the talo series it is possible to distinguish between OH-2 and OH-3 substitution by comparing the ¹³C NMR data. The compounds having a free OH-2 show typical values around 101.9–103.6 ppm for the anomeric carbon atom, whereas 2-O-substitution results in lower resonances (95.8–98.6 ppm) for C-1. The following mixtures (v/v) were used for TLC and flash and column chromatography: A (ethyl acetate-hexane 1:1), B (ethyl acetate-hexane 1:2), C (ethyl acetate-hexane 1:3), D (diethyl ether-hexane (5:4), E (dichloromethane-hexane-diethyl ether 10:4:1).

General procedure 1: synthesis of mono-xanthogenates 7, 8 and 25. A suspension of NaH (1.5 mmol) in 5 ml THF was cooled to 0°C under nitrogen. A mixture of the alcohol (1.0 mmol) and imidazole (10 mg) in 10 ml THF was then added dropwise. The suspension was allowed to warm to room temperature for 15 min, and then CS₂ (3.0 mmol) was added followed, after an additional 15 min, by CH₃I (2.0 mmol). When TLC indicated the complete consumption of the starting alcohol (15-45 min) acetic acid was added to destroy the excess of NaH. The suspension was filtered, the solvent was removed and the residue was extracted with 15 ml ether. The ether was washed with sat. NaHCO₃ (5 ml) and water $(2 \times 10 \text{ ml})$ and dried over MgSO₄. After the removal of the solvents the remainder was purified by flash chromatography to yield the xanthogenates. For physical data of the compounds 7, 8 and 25 see below.

General procedure 2: deoxygenation of mono-xanthogenates 7, 8 and 25. A solution of the xanthogenate (1.0 mmol) in 10 ml of toluene was heated to 80°C under nitrogen. A mixture of TTMSS (1.2 mmol) and azoisobutyronitrile (AIBN) (0.1 mmol) in 5 ml toluene was added dropwise over 20 min. When TLC indicated the reaction to be complete (2-4 h), the mixture was evaporated in vacuo and the residue submitted to flash chromatography with an appropriate eluant to yield the deoxy sugars 9, 10 and 26. Reductions with tributyltin hydride were performed as described above at 110°C using 3.0 equiv. of Bu₃SnH and 0.1 equiv. AIBN. For physical data of the compounds 9, 10 and 26 see below.

Methyl 4-O-benzyl-6-deoxy-3-O-methyl- α -D-mannopyranoside (4). A mixture of 3^5 (0.80 g) and dibutyltin oxide (0.74 g) in 30 ml of abs. toluene was refluxed with azeotropic removal of water for 16 h. The resulting solution was concentrated to about 15 ml and CH₃I (5 ml) and tetrabutylammonium iodide (1.10 g) were added. The mixture was stirred at 40° C for 18 h after which time TLC showed complete consumption of the starting material.

The mixture was evaporated and the residue submitted to flash chromatography on silica gel (eluant A) to yield 0.82 g (95%) of 4 as a syrup (R_f =0.38, A) with [α]_D+87.5° (c 0.55). ¹H NMR: δ 1.31 (d, 3 H, J 6.2 Hz, CH₃), 2.44 (d, 1 H, J 2.2 Hz, OH), 3.35 (s, 3 H, CH₃O-1), 3.36 (t, 1 H, J 9.2 Hz, H-4), 3.49 (s, 3 H, CH₃O-3), 3.54 (dd, 1 H, J 9.0, 3.4 Hz, H-3), 3.68 (dq, 1 H, J 9.6, 6.2 Hz, H-5), 4.04 (ddd, 1 H, J 3.4, 2.2, 1.6 Hz, H-2), 4.70 (d, 1 H, J 1.6 Hz, H-1), 4.73 (dd, 2 H, CH₂O-4), 7.35 (m, 5 H, ArH). ¹³C NMR: δ 17.9 (C-6), 54.7 (CH₃O-1), 57.4 (CH₃O-3), 67.0, 67.8 (C-2 and C-5), 75.1 (CH₂O-4), 79.8, 81.7 (C-3 and C-4), 100.0 (C-1), 127.6–138.5 (Ar).

Methyl 3,4-di-O-benzyl-6-deoxy- α -D-mannopyranoside (5) and methyl 2,4-di-O-benzyl-6-deoxy-\alpha-D-mannopyranoside (6). A mixture of 3 (536 mg) and dibutyltin oxide (498 mg) in 20 ml abs. toluene was refluxed with azeotropic removal of water for 18 h. The mixture was concentrated to about 10 ml. Thereafter, tetrabutylammonium iodide (738 mg) and benzyl bromide (855 mg) were added and the mixture was heated at 90°C for 1 h. After evaporation under reduced pressure the residue was chromatographed on silica gel (eluant A) to yield the 2,4-di-O-benzyl ether 6 (80 mg, 10 %, $R_f = 0.78$, A) which was characterized by comparision of its spectra with literature data. 18 Further elution yielded 5 (620 mg, 86 %) as a chromatographically homogeneous $(R_f = 0.64, A)$ syrup, $\lceil \alpha \rceil_D + 45.8^{\circ}$ (c 1.3). ¹H NMR: δ 1.34 (d, 3 H, J 6.0 Hz, CH₃), 2.44 (br s, 1 H, OH), 3.36 (s, 3 H, CH₃O-1), 3.47 (t, 1 H, J 9.2 Hz, H-4), 3.84 (dd, 1 H, J 8.8, 3.4 Hz, H-3), 4.04 (dd, 1 H, J 3.4, 1.6 Hz, H-2), 4.70 (m, 3 H), 4.78 (dd, 2 H, CH₂O), 7.34 (m, 10 H, ArH). ¹³C NMR: δ 18.6 (C-6), 54.9 (CH₃O-1), 67.2 and 68.6 (C-2 and C-5), 72.0 and 75.3 (2×CH₂O), 79.9 and 80.0 (C-3 and C-4), 99.8 (C-1), 127.1-137.7 (Ar).

Methyl 4-O-benzyl-6-deoxy-3-O-methyl-2-O-[(methylthio)thiocarbonyl]- α -D-mannopyranoside (7). The alcohol 4 (460 mg) was processed according to general procedure 1 and the residue purified by column chromatography on silica gel (eluant E) to yield 552 mg (91%) of 7 as a chromatographically homogeneous syrup $(R_{\rm f} = 0.52, E)$ with $\lceil \alpha \rceil_{\rm D} + 10.2^{\circ}$ (c 1.8). ¹H NMR: δ 1.34 (d, 3 H, J 6.2 Hz, CH₃), 2.59 (s, 3 H, SCH₃), 3.37 (s, 3 H, CH₃O-1), 3.42 (t, 1 H, J 9.4 Hz, H-4) 3.44 (s, 3 H, CH₃O-3), 3.75 (dq, 1 H, J 9.2, 6.2 Hz, H-5), 3.76 (dd, 1 H, J 9.6, 3.4 Hz, H-3), 4.77 (d, 1 H, J 1.6 Hz, H-1), 4.80 (dd, 2 H, CH₂O-4), 6.10 (dd, 1 H, J 3.4, 1.6 Hz, H-2), 7.35 (m, 5 H, ArH). ¹³C NMR: δ 18.0 (C-6), 19.0 (SCH₃), 54.9 (CH₃O-1), 57.7 (CH₃O-3), 67.4 (C-5), 75.2 (CH₂O-4), 76.8 (C-2), 80.0, 80.2 (C-3 and C-4), 97.5 (C-1), 127.0-138.5 (Ar), 215.9 (C=S).

Methyl 3,4-di-O-benzyl-6-deoxy-2-O-[(methylthio)thio-carbonyl]- α -D-mannopyranoside (8). The alcohol 5 (802 mg) was treated as described in general procedure 1. Flash chromatography (eluant A) yielded 8 (835 mg, 83%) as a syrup, $[\alpha]_D + 4.0^\circ$ (c 1.25). H NMR: δ 1.35

(d, 3 H, J 6.2 Hz, CH₃), 2.60 (s, 3 H, SCH₃), 3.37 (s, 3 H, CH₃O-1), 3.49 (t, 1 H, J 9.2 Hz, H-4), 3.78 (dq, 1 H, J 9.2, 6.2 Hz, H-5), 4.04 (dd, 1 H, J 9.2, 3.4 Hz, H-3), 4.68 and 4.74 (2 × dd, 4 H, 2 × CH₂O), 4.78 (br s, 1 H, H-1), 6.15 (dd, 1 H, J 3.4, 1.8 Hz, H-2), 7.32 (m, 10 H, ArH). ¹³C NMR: δ 18.4 (C-6), 19.3 (SCH₃), 55.0 (CH₃O-1), 67.6 (C-5), 71.8 (C-2), 75.3 (CH₂O), 77.3 and 78.0 (CH₂O and C-3), 80.1 (C-4), 97.4 (C-1), 127.2–138.0 (Ar), 215.6 (C=S).

Methyl 4-O-benzyl-2,6-dideoxy-3-O-methyl-α-D-arabino-hexopyranoside (9). The xanthogenate 7 (300 mg) was processed according to general procedure 2 and the residue was submitted to column chromatography using a hexane-ether gradient (starting with pure hexane and ending with eluant D) to yield 179 mg (84%) of 9 as a colorless syrup (R_f =0.61, D), [α]_D+125° (c 0.5). ¹H NMR: δ 1.29 (d, 3 H, J 6.2 Hz, CH₃), 1.56 (ddd, 1 H, J 13.2, 11.4, 3.8 Hz, H-2_{ax}), 2.27 (ddd, 1 H, J 13.2, 5.2, 1.4 Hz, H-2_{eq}), 3.03 (t, 1 H, J 9.2 Hz, H-4), 3.31 (s, 3 H, CH₃O-1), 3.44 (s, 3 H, CH₃O-3), 3.67 (m, 2 H), 4.75 (dd, 1 H, J 3.8, 1.4 Hz, H-1), 4.78 (dd, 2 H, CH₂O-4), 7.34 (m, 5 H, ArH). ¹³C NMR: δ 18.1 (C-6), 35.1 (C-2), 54.4 (CH₃O-1), 57.1 (CH₃O-3), 66.9 (C-5), 74.9 (CH₂O-4), 78.9 (C-3), 84.1 (C-4), 98.2 (C-1), 127.5–138.7 (Ar).

9 was also prepared by methylation of 14 in THF using NaH-MeI (85%). The resulting syrup showed the same physical data as described above.

Methyl 3,4-di-O-benzyl-2,6-dideoxy-α-D-arabino-hexopyranoside (10). The xanthogenate 8 (770 mg) was treated as described in general procedure 2. After chromatography (solvent D) 385 mg (66%) of 10 was obtained as a chromatographically homogeneous (R_f =0.65, D) syrup with [α]_D + 82.1° (c 1.5). ¹H NMR: δ 1.32 (d, 3 H, J 6.2 Hz, CH₃), 1.69 (ddd, 1 H, J 13.0, 11.4, 3.7 Hz, H-2_{ax}), 2.32 (ddd, 1 H, J 13.0, 5.0, 1.3 Hz, H-2_{eq}), 3.16 (t, 1 H, J 9.6 Hz, H-4), 3.33 (s, 3 H, CH₃O-1), 3.72 (dq, 1 H, J 9.6, 6.2 Hz, H-5), 3.95 (ddd, 1 H, J 11.4, 9.6, 5.0 Hz, H-3), 4.70–4.97 (m, 5 H), 7.34 (m, 10 H, ArH). ¹³C NMR: δ 18.7 (C-6), 36.0 (C-2), 54.5 (CH₃O-1), 67.1 (C-5), 71.7 and 75.7 (2 × CH₂O), 77.4 (C-3), 84.2 (C-4), 98.0 (C-1), 127.0–138.1 (Ar).

Methyl 2,6-dideoxy-α-D-arabino-hexopyranoside (11). A mixture of 10 (101 mg) and 10% Pd–C (20 mg) in 5 ml of ethyl acetate was stirred under a hydrogen atmosphere for 24 h. Filtration over a short silica gel column (ethyl acetate as the eluant) and evaporation of solvents gave 11 (43 mg, 90%) as a colorless syrup (R_f = 0.28, EtOAc), [α]_D + 131° (c 0.45) {Lit.³c [α]_D + 120° (c 2.0, H₂O)}. ¹H NMR data were in agreement with those given.³c ¹³C NMR: δ 18.4 (C-6), 38.2 (C-2), 54.9 (CH₃O-1), 67.5 and 69.3 (C-3 and C-5), 78.0 (C-4), 98.2 (C-1).

Methyl 2,6-dideoxy-3-O-methyl-α-D-arabino-hexopyranoside (12). A mixture of 9 (335 mg) and 10% Pd-C (50 mg) in 10 ml of ethyl acetate was stirred under a hydrogen atmosphere for 36 h. After filtration over a short silica gel column (eluant D) the solvents were evaporated off to give 198 mg (90%) of 12 as a chromatographically homogeneous syrup ($R_f = 0.28$, D), $[\alpha]_D + 1.01^\circ$ (c 1.6) {Lit.^{4a} $[\alpha]_D + 104.2^\circ$ (c 0.57)}. Spectroscopic data were in in full agreement with those reported.^{4a}

Methyl 4-O-benzyl-2,6-dideoxy-α-D-arabino-hexopyranoside (14) and methyl 4-O-benzyl-3,6-dideoxy-α-D-arabinohexopyranoside (15). A solution of 13¹⁰ (170 mg) in 10 ml toluene under a nitrogen atmosphere was heated under reflux. A mixture of AIBN (10 mg) and tributyltin hydride (480 mg) in 10 ml of toluene was added over a period of 45 min (the addition was repeated with half the amounts after 2 h). When all the starting material had reacted (TLC), the mixture was evaporated in vacuo and the remainder was submitted to flash chromatography (solvent D) to yield 57 mg (41%) of **14** ($R_f = 0.27$, D) and 42 mg (30%) of 15 ($R_f = 0.21$, D). Physical data for compound 14: $[\alpha]_D + 92.2^{\circ}$ (c 1.0). ¹H NMR: δ 1.34 (d, 3 H, J 6.4 Hz, CH₃), 1.68 (ddd, 1 H, J 12.8, 11.4, 1.4 Hz, H-2_{ax}), 2.15 (ddd, 1 H, J 12.8, 5.2, 1.2 Hz, H-2_{eq}), 2.99 (t, 1 H, J 9.2 Hz, H-4), 3.30 (s, 3 H, CH₃O-1), 3.70 (dq, 1 H, J 9.2, 6.4 Hz, H-5), 3.99 (ddd, 1 H, J 11.4, 9.2, 5.2 Hz, H-3), 4.74 (br s, 1 H, H-1), 4.75 (dd, 2 H, CH₂O-4), 7.34 (m, 5 H, ArH). ¹³C NMR: δ 18.3 (C-6), 37.6 (C-2), 54.6 (CH₃O-1), 66.8 and 68.7 (C-3, C-5), 75.0 (CH₂O-4), 86.4 (C-4), 98.2 (C-1), 127.6–138.2 (Ar).

Methyl 6-deoxy-4-O-methyl- α -D-talopyranoside (16). To a mixture of 2 (0.74 g), KOH (0.76 g) and DMSO (15 ml) was added MeI (1.44 g) dropwise with stirring at 10°C. Stirring was continued at room temperature for 1 h, the mixture was diluted with CH₂Cl₂ (15 ml) and washed with water $(5 \times 15 \text{ ml})$. The organic phase was dried with MgSO₄, filtered and evaporated under reduced pressure to yield the 4-O-methyl ether as a homogeneous syrup $(R_{\rm f} = 0.55, \, {\rm D})$. The crude syrup was dissolved in 50 ml of dry chloroform, and 90% aqueous trifluoroacetic acid (TFA) was added (1 ml). After 1 h the mixture was evaporated in vacuo and co-distilled with water and toluene. The residue was submitted to flash chromatography (eluant A) on SiO₂ to yield 16 (0.39 g, 67%) as colorless needles (from B), m.p. $76-77^{\circ}$ C; $[\alpha]_{D} + 119.2^{\circ}$ (c 2.8). ¹H NMR: δ 1.21 (d, 3 H, J 6.6 Hz, CH₃), 3.25 (s, 3 H, CH_3O-1), 3.33 (br s, 2 H, $2 \times OH$), 3.51 (s, 3 H, CH₃O-4), 3.55 (m, 2 H), 3.68 (t, 1 H, J 3.2 Hz, H-3), 3.75 (q, 1 H, J 6.6 Hz, H-5), 4.63 (s, 1 H, H-1). ¹³C NMR: δ 16.7 (C-6), 54.9 (CH₃O-1), 62.5 (CH₃O-4), 65.7, 66.6 and 70.7 (C-2, C-3 and C-5), 83.4 (C-4), 101.9 (C-1).

Methyl 4-O-benzyl-6-deoxy-α-D-talopyranoside (17). To a mixture of 2 (0.83 g) and KOH (0.85 g) in 20 ml DMSO was added benzyl bromide (1.30 g) dropwise. After 2 h methanol (2 ml) was added to destroy the excess of reagent and after an additional 30 min CH_2Cl_2 (60 ml)

and water (20 ml) were added. The organic layer was separated and washed with water (5 × 20 ml). After being dried with MgSO₄ the mixture was co-evaporated several times with water and finally toluene. The syrupy residue obtained could be used directly and was treated as described for 16. The remainder was purified by flash chromatography on silica gel to yield 17 (0.89 g, 87 %) as a syrup (R_f = 0.56, A) with [α]_D + 98.8° (c 1.8). ¹H NMR: δ 1.26 (d, 3 H, J 6.6 Hz, CH₃), 3.01 (d, 1 H, J 10.4 Hz, OH), 3.35 (s, 3 H, CH₃O-1), 3.43 (d, 1 H, J 12.0 Hz, OH), 3.62 (m, 2 H), 3.84 (m, 2 H), 4.73 (dd, 2 H, CH₂O-4), 4.74 (s, 1 H, H-1), 7.35 (m. 5 H, ArH).

Methyl 3,4-di-O-benzyl-6-deoxy- α -D-talopyranoside (18) and methyl 2,4-di-O-benzyl-6-deoxy-α-D-talopyranoside (19). To a solution of 17 (210 mg) in 10 ml toluene was added bis(tributyltin) oxide (467 mg) and the mixture was refluxed with azeotropic removal of water for 16 h. The solution was concentrated to about 5 ml, benzyl bromide (333 mg) and tetrabutylammonium bromide (288 mg) were added and the mixture was kept at 90°C for 5 h. The solvent was removed and the residue submitted to column chromatography (eluant D) to yield 18 (151 mg, 55%) as a syrup $(R_f = 0.46, D)$, $[\alpha]_D + 47.3^{\circ}(c \ 0.85)$. ¹H NMR: $\delta \ 1.24$ (d, 3 H, $J \ 6.6$ Hz, CH₃), 3.35 (s, 3 H, CH₃O-1), 3.68 (br s, 1 H), 3.74 (t, 1 H, J 2.8 Hz, H-3), 3.83 (q, 1 H, J 6.6 Hz, H-5), 3.98 (br s, 1 H), 4.22 (br s, 1 H), 4.74 and 4.79 ($2 \times dd$, 4 H, $2 \times CH_2O$), 4.80 (br s, 1 H, H-1), 7.35 (m, 10 H, ArH). ¹³C NMR: δ 17.4 (C-6), 55.2 (CH₃O-1), 66.4 (C-5), 67.9 (C-2), 69.8 (CH₂O), 74.4 and 75.5 (C-3 and CH₂O), 78.8 (C-4), 102.5 (C-1), 127.0–137.6 (Ar). Further elution gave **19** (98 mg, 36 %) as a syrup ($R_f = 0.36$, D).

Methyl 6-deoxy-4-O-methyl-2,3-O-thiocarbonyl-α-D-talopyranoside (20). A solution of 16 (500 mg) and TCDI (580 mg) in 20 ml of dry THF was refluxed under a nitrogen atmosphere for 2 h. The solvent was removed and the residue purified by flash chromatography on silica gel (eluant A) to yield a syrup ($R_{\rm f}$ = 0.68, A) which crystallized on treatment with diisopropyl ether to give 444 mg (73%) of 20, m.p. 138–140°C; [α]_D + 34.7° (c 0.5). ¹H NMR: δ 1.35 (d, 3 H, J 6.4 Hz, CH₃), 3.35 (dd, 1 H, J 5.0, 1.0 Hz, H-4), 3.40 (s, 3 H, CH₃O-1), 3.58 (s, 3 H, CH₃O-4), 3.84 (dq, 1 H, J 6.4, 1.0 Hz, H-5), 4.56 (dd, 1 H, J 7.1, 1.0 Hz, H-2), 5.04 (t, 1 H, J 7.0 Hz, H-3), 5.05 (br s, 1 H, H-1). ¹³C NMR: δ 16.2 (C-6), 55.4 (CH₃O-1), 62.4 and 63.4 (CH₃O-4 and C-5), 75.1, 77.7 and 78.0 (C-2, C-3 and C-4), 95.7 (C-1), 189.6 (C=S).

Methyl 2,6-dideoxy-4-O-methyl-α-D-lyxo-hexopyranoside (21). A mixture of 30 (120 mg) and 10 % Pd-on-charcoal (20 mg) in 5 ml of ethyl acetate was stirred under a hydrogen atmosphere for 24 h. Filtration of the reaction mixture through a short silica gel column and evaporation of the solvent afforded 21 (70 mg, 88 %) which crystallized on standing, m.p. 97–98°C; $[\alpha]_D + 163^\circ$ (c 1.0) {Lit. 13b m.p. 95–96°C; $[\alpha]_D + 168^\circ$ (c 0.6)}. H NMR

data were in accordance with those given in the literature. ^{13b} C NMR: δ 17.8 (C-6), 34.4 (C-2), 55.0 (CH₃O-1), 62.3 (CH₃O-4), 66.1, 66.2 (C-3, C-5), 81.5 (C-4), 98.4 (C-1).

21 was also prepared (37%) by treating 20 with Bu₃SnH as described for the synthesis of 14, to give material showing the same physical data as given above.

Methyl 6-deoxy-3,4-O-isopropylidene-α-D-talopyranoside (23) and methyl 6-deoxy-2-O-(6-deoxy-3,4-O-isopropylidene-α-D-talopyranosyl)-3,4-O-isopropylidene-α-D-talopyranoside (24). To a well-stirred solution of 2 (1.64 g) in 50 ml dry acetone was added dropwise a solution of 0.12 ml sulfuric acid in 2.5 ml acetone. After 1 h dry Na₂CO₃ was added, the solution was filtered and evaporated in vacuo and the residue was submitted to column chromatography on silica gel (eluant A). Three compounds were isolated: 135 mg (8%) of 2 ($R_f = 0.56$, A), 1.21 g (74%) of 23 as a chromatographically homogeneous syrup $(R_f = 0.32, A), [\alpha]_D + 80.0^{\circ} (c 1.6).$ ¹H NMR: δ 1.24 (d, 3 H, J 6.6 Hz, CH₃), 1.35 and 1.51 $[2 \text{ s}, 6 \text{ H}, C(CH_3)_2], 2.60 \text{ (br s}, 1 \text{ H}, OH), 3.42 \text{ (s}, 3 \text{ H},$ CH₃O-1), 3.67 (dd, 1 H, J 5.6, 3.4 Hz, H-2), 3.78 (dq, 1 H, J 6.6, 1.6 Hz, H-5), 4.09 (dd, 1 H, J 7.4, 1.6 Hz, H-4), 4.49 (dd, 1 H, J 7.4, 3.4 Hz, H-3), 4.63 (d, 1 H, J 5.6 Hz, H-1). 13 C NMR: δ 16.4 (C-6), 25.7 and 26.6 [C(CH₃)₂], 55.5 (CH₃O-1), 65.2 (C-5), 68.9 (C-2), 73.7 (C-3), 76.3 (C-4), 101.4 (C-1), 109.5 $[C(CH_3)_2]$. Further elution yielded the disaccharide 24 (105 mg, 6%) as a syrup $(R_{\rm f}=0.10,\,{\rm A})$. The isopropylidene derivative 23 was also synthesized (60% yield), by treating methyl 6-deoxyα-D-talopyranoside with 2,2-dimethoxypropane and pyridinium p-toluenesulfonate (PPTS) in acetone as the solvent, showing the same physical data as given above. In addition, compound 2 was obtained in a 21 % yield.

Methyl 6-deoxy-3,4-O-isopropylidene-2-O-[(methylthio)-thiocarbonyl]-α-D-talopyranoside (25). The alcohol 23 (1.18 g) was treated according to general procedure 1, to yield 1.41 g (88%) of 25 as a syrup (R_f =0.60, E) after flash chromatography (eluant E), [α]_D + 105.4° (c 1.1). ¹H NMR: δ 1.26 (d, 3 H, J 6.6 Hz, CH₃), 1.33 and 1.53 [2 s, 6 H, C(CH₃)₂], 2.60 (s, 3 H, SCH₃), 3.41 (s, 3 H, CH₃O-1), 3.89 (dq, 1 H, J 6.6, 1.8 Hz, H-5), 4.17 (dd, 1 H, J 7.4, 1.8 Hz, H-4), 4.75 (dd, 1 H, J 7.4, 3.0 Hz, H-3), 4.92 (d, 1 H, J 6.2 Hz, H-1), 5.79 (dd, 1 H, J 6.2, 3.0 Hz, H-2). ¹³C NMR: δ 16.2 (C-6), 19.9 (SCH₃), 25.9 and 26.7 [C(CH₃)₂], 55.5 (CH₃O-1), 66.1 (C-5), 71.7 (C-2), 76.5 (C-3), 79.0 (C-4), 98.3 (C-1), 110.2 [C(CH₃)₂], 215.3 (C=S).

Methyl 2,6-dideoxy-3,4-O-isopropylidene-α-D-talopyranoside (26). A solution of 25 (1.12 g) in 60 ml toluene was processed according to general procedure 2. After 2 h the mixture was evaporated in vacuo and the residue was submitted to column chromatography (eluant F) to give 0.58 g (79%) of 26 as a colorless syrup (R_f =0.44, F), $[\alpha]_D$ +61.0° (c 1.55) {Lit. $[\alpha]_D$ +59.2° (c 0.9, CH₂Cl₂)}. ¹H NMR data were in accordance with those given in the literature. ¹⁶ ¹³C NMR: δ 16.7 (C-6), 25.8 and 27.3 [C(CH₃)₂], 30.9 (C-2), 55.0 (CH₃O-1), 60.5 (C-5), 70.6 (C-3), 75.6 (C-4), 97.2 (C-1), 108.3 [C(CH₃)₂].

Methyl 2,6-dideoxy-α-D-lyxo-hexopyranoside (27). To a solution of 26 (0.54 g) in 50 ml of dry MeOH was added 2 ml methanolic HCl and the solution was left at $+4^{\circ}$ C for 16 h. The solution was neutralized with ion-exchange resin (Ionenaustauscher III, Merck), filtered and evaporated in vacuo, and the residue was purified by flash chromatography on silica gel with ether as the eluant to yield 27 (0.36 g, 81 %) as a syrup, $[\alpha]_D + 118^{\circ}$ (c 0.6) {Lit. $[\alpha]_D + 125.2^{\circ}$ (c 0.8, CH₂Cl₂)}. $[\alpha]_D + 125.2^{\circ}$ (c 0.8, CH₂Cl₂)}.

Methyl 4-O-acetyl-2,6-dideoxy-α-D-lyxo-hexopyranoside (28). A mixture of 31 (205 mg) and 10 % Pd-on-charcoal (40 mg) in 10 ml ethyl acetate was stirred under a hydrogen atmosphere for 24 h. The mixture was filtered and purified by chromatography (ethyl acetate as the eluant) to yield 121 mg (86%) of 28 as a colorless syrup (R_f = 0.20, EtOAc) which crystallized on treatment with eluant B, m.p. 87–89°C; [α]_D + 130.1° (c 0.85) {Lit. ^{17a} m.p. 89°C; [α]_D + 129° (c 0.5)}. ¹³C NMR: δ 17.4 (C-6), 21.5 (OAc), 33.3 (C-2), 55.2 (CH₃O-1), 64.6 and 65.0 (C-3, C-5), 73.0 (C-4), 98.5 (C-1), 170.9 (C=O). ¹H NMR data were in accordance with those reported. ¹⁶

Methyl 3-O-benzyl-2,6-dideoxy-α-D-lyxo-hexopyranoside (29). A mixture of 27 (0.48 g) and dibutyltin oxide (0.73 g) in 40 ml of toluene was refluxed with azeotropic removal of water for 16 h. The mixture was cooled and concentrated to about 20 ml. Benzyl bromide (1.26 g) and tetrabutylammonium iodide (1.09 g) were added and the mixture was kept at 90°C for 2 h. The solvent was removed and the residue purified by column chromatography (eluant B) to yield 29 (0.62 g, 83 %) as a colorless syrup $(R_f = 0.45, B), [\alpha]_D + 115^{\circ} (c 0.72)$ {Lit. 19 [α]_D + 83° (c 2.5)}. 1 H NMR: δ 1.32 (d, 3 H, J 6.6 Hz, CH₃), 1.95 (m, 2 H), 2.12 (br s, 1 H, OH), 3.32 (s, 3 H, CH₃O-1), 3.84 (m, 3 H), 4.59 (s, 2 H, CH₂O-3), 4.82 (d, 1 H, J 3.2 Hz, H-1), 7.34 (m, 5 H, ArH). ¹³C NMR: δ 17.4 (C-6), 30.4 (C-2), 55.1 (CH₃O-1), 65.5 (C-5), 68.8, 70.1 (C-4, CH₂O-3), 73.0 (C-3), 98.4 (C-1), 127.0–137.3 (Ar).

Methyl 3-O-benzyl-2,6-dideoxy-4-O-methyl-α-D-lyxo-hexopyranoside (30). The benzyl ether 29 (218 mg) was methylated with NaH–MeI in THF in the usual manner to yield 183 mg (79%) of 30 as a syrup (R_t = 0.55, C) after chromatography on silica gel (eluant C), [α]_D + 114.6° (c 0.5) {Lit. 17a [α]_D + 108° (c 0.58)}. 1H NMR: δ 1.27 (d, 3 H, J 6.4 Hz, CH₃), 1.93 (ddd, 1 H, J 12.6, 5.0, 1.2 Hz, H-2_{eq}), 2.08 (ddd, 1 H, J 12.6, 11.8, 3.6 Hz, H-2_{ax}), 3.30 (s, 3 H, CH₃O-1), 3.34 (d, 1 H, J 2.4 Hz, H-4), 3.63

(s, 3 H, CH₃O-4), 3.79 (q, 1 H, J 6.4 Hz, H-5), 3.85 (ddd, 1 H, J 11.8, 5.0, 2.4 Hz, H-3), 4.61 (s, 2 H, CH₂O-3), 4.82 (d, 1 H, J 3.6 Hz, H-1), 7.34 (m, 5 H, ArH). ¹³C NMR: δ 17.7 (C-6), 31.1 (C-2), 55.0 (CH₃O-1), 61.5 (CH₃O-4), 66.5 (C-5), 70.4 and 74.9 (C-3 and CH₂O-3), 78.7 (C-4), 98.7 (C-1), 126.7–137.9 (Ar).

Methyl 4-O-acetyl-3-O-benzyl-2,6-dideoxy-α-D-lyxo-hexopyranoside (31). The benzyl ether 29 (160 mg) was acetylated with acetic anhydride–triethylamine–DMAP in CH₂Cl₂ to yield 160 mg (86%) of 31, $[α]_D + 160.2^\circ$ (c 0.5) {Lit. 17a $[α]_D + 168^\circ$ (c 0.38)}. 1 H NMR: δ 1.18 (d, 3 H, J 6.6 Hz, CH₃), 1.92 (dd, 2 H, J 12.5, 5.6 Hz, H-2_{eq}), 2.02 (ddd, 1 H, J 12.5, 11.6, 3.8 Hz, H-2_{ax}), 2.18 (s, 3 H, OAc), 3.33 (s, 3 H, CH₃O-1), 3.90 (ddd, 1 H, J 11.6, 5.6, 2.6 Hz, H-3), 3.94 (q, 1 H, J 3.8 Hz, H-5), 4.55 (dd, 2 H, CH₂O-3), 4.85 (d, 1 H, J 3.8 Hz, H-1), 5.34 (d, 1 H, J 2.6 Hz, H-4), 7.31 (m, 5 H, ArH). 13 C NMR: δ 17.5 (C-1), 21.8 (OAc), 31.8 (C-2), 55.1 (CH₃O-1), 64.8 (C-5), 69.1, 70.3 and 71.4 (C-3, C-4 and CH₂O-4), 98.6 (C-6), 127.0–137.4 (Ar), 169.8 (C=O).

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References

- For a review on deoxy sugars, see Hanessian, S. and Haskell, T. H. In: Pigman, W. and Horton, D., Eds., The Carbohydrates, 2nd ed., Academic Press, New York 1970, Vol. II A, pp. 139-212.
- (a) Mallams, A. K. In: Kennedy, J. F., Ed., Carbohydrate Chemistry, Clarendon Press, Oxford 1988, pp. 73-133;
 (b) Berdy, J. CRC handbook of Antibiotic Compounds, CRC Press, Boca Raton 1980, Vol. 1-3.
- (a) Gilleron, M., Fournie, J. J., Pougny, J. R. and Puzo, G. J. Carbohydr. Chem. 7 (1988) 733; (b) Durette, P. L. Synthesis (1980) 1037; (c) Stanek J., Jr., Marek, M. and Jary, J. Carbohydr. Res. 64 (1978) 315.
- (a) Dornhagen, J., Klausener, A., Runsink, J. and Scharf, H.-D. *Liebigs Ann. Chem.* (1985) 1838 and literature cited therein; (b) Yasuda, S. and Matsumoto, T. *Tetrahedron Lett.* (1969) 4393.
- Kjølberg, O. and Neumann, K. Acta Chem. Scand. 46 (1992) 877.
- For a review, see David, S. and Hanessian, S. Tetrahedron 41 (1985) 643.
- Czernecki, S., Georgoulis, C. and Provelenghiou, C. Tetrahedron Lett. (1976) 3535.
- 8. For a review on deoxygenations based on the thiocarbonyl group, see Barton, D. H. R. Tetrahedron 48 (1992) 2529.
- Haque, M. E., Kikuchi, T., Kanemitsu, K. and Tsuda, Y. Chem. Pharm. Bull. 35 (1987) 1016.
- 10. Neumann, K. Acta Chem. Scand. 46 (1992) 913.
- 11. Eis, M. J. and Ganem, B. Carbohydr. Res. 176 (1988) 316.
- 12. Aspinall, G. O. and Takeo, K. *Carbohydr. Res. 121* (1983)
- (a) Yoshimura, J., Aqeel, A., Hong, N., Sato, K.-I. and Hashimoto, H. Carbohydr. Res. 155 (1986) 236; (b) Jary, J. and Marek, M. Collect. Czech. Chem. Commun. 46 (1981) 2410 and references cited therein; (c) Brimacombe, J. S. and Portsmouth, D. Carbohydr. Res. 1 (1965) 128.

- Shashkov, A. S., Yashunskij, D. V., Zhdanovich, Y. V. and Lokshin, G. B. Bioorg. Khim. 17 (1991) 410; Chem. Abstr. 115 (1991) 72059s.
- 15. For an example, see Hughes, N. A. Carbohydr. Res. 7 (1968)
- 16. Thiem, J. and Meyer, B. Chem. Ber. 113 (1980) 3058 and literature cited therein.
- 17. (a) Roush, W. R. and Straub, J. A. Tetrahedron Lett. (1986) 3349; (b) for the assignment of the correct structure, see
- Thiem, J. and Meyer, B. J. Chem. Soc., Perkin Trans. 2 (1979) 1331.
- 18. Fang, Y., Kong, F. and Wang, Q. J. Carbohydr. Chem. 6 (1987) 169.
- Brimacombe, J. S., Portsmouth, D. and Stacey, M. J. Chem. Soc. (1964) 5614.

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