Conformational Properties of the Amino Acid Residues L-Cysteine, L-Serine and L-Cystine

C. H. Görbitz

Department of Chemistry, University of Oslo, P.O. Box 1033, Blindern N-0315 Oslo 3, Norway

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The correlation between torsion angles of the main chain and the orientation of the side chain in the solid phase has been investigated for the amino acids L-cysteine, L-serine and L-cystine. Information has been obtained from the Cambridge Structural Database.

In the crystal structures of cysteine and serine the torsion angle N-C^a-C'-N (ψ) is often close to 0 or 180°. When this is the case, N-C^a-C'-O'/S' (χ^1) is almost exclusively in the vicinity of 60° (g^+). When ψ is in the interval 30-150°, only trans (t) or gauche⁻ (g^-) is observed. Thus, χ^1 is strongly dependent on the conformation of the main chain for these residues. Force-field energy calculations agree reasonably well with the experimental results.

For L-cystine, g^- is more frequently observed than g^+ at χ^1 . χ^2 and χ^3 (disulfide bond) are both around $+/-85^\circ$. The sign sequences for the five torsion angles along the disulfide bridge $(\chi^1, \chi^2, \chi^3, \chi^{2'}, \chi^{1'})$ are: cyclic peptides, --+- or other; non-cyclic with twofold axis, +--+ or +++++; other non-cyclic, ---- or +++++

The connection between the values of the torsion angles φ and ψ for various secondary structures of a peptide chain, usually presented in the form of a Ramachandran map, is familiar to all peptide chemists. In contrast, the less obvious connection between main-chain conformation and allowed or preferred orientations of the side chain is only rarely commented upon. Most papers present the two accompanying sets of torsion angles as largely independent entities. There are, however, some theoretical studies¹⁻³ which indicate that the presence of atoms beyond C⁶ in the side chain does have an effect on the allowed values of ϕ and ψ. Conformational maps have been produced showing these restrictions. Furthermore, a survey of crystal structures of peptides⁴ has demonstrated that χ¹ at 60° is disfavored when ψ deviates much from 0 or 180° and that χ^1 at 180° is less frequently observed when ψ is close to these angle values. Previous investigations^{5,6} have also indicated that in general the distribution between the three χ^1 rotamers is slightly different in peptides and proteins. The purpose of this work was to investigate the existence of a connection between side-chain and main-chain conformations for the amino acids L-cysteine and L-serine. Conformational properties of the disulfide bridge in L-cystine have also been analyzed.

Methodology

All crystallographic data were retrieved from the Cambridge Structural Database (CSD, January 1989 release).⁷ The number of structures analyzed is 20 for serine,⁸⁻²² containing 26 crystallographically independent residues, 8 for

cysteine, ²³⁻³⁰ containing 9 residues and 14 for cystine ³¹⁻⁴³ each containing one disulfide bridge, four of which have a twofold axis through the S-S bond. Structures in the first group all have R factors less than 0.085, whereas for cysteine three structures have R factors in the range 0.09-0.14. Some of the cystine structures have R factors up to 0.13, but all have been retained due to the small number of molecules available. No side-chain disorder was allowed. In cases of duplicate structures, the more precise determination was used. Data for cysteine has been supplemented with two structures, ⁴⁴ and torsion angles for three D-serine

Table 1. References for REFCODes.

REFCOD	Ref.	REFCOD	Ref.	REFCOD	Ref.	
Serine		PIVSMA	18	Cystine		
		SERASC10	19	•		
BUFFEN10	8	SERGLY	20	BEWDIQ10	31	
BUFFIR10	8	SERTYR10	21	BICKON	32	
CGPSAQ	9	FEPFOV	22	CYSMEC	33	
COBXAS	8			CYSTBR	34	
COBXEW	8	Cysteine		CYSTCL02	35	
CSEHSM	10	-		CYSTIN10	36	
CUJHIY	11	CYGNAI	23	DGLYCH01	37	
CUJHOE	11	CYSCLM10	24	DUNLON	38	
CYSESE	12	DLCYST	25	GAGDIB	39	
DIYZOA	13	ECYSCU	26	LCSTIM	40	
DLSERN11	14	GLUTAS02	27	LCYSCC	41	
FERCRN10	15	LCYSTN	28	LCYSTI10	42	
LALLSE	16	LCYSTN21	29	LCYSTI11	43	
LSERIN10	17	NALCYS10	30	TANDEM30	32	
LSERMH10	14					

residues were inverted to the equivalent conformation of L-serine. Cystine structures include molecules where the residue is part of a large ring system, but not cyclo-cystine (Cys-Cys). The nomenclature and conventions used follow the recommendations of IUPAC.⁴⁵ When referring to a CSD entry, the database identification reference code (REFCOD) was used. REFCOD references are given in Table 1.

Energy minimizations were performed with the AMBER force field^{46,47} which is an integrated part of the SYBYL Version 5.2 computer program. ⁴⁸ Electrostatic interactions were used and all hydrogen atoms were included. The dielectric constant was set to 80 which should be appropriate for mimicking an aqueous molecular environment. The cut-off criteria was an energy change less than 0.00025 kJ mol⁻¹. Conformational searches were carried out with the SEARCH function of the SYBYL program. Relatively short intramolecular contacts were permitted, down to 0.93 times the sum of the van der Waals' radii. Instead of the default geometry of the *trans* peptide bond, new parameters were calculated using 264 peptide bonds in 116 structures of *R* factor < 0.075 from the CSD. A list of references is available from the author on request.

Description and discussion

Serine and cysteine

 χ^1 and χ^2 . For the study of cysteine, only unsubstituted residues with a free –SH group were used. The distribution

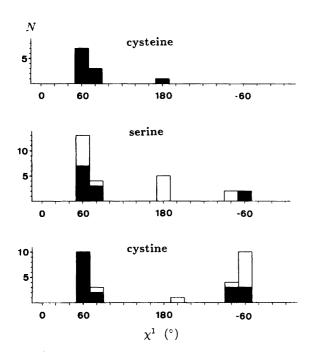
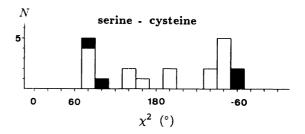
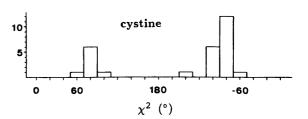


Fig. 1. $χ^1$ distribution for cysteine, serine and cystine. The white and black columns represent residues in cyclic and non-cyclic molecules, respectively. In the diagram for cystine, molecules with a twofold axis are given double weight.





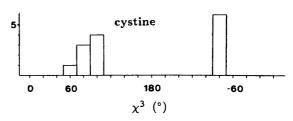


Fig. 2. χ^2 distributions for cysteine (black)/serine (top) and cystine (middle) and χ^3 distribution for cystine (bottom).

of χ^1 (Fig. 1) shows a highly (>99.9%) significant preference for the g^+ conformation (91%) which has not been described in earlier surveys. For serine the distribution also shows a preference for g^+ (65%), and upon removal of all residues which are part of cyclic systems, this preference is even more pronounced (83%). The common distribution of χ^2 for cysteine and serine is given in Fig. 2. Peaks are found around $+/-80^\circ$ and $+/-150^\circ$. It should be noted that the positions of hydrogen atoms may be subject to relatively large errors, and that the hydrogen bonding for serine, and to a lesser extent for cysteine, may conceal any inherent preferences of the terminal groups towards particular torsion angles.

Correlation with main-chain. Fig. 3 depicts the distribution of ψ against χ^1 for serine and cysteine. When the N-C^a-C'-N group is close to planar (ψ close to 0° or 180°), the g^+ conformation is totally dominant. On the other hand, g^+ is not observed for residues with ψ in the interval 100–150°. The Ramachandran plot of (φ, ψ) for 23 residues is shown in Fig. 4. It can be seen that all but two structures fit nicely into four well-defined groups. These may be described in more detail.

Group (A). ψ is close to 180° and φ to -164° (one residue at -136°). This group consists of three serine residues in large ring systems (2 · CUJHIY, CUJHOE), as well as two C-terminal serines (DIYZOA, LALLSE) and one

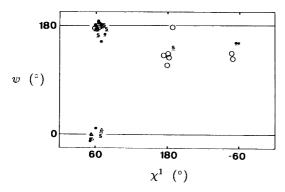


Fig. 3. Distribution of ψ against χ^1 for cysteine and serine. In Figs. 3 and 4 the following symbols are used: Δ , Serine in cyclic dipeptide; \bigcirc , Cyclic structure with serine; \blacksquare , Linear structure with serine; S, Cysteine in linear structure. Underlined symbols represent structures with *C*-terminal carboxy, carboxylate or ester groups. For these molecules ψ_T^{-1} or ψ_T^{-2} is plotted. In Fig. 3, the value closest to 180° has been used, in Fig. 4 the value giving the better fit to one of the four groups indicated in the map.

C-terminal cysteine (NALCYS10). The conformation is very extended; ψ -values close to 180° are in general quite rare. The χ^1 distribution g^+ :t: g^- is 5:1:0.

Group (B). ψ is around 0°, and ϕ is around -80° . This is in the 'bridge region' connecting the regions for extended chain and helix in the Ramachandran plot. The parameters correspond to a type I β turn, but the group consists of three C-terminal serine amides (BUFFEN10, BUFFIR10, COBXAS), one C-terminal serine methyl ester (COBXEW) and glutathione (γ -Glu-Cys-Gly, GLUTAS02), none of which are involved in such turns. All χ^1 are g^+ .

Group (C). (φ, ψ) is around (-55°, 130°) which are the parameters for the second residue in the type II β turn. All serines are located in such β turns in rather similar large

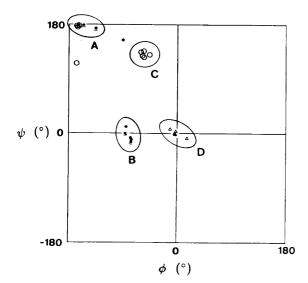


Fig. 4. Ramachandran plot of (ϕ,ψ) for cysteine and serine. Letters refer to groups described in the text.

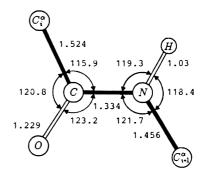


Fig. 5. Bond lengths (Å) and bond angles (°) of the *trans* peptide bond. Standard deviations for mean values are 0.001 Å and 0.1°, respectively. For bond angles involving H the standard deviations are 0.3°. The N–H value is derived from neutron diffraction studies, X-ray structures give 0.934(6) Å for this bond length.

ring structures (2 · CUJHIY, CUJHOE, 2 · FERCRN10). The χ^1 distribution $g^+:t:g^-$ is 0:3:2.

Group (D). Cyclic dipeptides (dioxopiperazines), in which both φ and ψ necessarily are close to 0° (CSEHSM, $2 \cdot \text{CYSESE}$, SERTYR10, FEPFOV). Thus the connection between these two torsion angles in this group is of limited interest in this study, but it is still interesting that χ^1 is invariably found in the g^+ conformation. This group is not discussed further.

Computation of sterically allowed conformations. Earlier papers $^{1-3}$ have documented that the presence of a C $^{\gamma}$ -atom leads to slightly different allowed regions of the Ramachandran map depending on which staggered position is occupied. To see if the χ^1 populations described above could be

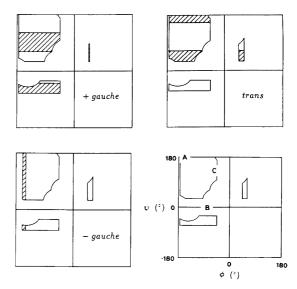


Fig. 6. Theoretical Ramachandran plots for cysteine and serine. Solid lines enclose allowed values of ϕ and ψ for each χ^1 rotamer of serine, lined regions are disallowed for cysteine.

$$C_{i}^{\beta}$$
 C_{i}^{β}
 C_{i}^{β}
 C_{i+1}^{β}
 C_{i+1}^{β}
 C_{i+1}^{β}
 C_{i+1}^{β}
 C_{i+1}^{β}
 C_{i+1}^{β}

Scheme 1.

explained by such differences, similar maps were calculated for serine and cysteine. The molecules N-acetyl-L-Cys-N-methylamide and N-acetyl-L-Ser-N-methylamide were used as model peptides. The geometry of the trans peptide bond from a survey of crystal structures is given in Fig. 5. The values indicated were used during the search procedure except for the two angles involving the amide proton which were raised by 0.3° (to 19.6 and 18.7°) to obtain a planar arrangement around the nitrogen atom. ω was 180° and $\tau(C^{\alpha})$ (N- C^{α} -C') was kept at 111.0° . The results are shown in Fig. 6.

The maps for serine show that the differences between permitted regions for the three χ^1 rotamers are marginal, and in most cases these can be neglected. The Ramachandran plots for cysteine however, demonstrate that the larger van der Waals' radius of a sulfur atom compared with an oxygen atom greatly reduces the rotational freedom of the side chain for this residue. In the experimental (A) conformation, g^+ is the only allowed rotamer for cysteine,

 g^- and t give too close contacts with O_{i-1} and H_{i+1} , respectively. On the other hand, for serine all rotamers are allowed. (B) is apparently disallowed for both residues regardless of the orientation of the side chain. In this region, steric conflict exists between N_i and H_{i+1} (Scheme 1).

This short intramolecular contact may be relieved fairly easily by a small opening of $\tau(C^{\alpha})$. Indeed, experimental results⁴⁹ show that when ψ is close to 0°, the mean value for $\tau(C^{\alpha})$ is about 4° larger than in the extended region (ψ in the interval 120–170°). Making this adjustment, one finds that all rotamers are possible for both residues. This is also the case for (C).

In an attempt to explain further the conformational preferences of χ^1 , the three main-chain configurations have been energy-minimized with the AMBER force field. In the first step ϕ and ψ were kept fixed to experimental values. χ^1 was not fixed, but tended to change very little. In the next step all constraints were released to let the refinement proceed to the closest local minimum. The results are given in Table 2.

The energies obtained from the constrained refinements generally give and adequate agreement with the experimental distributions which may be summarized as follows (1) Serine (A): g^- is not observed and is calculated to have the highest energy. The experimental preference for g^+ over t is unexplained. (2) Serine (B): t has high energy and is unobserved. The preference for g^+ over g^- is unexplained. (3) Serine (C): g^+ is not observed and has the highest energy. t and g^- are both observed, but a discrepancy exists as t is calculated to have significantly higher

Table 2. Torsion angles (°) and calculated energies (kJ mol⁻¹) for minimized conformers of serine and cysteine.

Residue	Main chain	Constrained refinement				Free refinement				
		φ	ψ	χ¹	Ε	φ	ψ	χ¹	Ε	
Serine	(A)	-164	176	60	0.0ª	-169	178	55	3.8	
	` ,	-164	176	180	0.5	-169	-171	-176	0.0	
		-164	176	-60	16.6	-140	163	-57	6.8	
Serine	(B)	-80	-6	60	0.2	-66	-48	44	2.3	
		-80	-6	180	31.4	-57	-44	-175	32.6	
		-80	-6	-60	0.0	-48	-40	-44	0.0	
Serine	(C)	-55	130	60	20.5	-79	166	45	21.3	
		-55	130	180	18.0	-64	136	-174	32.3	
		-55	130	-60	0.0	-71	64	-50	0.0 ^b	
Cysteine	(A)	-164	176	60	0.0	-168	163	69	0.2	
		-164	176	177	9.9	-158	99	172	0.0	
		-164	176	-60	10.1	-156	102	-75	26.2	
Cysteine	(B)	-80	-6	60	0.0	-75	-21	70	1.9	
		-80	-6	180	4.5	-64	-43	180	0.0	
		-80	-6	-60	17.2	-65	-30	-65	21.2	
Cysteine	(C)	-55	130	60	20.5	-67	142	68	13.4	
•	` '	-55	130	180	0.0	-66	129	179	0.0	
		-55	130	-60	24.7	-65	125	-66	22.5	

^aLowest energy staggered position set to E = 0.0. ^bStabilized by an intramolecular hydrogen bond.

energy than g^- . (4) Cysteine: the experimental material is to limited to allow any comparison with the theoretical data.

The results of the free refinement show that considerable alterations take place when the constraints are released. Molecules in the almost planar (B) conformation with $\psi = -6^{\circ}$ ($\phi = -80^{\circ}$) are shifted to positions in a (ϕ , ψ) map close that of an α -helix (-58° , -47°). For (C) too, substantial shifts are observed. A particular case is serine (C) g^{-} which ends up with an intramolecular hydrogen bond between O_{i-1} and N_{i+1} .

In short, it seems that due to the large shifts for ϕ and ψ in the free refinement, the contrained refinement is most appropriate for studies of χ^1 distributions for a set of (ϕ,ψ) values. The considerations above may then explain some of the experimental results, but at the same time several observations remain unresolved. Taking into account crystallattice forces may possibly improve the results, 50 but reliable theoretical rotamer populations are no doubt very hard to obtain. Finally, it should also be born in mind that this investigation is based on a limited number of experimental structures, and that some of the features described in the text may eventually turn out to be statistical coincidences.

Cystine

 χ^{I} . The distribution shown for cystine (Fig. 1) deviates significantly from those of cysteine and serine. The *trans* conformation is still quite rare, but for this residue g^{-} is dominant over g^{+} .

 χ^2 and χ^3 . The distributions of χ^2 and χ^3 (disulfide bond) are depicted in Fig. 2. The *trans* rotamer is absent for these two torsion angles, and the *gauche* rotamers are significantly distorted from $+/-60^{\circ}$ to positions at approximately $+/-85^{\circ}$.

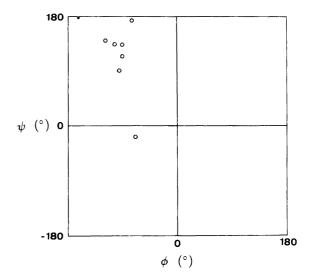


Fig. 7. Ramachandran plot of (ϕ, ψ) for cystine: \bigcirc , cyclic structures; \bigcirc , non-cyclic structure.

Correlation with the main chain. A Ramachandran plot is given in Fig. 7. There are only 8 points in the diagram, of which 7 represent residues in cyclic systems (BEWDIQ10, $2 \cdot BICKON$, DUNLON, GAGDIB, $2 \cdot TANDEM30$). The scatter is quite wide, and the points are not located in the groups described for serine and cysteine. The only non-cyclic structure however (DGLYCH01), fits nicely with the parameters for group (A) with the side chain in the g^+ conformation. There is no indication in this material that the conformational preferences for (φ, ψ, χ^1) and $(\varphi', \psi', \chi^{1'})$ are any different from those of a single cysteine residue. The appearance of the Ramachandran plot is different from the one in Fig. 4 because most of the points represent cyclic systems. Severe restrictions limit the possible conformations of the disulfide bridge in these molecules.

Table 3. Torsion angles (°) in the disulfide bridge in cystine structures with separations between C^{α} atoms (Å).

Compound	NCCS	ccss	CSSC	SSCC	SCCN	Signs	2*	O ^b	C _α ····C _α	REFCOD
Boc-Cys-Pro-Aib-Cys-NHCH ₃	-169.1	-138.7	81.5	73.3	-71.3	t-++-		*	5.3	BEWDIQ10
Tandem · 14 H ₂ O	-53.9	-87.8	101.7	-84.2	-61.0	+		*	3.8	BICKON
L-Cystine dimethyl ester · 2 HCl · H₂O	-75.9	-79.2	-84.4	-77.4	-51.7				5.8	CYSMEC
L-Cystine · 2 HBr	70.6	84.5	77.6	84.5	70.6	+++++	*		5.0	CYSTBR
L-Cystine · 2 HCl	68.7	-88.5	-82.5	-88.5	68.7	++	*		6.1	CYSTCL02
L-Cystine · 2 HBr · 2 H₂O	-54.9	-82.3	-79.8	-70.0	-85.3				5.7	CYSTIN10
N,N'-Diglycyl-∟-cystine · 2 H ₂ O	60.1	-95.2	-84.4	-95.2	60.1	++	*		6.1	DGLYCH01
Pressinoic acid · H ₂ O	79.6	92.9	95.4	-70.7	-60.9	+++		*	5.0	DUNLON
Boc-Cys-Val-Aib-Ala-Leu-Cys-NHCH ₃	-54.9	-101.1	101.2	-84.1	-63.1	+		*	4.0	GAGDIB
L-Cystine diamide · 2 HCl	65.0	-94.6	-81.4	-94.6	65.0	++	*		6.1	LCSTIM
L-Cystine · 2 HCl · 2 H₂O	-55.8	-80.6	-79.9	-70.8	-85.1				5.7	LCYSCC
L-Cystine (hexagonal form)	55.2	81.6	73.7	81.6	55.2	+++++			5.9	LCYSTI10
L-Cystine (tetragonal form)	64.3	75.1	69.3	66.9	51.2	+++++			5.6	LCYSTI11
Tandem · 12 H ₂ O	-52.0	-99.9	99.7	-79.1	-64.0	+		*	3.9	TANDEM30

^a Molecule with a twofold axis through the S-S bond. ^bCyclic peptides.

Table 4. Allowed sign sequences for $\chi^1\chi^2\chi^3\chi^2'\chi^{1\prime}$ torsion angles in the disulfide bridge of cystine. Signs used for the values are (in °): +, χ^2 , χ^3 , $\chi^{2\prime}$ [70, 100] or χ^1 , χ^1 [50, 70]; -, χ^2 , χ^3 , $\chi^{2\prime}$ [-70, -100] or χ^1 , χ^1 [-50, -70]; t, χ^1 , χ^1 [170, 190].

$\frac{\chi^2\chi^3\chi^{2'}}{+++}$	Total	Sign sequences ^b									
	6	+++++	++++t	++++~	t+++t	t+++-	-+++-				
	6	++	+t	+	tt	t					
 ++-	9	+++	t++	-++							
+-+	6	t+-+t									
+	9	t++	t+t	t+							
+ -+-	6	+									
Sum	42	26									

^aTheoretical number of sign combinations. Symmetry has been included; ++++-=-++++ and so on. ^bReadily accessible conformations.

Disulfide bridge conformation. There are three possible rotamers at χ^1 , and two each at χ^2 and χ^3 giving 42 possible combinations for the bridge $\chi^1\chi^2\chi^3\chi^{2'}\chi^{1'}$ (symmetry included, see below). Some of these are prohibited due to steric hindrance, and only four have been found more than once in crystal structures (Table 3).

In the cyclic systems encompassing 14- and 20-membered rings, conformations of the disulfide bridge which give relatively short C^{α} - C^{α} separations are always preferred. In particular, the sign combination --+-, called the short right-hand hook, occurs on three different occasions. The left-hand spiral ---- and the right-hand spiral ++++ are both observed in three structures. The former is the predominant conformation in proteins. The sign sequence +---+ is seen in three structures of noncyclic molecules, all having a twofold axis through the central S-S bond. Even when crystallographic symmetry is not present, there seems to be a marked tendency for symmetry at the S-S bond with respect to the sign of the torsion angles. Only two structures exist where this is not the case.

For the C-S-S-C torsion angle, previous investigations give peaks in the distribution for the values -90 and $90^{\circ}.^{51}$ It is thus rather peculiar that in Table 3, no values are indicated in the intervals <-85, -95> and <85, 95>. The number of structures is however, too limited for one to tell if this observation has any significance.

Theoretical calculations. For the CH₂–S–S–CH₂ torsion angle AMBER combines a threefold and a twofold potential to ensure the well-known gauche tendency. However, for C^{α} – C^{β} –S–S only X–CH₂–S–X torsional parameters are available which describe a standard threefold potential. Evidently, these are inadequate for reproducing experimental values of χ^2 in cystine. Therefore, no energy calculations on conformations of the disulfide bridge have been performed. Instead the properties of cystine have been investigated in a more simple manner. In this procedure, all five torsion angles were varied simultaneously with χ^1 and $\chi^{1\prime}$ set to one of the three intervals [50, 70], [170, 190] or [-50, -70] while χ^2 , χ^3 and $\chi^{2\prime}$ were confined to the experimental intervals [70, 100] and [-70, -100]. Table 4 gives a

complete list of allowed sign combinations. It can be seen that only 26 out of 42 conformations are easily accessible. In particular, structures in which the three central bonds have sign sequences +++ or --- have extended conformational freedom for χ^1 and $\chi^{1\prime}$. It should be noted that sign combinations other than those indicated in Table 4 may occur if the torsion angles deviate substantially from the values normally observed. An example is the structure of BEWDIQ10 with a rather unique C-C-S-S torsion angle of -139° . The sign combination is t^-++- , which is not included in Table 4. The ring system in this molecule is rather strained, and it seems very unlikely that the same conformation would be found in a non-cyclic molecule.

Concluding remarks

It is rather surprising to see how the introduction of a serine or cysteine residue into a linear peptide seems to lock the main chain into one of only two distinct conformations in the solid phase, with the type II β turn accounting for a third possibility in cyclic compounds. Furthermore a firm connection exists between the conformation of the side chain and the main chain. The serine χ^1 distribution in proteins is 0.48:0.24:0.29 for g^+ : $t:g^-$, which deviates from the figures for serine in this survey (0.65:0.19:0.15). This may be attributed to the fact that most residues in peptides are in well-defined conformations with a preference for g^+ , in proteins a large part (ca. 41%)⁵² of the residues are in reverse turns where the preference may be tilted towards t and g^- conformations.

A similar argument may be used to explain the higher percentage of t and g^- rotamers for cysteine/cystine in proteins. The combined (major part is cystine) distributions $g^+:t:g^-$ is $0.14:0.25:0.61,^6$ figures presented in this work for peptides are 0.91:0.00:0.09 and 0.46:0.04:0.50 for cysteine and cystine, respectively. The cystine residues play a crucial role in the folding of many proteins by virtue of their ability to form links between different polypeptide chains. Short interchain distances may then impose the same restrictions on the sign combination along the disulfide bridge as described above for the smaller cyclic molecules.

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