Tobacco Chemistry. 70.* Six New Cembrane-Derived Compounds from Tobacco

Inger Wahlberg, Ann-Marie Eklund and Curt R. Enzell

Reserca AB, P.O. Box 17007, S-104 62 Stockholm, Sweden

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Six new compounds of probable cembrane origin have been isolated from Greek tobacco. They have been identified as (E)-5-hydroxy-5-isopropylhept-3-en-2-one (1), (E)-4-isopropyl-7-oxooct-5-en-4-olide (2), (2E,6E)-5-isopropyl-2-methyl-8-oxonona-2,6-dienal (3), (E)-4-hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione (4), (E)-3-hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoic acid (5) and (3E,7E)-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-diene-2,12-dione (6) by spectral methods and synthesis (1,2,4-6). The biogenesis of 1-6 is discussed.

The aroma fractions isolable from tobacco are noted for their large content and wide array of compounds, which are postulated to arise by biodegradation of diterpenoids, carotenoids and higher isoprenoids.² As an addition to these, we now report the isolation of six new cembrane-derived compounds (1–6) from tobacco.³

Results

Repetitive liquid chromatography was used to isolate compounds 1–4 from a diethyl ether extract of sun-cured leaves of Greek tobacco and compounds 5 and 6 from a chloroform extract of flowers of Greek tobacco.

Structure determination. The first new compound (1), $C_{10}H_{18}O_2$, is an α,β -unsaturated ketone [IR: 1679 cm⁻¹; ¹H NMR doublets at δ 6.34 and 6.75 (J=15.9 Hz)]. The keto group is linked to a methyl group (¹H NMR 3 H singlet δ 2.29) and the β -carbon of the enone moiety to a fully substituted hydroxy-bearing carbon atom [¹³C NMR: δ 77.8 (s); OH-absorption in the IR spectrum]. Since the latter carbon atom is also attached to an ethyl and an isopropyl group, 1 was identified as (E)-5-hydroxy-5-isopropylhept-3-en-2-one. This formulation was consistent with the mass spectrum, which includes peaks at m/z 141, 127, 99, 81, and 43 corresponding to ions arising from the fragmentation reactions outlined in Scheme 1.

^{*} For part 69 see Ref. 1.

Confirmatory structural evidence was obtained by synthesis involving bromination of (\pm) -(E)-5-isopropylhept-3-en-2-one $(7)^4$ using N-bromosuccinimide (NBS) and tertbutyl hydroperoxide in CCl₄ to give 8 and subsequent acid-catalyzed hydrolysis.

The second new compound (2), $C_{11}H_{16}O_3$, also contains an (E)-3-oxobut-1-enyl moiety that is linked to a fully substituted oxygen-bearing carbon atom [13 C NMR δ 89.7 (s)]. The latter is part of a five-membered ring lactone system (IR band at 1785 cm $^{-1}$) and carries an isopropyl substituent. These results allowed the identification of 2 as (E)-4-isopropyl-7-oxooct-5-en-4-olide.

A racemic sample of (E)-4-isopropyl-7-oxooct-5-en-4-olide, the spectral data of which were identical with those

of the natural product (2), was obtained in a low yield as follows. (\pm) -(E)-4-Isopropyl-7-oxooct-5-enenitrile (9), prepared by analogy with a method used for the synthesis of (\pm) -(5E)-4-isopropyl-7-methylocta-5,7-dienenitrile,5 was brominated using NBS. The bromo compound formed (10) was treated with acid to give, as the major product, a compound, which was identified as (\pm) -(E)-4-hydroxy-4-isopropyl-7-oxooct-5-enenitrile (11) from its spectral data, and as the minor product compound 2.

The third new compound (3), $C_{13}H_{20}O_2$, incorporates an aldehyde group and an oxo group, both being α,β -unsaturated [IR: 2711, 1693 and 1625 cm⁻¹; ¹³C NMR: δ 132.4 (d), 140.4 (s), 148.2 (d), 151.5 (d), 194.9 (d) and 197.9 (s)]. The ¹H NMR spectrum was consistent with the presence of

an isopropyl group and two methyl groups, of which one is vinylic and the other is linked to the keto group [δ : 0.93 (d), 0.98 (d), 1.75 (dt) and 2.26 (s)]. These structural fragments were allocated to a (2E,6E)-5-isopropyl-2-methyl-8-oxonona-2,6-dienal structure with the aid of the 1 H- 1 H shift correlation spectrum. The presence of crosspeaks due to long-range coupling between H-3 and H-10 and between H-7 and H-9 was thereby useful from a diagnostic point of view. The assignment of E-geometries to the 2,3 and 6,7 double bonds followed from the shielding of the aldehydic proton, δ 9.38, 6 and the magnitude of the vicinal coupling constant, J=15.9 Hz, respectively. Structural information was also provided by the mass spectrum, which contains characteristic peaks at m/z 166, 125, 84 and 43 (Scheme 1).

Attempts to synthesize 3 from norsolanadione (12) or 2-isopropyl-5-oxohexanal were unsuccessful.

The fourth new compound (4), $C_{15}H_{26}O_3$, possesses two oxo groups (^{13}C NMR signals at δ 209.4 and 210.4; IR band at 1710 cm $^{-1}$), which were allocated by the use of $^{1}H_{-}^{1}H$ shift correlation spectroscopy to partial structures **A** and **B**. These were linked via a fully substituted carbon atom, which carries a hydroxy and a methyl group (^{13}C NMR signal at δ 71.6; ^{1}H NMR methyl singlet at δ 1.28; OH-absorption in the IR spectrum) to an (E)-4-hydroxy-7-iso-propyl-4-methylundec-5-ene-2,10-dione structure. This assignment was corroborated by the mass spectrum, which includes diagnostically useful ions of masses 196, 178, 135, 71, and 43 (Scheme 1).

The synthesis of racemic 4 was performed as follows. (\pm) -Norsolanadione monoacetal $(13)^7$ was converted by a Reformatsky reaction using zinc and methyl bromoacetate into a mixture of the C-3 epimers of the methyl ester 14 [1H NMR: three-proton singlets at δ 1.30, 1.31 and 3.68]. Alkaline hydrolysis gave the acid 15, which was treated with methyllithium in THF. The desired diastereomeric ketones 16 and 17 were obtained as the major products, while a mixture of the C-4 isomers of the diol 18 (IR: 3613 and 3364 cm⁻¹), the monool 19 and norsolanadione monoacetal (13) were minor products. The diol 19 is evidently formed by dialkylation of 15, while norsolanadione monoacetal (13) is likely to arise via a retro-aldol reaction occurring with 16 and 17. Compound 18 was hydrolyzed to give (E)-2-hydroxy-5-isopropyl-2-methylnon-3-en-8-one (20), which is a tobacco constituent.7

Hydrolysis converted 17 into the desired (\pm) -(E)-4-hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione, which was identical with the new tobacco constituent 4. A C-4 epimer of 4 (21), hitherto not encountered in tobacco, was obtained by hydrolysis of 16.

One of the C-3 isomers of (6S)-(E)-3-hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoic acid (22) has previously been found in tobacco. We have now isolated this acid and an isomer thereof (5) as the corresponding methyl esters (23, 24) from tobacco. Their structures were confirmed by direct comparison with authentic racemic samples prepared

by removal of the protective group in 14 and subsequent separation.

With a sample of 14 in hand we also prepared the two (E)-1,3-dihydroxy-6-isopropyl-3-methyldec-4-en-9-ones epimeric at C-3 (25, 26) via reduction using LAH and acid-catalyzed hydrolysis. The most polar of these (25) gave rise to spectral data identical with those reported for a compound previously found in tobacco.⁹

The sixth new compound (6), $C_{17}H_{28}O_3$ contains two oxo groups, of which one is α,β -unsaturated [IR: 1718 and 1676 cm⁻¹; ¹³C NMR: δ 209.1 (s) and 198.2 (s)]. It also possesses one tertiary hydroxy group [IR: 3610 and 3430 cm⁻¹; ¹³C NMR: δ 72.5 (s)] and two 1,2-disubstituted double bonds [¹³C NMR: δ 130.2 (d), 134.2 (d), 137.9 (d) and 143.5 (d)]. These were incorporated into a (3*E*,7*E*)-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-diene-2,12-dione structure by using information from the ¹H–¹H shift correlation and mass spectra (Scheme 1).

The structure assigned to 6 was confirmed by synthesis. This involved the reaction of (±)-norsolanadione monoacetal (13) with methyl (E)-4-bromobut-2-enoate and zinc to give a 1:1 mixture of the C-5 epimers of the methyl ester 27 (¹H NMR: three proton singlets at δ 1.30, 1.32 and 3.72). Hydrolysis under alkaline conditions converted 27 into the corresponding acid 28. The latter was treated with methyllithium to give, as the major product, a 1:1 mixture of the C-6 epimers of the methyl ketone 29 (¹H NMR: three-proton singlets at δ 1.30, 1.33 and 2.25) and as the minor product diol 30 (IR: 3610 and 3430 cm⁻¹) formed by dialkylation of 28. The protective group in 29 was removed by hydrolysis under acidic conditions to give an inseparable mixture of the C-6 epimers of (3E,7E)-6-hydroxy-9isopropyl-6-methyltrideca-3,7-diene-2,12-dione. A comparison of the ¹H and ¹³C NMR data revealed that the new compound (6) was identical with one of the epimers in this mixture. The other C-6 epimer (31) has hitherto not been isolated from tobacco.

Since the new compounds (1-6) were isolated in minute quantities only (1.2-6.2 mg), it has not been possible to determine their absolute configurations.

Biogenesis. Compounds 1–6 are evidently additions to the large group of isopropyl-containing irregular isoprenoids isolable from the flavour fractions of tobacco. Available results suggest that these compounds arise by biodegradation of the cembranic diterpenoids that are present in substantial amounts in the cuticular wax of the tobacco leaf and flower. The degradation reactions are assumed to be initiated by rupture of the bonds indicated in Scheme 2.² The key metabolites formed undergo subsequent chemical alterations which may involve loss of carbon atoms.

Thus as outlined in Scheme 3, the cembra-2,7,11-triene-4,6-diols 32 and 33^{10} are the postulated precursors of norsolanadione (12),¹¹ the C_{12} key metabolite. The prerequisite breakage of their 4,5 and 11,12 bonds may take place via the intermediate formation of the *seco*-aldehyde 34, the

seco-acids 35 and 36, and/or the ketols 37 and 38 and the seco-diketone 39, which are all tobacco constituents. 12-14

Scheme 2.

Norsolanadione (12), in turn, undergoes loss of one or two carbon atoms with the formation of the C_{11} acid 40 and the C_{10} enone 7; 7 may also be formed by decarboxylation of 40. Allylic oxidation converts 40 and 7 into 2 and 1, respectively. It is noteworthy that 1 and 2 are the first degraded cembranoids encountered in tobacco, in which the isopropyl-bearing carbon atom also carries oxygen (C-1 in the parent cembranoids).

The C_{13} aldehyde 3 may formally arise via rupture of the 4,5 and 10,11 bonds in the parent cembranoid. This mode of formation was rendered likely by the recent isolation from tobacco of the plausible intermediate 41,15 but differs from that of the previously known C_{13} compounds which invariably involves breakage of the 5,6 and 11,12 bonds.

The formation of the C_{14} acids 5 and 22 in tobacco is explained by rupture of the 6,7 and 11,12 bonds in a rele-

vant cembranoid precursor and subsequent oxidation of the intermediate C_{14} aldehyde (Scheme 2).² The biogenesis of 4 and 6, on the other hand, would require the rupture of the 7,8 and 11,12 (Scheme 2) and the 8,9 and 11,12 bonds, respectively, but further insight into the course of these reactions remains to be gained.

Experimental

Optical rotations were recorded on a Perkin-Elmer 241 polarimeter. For other instrumental details see Ref. 16.

Isolation. (E)-5-Hydroxy-5-isopropylhept-3-en-2-one (1, 6.2 mg) and (2E,6E)-5-isopropyl-2-methyl-8-oxonona-2,6-dienal (3, 1.2 mg) were isolated from fraction B7 and (E)-4-isopropyl-7-oxooct-5-en-4-olide (2, 4.8 mg) and (E)-4-hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione (4, 1.6 mg) from fraction B9 of an Et₂O extract of 295 kg of Greek sun-cured tobacco¹⁷ by chromatography over silica gel using hexane/EtOAc as the eluent followed by HPLC using columns packed with Spherisorb 5 and Spherisorb 5 Nitrile and hexane/EtOAc 60:40 as the eluent.

(6S)-(E)-3-Hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoic acid (22) and an epimer thereof (5) were isolated as the corresponding methyl esters (24, 11.2 mg and 23, 2.5 mg) from fraction 2 (64 mg) of the acidic portion of a CHCl₃ extract of flowers of Greek tobacco¹⁶ by repetitive HPLC using columns packed with Spherisorb 5 Nitrile (hexane/EtOAc 60:40) and Spherisorb 5 (hexane/EtOAc 75:25).

(3E,7E)-6-Hydroxy-9-isopropyl-6-methyltrideca-3,7-diene-2,12-dione (6, 1.2 mg) was obtained from fraction C3

(272 mg) of a CHCl₃ extract of flowers of Greek tobacco¹⁸ by HPLC using columns packed with Spherisorb 5 (hexane/EtOAc 40:60) and Spherisorb 5 Nitrile (hexane/EtOAc 1:1).

(*E*)-5-Hydroxy-5-isopropylhept-3-en-2-one (1) was an oil and had $[\alpha]_D$ –4.1° (*c* 0.41, CHCl₃); (Found: $[M-29]^+$ 141.0884. Calc. for C₈H₁₃O₂: 141.0915); IR (CCl₄): 3616, 3450, 1699, 1679, 1627 and 990 cm⁻¹; ¹H NMR (CDCl₃): δ 0.86 (t, *J* 7.5 Hz, H-7), 0.91 (d, *J* 6.8 Hz)/0.93 (d, *J* 6.8 Hz) (H-9/H-10), 1.63 (dq, *J* 7.5 and –14.0 Hz, H-6a), 1.68 (dq, *J* 7.5 and –14.0 Hz, H-6b), 1.84 (septet, *J* 6.8 Hz, H-8), 2.29 (s, H-1), 6.34 (d, *J* 15.9 Hz, H-3) and 6.75 (d, *J* 15.9 Hz, H-4); ¹³C NMR (CDCl₃): δ 7.7 (C-7), 16.4/17.6 (C-9/C-10), 28.0 (C-1), 30.9 (C-6), 36.1 (C-8), 77.8 (C-5), 129.2 (C-3), 150.8 (C-4) and 198.0 (C-2); MS [m/z (%, composition)]: 141 (11, M-29), 127 (56, C₇H₁₁O₂ and C₈H₁₅O), 113 (16, C₇H₁₃O), 109 (10, C₇H₉O and C₈H₁₃), 99 (31, C₆H₁₁O), 85 (16), 81 (11, C₆H₉), 71 (22, C₄H₇O), 57 (24, C₃H₅O) and 43 (100).

(*E*)-4-Isopropyl-7-oxooct-5-en-4-olide (2) was an oil and had $[\alpha]_D$ +0.7° (*c* 0.43, CHCl₃); (Found: M^{++} 196.1108. Calc. for C₁₁H₁₆O₃: 196.1100); IR (CCl₄): 1785, 1705, 1685 and 1635 cm⁻¹; ¹H NMR (CDCl₃): δ 0.98 (d, *J* 6.4 Hz)/1.00 (d, *J* 7.0 Hz) (H-10/H-11), 2.29 (s, H-8), 6.36 (d, *J* 16.0 Hz, H-6) and 6.76 (d, *J* 16.0 Hz, H-5); ¹³C NMR (CDCl₃): δ 17.0/17.1 (C-10/ C-11), 28.2 (C-3), 29.0 (C-8), 30.4 (C-2), 37.2 (C-9), 89.7 (C-4), 129.1 (C-6), 143.9 (C-5), 176.1 (C-1) and 197.1 (C-7); MS [m/z (%, composition)]: 196 (4, M), 181 (1), 154 (40, C₈H₁₀O₃), 153 (100, C₈H₉O₃ and C₉H₁₃O₂), 125 (32, C₇H₉O₂ and C₈H₁₃O), 111 (13, C₆H₇O₂ and C₇H₁₁O), 97 (27, C₅H₅O₂ and C₆H₉O), 84 (7, C₅H₈O), 69 (9, C₄H₅O and C₅H₉), 55 (19, C₃H₃O and C₄H₇) and 43 (66, C₂H₃O and C₄H₇).

(2E,6E)-5-Isopropyl-2-methyl-8-oxonona-2,6-dienal (3) was an oil and had $[\alpha]_D +3.5^\circ$ (c 0.26, CHCl₃); (Found: $[M-43]^+$ 165.1256. Calc. for $C_{11}H_{17}O$: 165.1279); IR (CCl₄): 2820, 2711, 1693, 1625, 1371, 1359 and 985 cm⁻¹; ¹H NMR (CDCl₃): δ 0.93 (d, J 6.8 Hz)/0.98 (d, J 6.8 Hz) (H-12/H-13), 1.75 (dt, J 0.9 and 1.4 Hz, H-10), 1.80 (d septet, J 5.9 and 6.8 Hz, H-11), 2.20 (ddd, J 5.2, 5.9 and 8.2 Hz, H-5), 2.26 (s, H-9), 2.42 (dddq, J 0.9, 7.3, 8.2 and -15.2 Hz, H-4a), 2.59 (dddq, J 0.9, 5.2, 7.3 and -15.2 Hz, H-4b), 6.07 (dd, J 0.8 and 15.9 Hz, H-7), 6.39 (tq, J 1.4 and $7.3~{\rm Hz},\,{\rm H}\text{-}3),\,6.63~({\rm dd},\,J\,9.2~{\rm and}\,15.9~{\rm Hz},\,{\rm H}\text{-}6)$ and $9.38~({\rm s},$ H-1); ¹³C NMR (CDCl₃): δ 9.5 (C-10), 19.1/20.6 (C-12/ C-13), 27.4 (C-9), 31.0 (C-4), 31.5 (C-11), 48.7 (C-5), 132.4 (C-7), 140.4 (C-2), 148.2 (C-6), 151.5 (C-3), 194.9 (C-1) and 197.9 (C-8); MS [m/z (%, composition)]: 193 (1, M-15), 166 (1), 165 (2), 147 (1), 126 (16, $C_8H_{14}O$), 125 $(35, C_8H_{13}O), 107 (7, C_8H_{11}), 95 (10, C_6H_7O) and C_7H_{11}, 84$ $(22, C_5H_8O)$, 69 $(7, C_4H_5O)$ and C_5H_9 , 55 $(23, C_4H_7)$ and C_3H_3O) and 43 (100).

(*E*)-4-Hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione (**4**) was an oil and had $[\alpha]_D$ +6.2° (*c* 0.13, CHCl₃); (Found: $[M-18]^+$ 236.1793. Calc. for $C_{15}H_{24}O_2$: 236.1776); IR (CCl₄): 3500, 1710, 1385, 1365 and 980 cm⁻¹; ¹H NMR (CDCl₃): δ 0.81 (d, *J* 6.8 Hz)/0.86 (d, *J* 6.6 Hz) (H-14/

H-15), 1.28 (s, H-12), 1.44 (m, H-8a), 1.56 (m, H-13), 1.70 (m, H-7), 1.74 (m, H-8b), 2.11 (s, H-11), 2.17 (s, H-1), 2.28 (m, H-9a), 2.39 (m, H-9b), 2.63 (d, J –17.0 Hz, H-3a), 2.78 (d, J –17.0 Hz, H-3b), 4.03 (br s, –OH), 5.33 (dd, J 9.0 and 15.6 Hz, H-6) and 5.45 (d, J 15.6 Hz, H-5); ¹³C NMR (CDCl₃): δ 19.1/20.7 (C-14/C-15), 26.1 (C-8), 28.8 (C-12), 30.0 (C-11), 31.8 (C-1), 32.1 (C-13), 41.9 (C-9), 48.6 (C-7), 53.1 (C-3), 71.6 (C-4), 129.7 (C-6), 137.7 (C-5), 209.4 (C-10) and 210.4 (C-2); MS [m/z (%, composition)]: 236 (1, M–18), 218 (3, $C_{15}H_{22}O$), 196 (3), 178 (4, $C_{12}H_{18}O$), 163 (2, $C_{11}H_{15}O$), 151 (3), 135 (15, $C_{10}H_{15}$), 121 (12, $C_{9}H_{13}$), 109 (16, $C_{8}H_{13}$ and $C_{7}H_{9}O$), 95 (21, $C_{6}H_{7}O$ and $C_{7}H_{11}$), 81 (9, $C_{6}H_{9}$), 71 (13, $C_{4}H_{7}O$), 55 (11, $C_{4}H_{7}$ and $C_{3}H_{3}O$) and 43 (100, $C_{2}H_{3}O$ and $C_{3}H_{7}$).

(*E*)-Methyl 3-hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoate (**23**) was an oil and had $[\alpha]_D$ –4.2° (*c* 0.24, CH₃OH); (Found: $[M-36]^{+-}$ 234.1623. Calc. for C₁₅H₂₂O₂: 234.1620); IR (CCl₄): 3519, 1721, 1386, 1367 and 982 cm⁻¹; ¹H NMR (CDCl₃): δ 0.80 (d, *J* 6.8 Hz)/0.85 (d, *J* 6.6 Hz) (H-13/H-14), 1.32 (s, H-11), 2.12 (s, H-10), 2.56 (d, *J* –15.7 Hz, H-2a), 2.59 (d, *J* –15.7 Hz, H-2b), 3.68 (s, –OCH₃), 5.37 (dd, *J* 8.6 and 15.3 Hz, H-5) and 5.45 (d, *J* 15.3 Hz, H-4); ¹³C NMR (CDCl₃): δ 19.0/20.7 (C-13/C-14), 26.2 (C-7), 28.9 (C-11), 30.0 (C-10), 32.0 (C-12), 41.9 (C-8), 45.3 (C-2), 48.5 (C-6), 51.7 (–OCH₃), 71.0 (C-3), 130.0 (C-5), 137.1 (C-4), 173.1 (C-1) and 209.2 (C-9); MS [m/z (%)]: 234 (5, M-36), 153 (5), 135 (9), 125 (9), 121 (19), 107 (8), 97 (13), 95 (16), 93 (13), 85 (7), 81 (8), 71 (11), 69 (11), 55 (11) and 43 (100).

(6S)-(E)-Methyl 3-hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoate (24) was an oil and had $[\alpha]_D$ -3.8° (c 1.12, CH₃OH); IR (CCl₄): 3523, 1720, 1385, 1367 and 982 cm⁻¹; ¹H NMR (CDCl₃): δ 0.81 (d, J 6.8 Hz)/ 0.87 (d, J 6.6 Hz) (H-13/H-14), 1.32 (s, H-11), 2.12 (s, H-10), 2.57 (d, J -15.7 Hz, H-2a), 2.60 (d, J -15.7 Hz, H-2b), 3.69 (s, -OCH₃), 5.37 (dd, J 8.8 and 15.6 Hz, H-5) and 5.47 (d, J 15.6 Hz, H-4); ¹³C NMR (CDCl₃): δ 18.9/20.7 (C-13/C-14), 26.1 (C-7), 28.7 (C-11), 30.0 (C-10), 32.0 (C-12), 41.9 (C-8), 45.4 (C-2), 48.6 (C-6), 51.7 (–OCH₃), 71.0 (C-3), 130.0 (C-5), 137.3 (C-4), 173.1 (C-1) and 209.2 (C-9); MS $[m/z \ (\%)]$: 234 (5, M-36), 153 (4), 135 (8), 125 (8), 121 (17), 107 (8), 97 (11), 95 (14), 93 (11), 85 (6), 81 (7), 71 (10), 69 (9), 55 (10) and 43 (100). The optical rotation, IR, ¹H NMR and MS data agreed well enough with corresponding data previously reported for an authentic sample to establish the identity of 24.8

(3E,7E)-6-Hydroxy-9-isopropyl-6-methyltrideca-3,7-diene-2,12-dione (**6**) was an oil and had [α]_D 0° (c 0.16, CHCl₃); (Found: [M-18]⁺ 262.1976. Calc. for C₁₇H₂₆O₂: 262.1933); IR (CCl₄): 3610, 3421, 1718, 1676, 1628 and 982 cm⁻¹; ¹H NMR (CDCl₃): δ 0.83 (d, J 6.8 Hz)/ 0.88 (d, J 6.7 Hz) (H-16/H-17), 1.34 (s, H-14), 2.13 (s, H-13), 2.25 (s, H-1), 2.35 (m, H-11), 2.46 (dd, J 1.4 and 7.5 Hz, H-5a and H-5b), 5.38 (dd, J 8.9 and 15.6 Hz, H-8), 5.48 (d, J 15.6 Hz, H-7), 6.11 (dt, J 1.4 and 16.0 Hz, H-3) and 6.81 (dt, J 7.5 and 16.0 Hz, H-4); ¹³C NMR (CDCl₃): δ 19.1/20.7 (C-16/C-17), 26.1 (C-10), 26.8 (C-1), 28.9 (C-14), 30.0

(C-13), 32.0 (C-15), 42.0 (C-11), 45.7 (C-5), 48.7 (C-9), 72.5 (C-6), 130.2 (C-8), 134.2 (C-3), 137.9 (C-7), 143.5 (C-4), 198.2 (C-2) and 209.1 (C-12); MS [m/z (% composition]: 262 (2, M-18) 204 (1, $C_{14}H_{20}O$), 197 (21, $C_{12}H_{21}O_2$), 179 (10, $C_{12}H_{19}O$), 161 (19, $C_{12}H_{17}$), 139 (16, $C_{9}H_{15}O$), 121 (63, $C_{9}H_{13}$), 109 (33, $C_{8}H_{13}$ and $C_{7}H_{9}O$), 97 (27, $C_{7}H_{13}$ and $C_{6}H_{9}O$), 84 (23, $C_{5}H_{8}O$), 71 (54, $C_{4}H_{7}O$), 55 (18, $C_{4}H_{7}$ and $C_{3}H_{3}O$) and 43 (100, $C_{3}H_{7}$ and $C_{2}H_{3}O$).

Preparation of (\pm)-(E)-5-hydroxy-5-isopropylhept-3-en-2-one (1). To a solution of 19.6 mg of (\pm)-(E)-5-isopropylhept-3-en-2-one (7)⁴ in 5 ml of CCl₄ was added a solution of 15.8 mg of NBS and 30 µl of tert-butyl hydroperoxide in 0.5 ml of CCl₄. The reaction mixture was refluxed for 2 h, diluted with water and extracted with CCl₄. The organic phase was washed with water and concentrated. The residue was separated by chromatography over silica gel (hexane/EtOAc 98:2) to give 6.3 mg of starting material and 10.9 mg of (\pm)-(E)-5-bromo-5-isopropylhept-3-en-2-one (8), which had ¹H NMR (CDCl₃): δ 0.97 (d, J 6.9 Hz)/1.10 (d, J 6.6 Hz) (H-9/H-10), 1.01 (t, J 7.2 Hz, H-7), 1.85–2.05 (m, H-6), 2.13 (septet, J 6.8 Hz, H-8), 2.30 (s, H-1), 6.35 (d, J 15.7 Hz, H-3) and 6.68 (d, J 15.7 Hz, H-4).

To a solution of 4 ml of THF, 0.4 ml of water, and 0.2 ml of aqueous $\rm H_2SO_4$ (5%) was added a solution of 10 mg of 8 in 1 ml of THF. The reaction mixture was refluxed for 15 h. Work-up and separation by chromatography over silica gel (hexane/EtOAc 10:90) gave 2.1 mg of (\pm)-(E)-5-hydroxy-5-isopropylhept-3-en-2-one, the IR, 1 H and 13 C NMR and mass spectra of which were identical with those of the naturally occurring 1.

Preparation of (±)-(E)-4-isopropyl-7-oxooct-5-en-4-olide (2). To a solution of 188 mg of (±)-(E)-4-isopropyl-7-oxooct-5-enenitrile (9) in 6 ml of CCl₄ was added a solution of 259 mg of NBS and 35 μl of tert-butyl hydroperoxide in 1 ml of CCl₄. The reaction mixture was refluxed for 1.5 h, diluted with water and extracted with CCl₄. The organic phase was washed with water and concentrated. The residue was separated by chromatography over silica gel (hexane/EtOAc gradient) to afford 33.7 mg of starting material (9) and 60.5 mg of (±)-(E)-4-bromo-4-isopropyl-7-oxooct-5-enenitrile (10), which had ¹H NMR (CDCl₃): δ 0.99 (d, J 6.6 Hz)/1.12 (d, J 6.6 Hz) (H-10/ H-11), 1.95 (septet, J 6.6 Hz, H-9), 2.33 (s, H-8), 6.42 (d, J 15.5 Hz, H-6) and 6.52 (d, J 15.5 Hz, H-5).

A solution of 60.5 mg of 10 and 0.8 ml of aqueous H_2SO_4 (20%) in 6 ml of THF and 0.4 ml of water was refluxed for 30 h. Work-up and separation by HPLC (Spherisorb 5; hexane/EtOAc 60:40) yielded 11.6 mg of (\pm)-(E)-4-hydroxy-4-isopropyl-7-oxooct-5-enenitrile (11), which had IR (CCl₄): 3618, 3496, 1702, 1682 and 1629 cm⁻¹; ¹H NMR (CDCl₃): δ 0.95 (d, J 6.9 Hz)/0.95 (d, J 6.6 Hz) (H-10/H-11), 1.84 (septet, J 6.8 Hz, H-9), 2.31 (s, H-8), 6.37 (d, J 15.7 Hz, H-6) and 6.66 (d, J 15.7 Hz, H-5); MS [m/z (%)]: 162 (0.5, M-33), 152 (20), 124 (6), 113 (29), 110 (19), 95 (20), 82 (13), 71 (13), 67 (7), 55 (12) and 43 (100) and 2.5

mg of (\pm) -(E)-4-isopropyl-7-oxooct-5-en-4-olide, the spectral data of which were identical with those of the naturally occurring 2.

Preparation of (±)-(E)-methyl 9,9-ethylenedioxy-3-hydroxy-6-isopropyl-3-methyldec-4-enoate (14). A mixture of 288 mg of racemic norsolanadione monoacetal (13)⁷ and 133 mg of activated zinc powder in 12 ml of dry benzene was refluxed with 195 µl of methyl bromoacetate under nitrogen for 30 min. The reaction mixture was cooled (0°C), stirred with aqueous acetic acid (10%) for 30 min. and extracted with Et₂O. The organic phase was washed with aqueous acetic acid (10%), aqueous NaHCO3 and water, dried and concentrated. Flash chromatography over silica gel using hexane/EtOAc (80:20) as the eluent gave 343 mg of a 1:1 mixture of the C-3 epimers of racemic (E)-methyl 9,9-ethylenedioxy-3-hydroxy-6-isopropyl-3methyldec-4-enoate (14), which had ¹H NMR (CDCl₃): δ 0.80 (d, J 6.7 Hz)/0.84 (0.86) (d, J 6.7 Hz) (H-13/H-14),1.30 (s) / 1.31 (s) (H-10/H-11), 2.55 (d, J - 15.5 Hz, H-2a),2.59 (2.60) (d, J - 15.5 Hz, H-2b), 3.68 (3.69) (s, -OCH₃)3.92 (m, -OCH₂CH₂O-), 5.40 (5.41) (dd, J 8.3 and 15.6 Hz, H-5) and 5.46 (d, J 15.6 Hz, H-4); MS [m/z (%)]: 314 (0.2, M), 299 (1), 296 (0.1), 253 (0.4), 234 (2), 225 (3), 139 (2), 121 (3), 99 (10), 87 (100), 81 (4), 71 (3), 59 (5), 55 (5) and 43 (35).

Preparation of (\pm) -(E)-9,9-ethylenedioxy-3-hydroxy-6isopropyl-3-methyldec-4-enoic acid (15). A solution of 49 mg of 14 in 3 ml of ethanol and 0.1 ml of aqueous KOH (45 %) was kept at 50 °C and under nitrogen for 3 h. The reaction mixture was poured into water (0°C), acidified with acetic acid and extracted with EtOAc. The organic phase was washed with water, dried and concentrated to give 44 mg of a 1:1 mixture of the C-3 diastereoisomers of (\pm) -(E)-9,9-ethylenedioxy-3-hydroxy-6-isopropyl-3-methyldec-4-enoic acid (15), which had ¹H NMR (CDCl₃): δ 0.80 (0.82) (d, J 6.8 Hz)/0.85 (0.86) (d, J 6.7 Hz) (H-13/ H-14), 1.30 (1.31) (s, H-10), 1.35 (1.36) (s, H-11), 2.58 (2.61) [d, J - 15.2 (-15.6) Hz, H-2a], 2.60 (2.65) [d, J-15.2 (-15.6) Hz, H-2b], 3.94 (m, -OCH₂CH₂O-), 5.42 (5.44) (dd, J 8.6 and 15.7 Hz, H-5) and 5.50 (5.47) (d, J 15.7 Hz, H-4).

Treatment of (\pm) -(E)-9,9-ethylenedioxy-3-hydroxy-6-iso-propyl-3-methyldec-4-enoic acid (15) with methyllithium. To a solution of 33 mg of 15 in 3 ml of dry THF was added 1 ml of a dry solution of methyllithium in Et₂O (5%). After being stirred under nitrogen at 0°C for 45 min, the reaction mixture was slowly poured into 50 ml of aqueous acetic acid (10%) kept at 0°C for 30 min. Work-up and separation by HPLC (Spherisorb 5; hexane/EtOAc 60:40) gave 1.6 mg of racemic norsolanadione monoacetal (13), 2.4 mg of (\pm)-(E)-8,8-ethylenedioxy-5-isopropyl-2-methylnon-3-en-2-ol (19), 5.7 mg of (\pm)-(E)-10,10-ethylenedioxy-4-hydroxy-7-isopropyl-4-methylundec-5-en-2-one (16), 7.1 mg of a C-4 diastereoisomer (17) of 16, 1.0 mg of a 1:1 mixture of C-4

diastereoisomers of (\pm) -(E)-10,10-ethylenedioxy-7-isopropyl-2,4-dimethylundec-5-ene-2,4-diol (18) and 4.0 mg of starting material (15).

(±)-(E)-8,8-Ethylenedioxy-5-isopropyl-2-methylnon-3-en-2-ol (19) had ¹H NMR (CDCl₃): δ 0.82 (d, J 6.7 Hz)/0.87 (d, J 6.6 Hz) (H-12/H-13), 1.31 (s)/1.32 (s)/1.32 (s) (H-1/H-9/H-10), 3.93 (m –OCH₂CH₂O–), 5.36 (dd, J 9.0 and 15.7 Hz, H-4) and 5.56 (d J 15.7 Hz, H-3). MS [m/z (%)]: 256 (0.2, M), 241 (2), 223 (0.5), 195 (1), 176 (3), 136 (5), 121 (4), 111 (9), 99 (9), 87 (100), 81 (4), 71 (9), 69 (9), 59 (21), 55 (11) and 43 (89).

(±)-(E)-10,10-Ethylenedioxy-4-hydroxy-7-isopropyl-4-methylundec-5-en-2-one (**16**) had ¹H NMR (CDCl₃): δ 0.80 (d, J 6.8 Hz)/0.84 (d, J 6.8 Hz) (H-14/H-15), 1.28 (s, H-12), 1.30 (s, H-11), 2.16 (s, H-1), 2.60 (d, J –16.9 Hz, H-3a), 2.78 (d, J –16.9 Hz, H-3b), 3.92 (m, –OC H_2 C H_2 O–), 5.37 (dd, J 8.5 and 15.8 Hz, H-6) and 5.44 (d, J 15.8 Hz, H-5); ¹³C NMR (CDCl₃): δ 18.9/20.8 (C-14/C-15), 23.8 (C-11), 26.7 (C-8), 28.9 (C-12), 31.9 (C-1), 32.0 (C-13), 37.3 (C-9), 49.1 (C-7), 53.0 (C-3), 64.6/64.6 (–OC H_2 C H_2 O–), 71.7 (C-4), 110.2 (C-10), 130.1 (C-6), 137.0 (C-5) and 210.5 (C-2).

(±)-(E)-10,10-Ethylenedioxy-4-hydroxy-7-isopropyl-4-methylundec-5-en-2-one (17) had ¹H NMR (CDCl₃): δ 0.80 (d, J 6.7 Hz)/0.86 (d, J 6.7 Hz) (H-14/H-15), 1.28 (s, H-12), 1.30 (s, H-11), 2.16 (s, H-1), 2.60 (d, J –16.8 Hz, H-3a), 2.79 (d, J –16.8 Hz, H-3b), 3.93 (m –OC H_2 C H_2 O–), 5.35 (dd, J 8.9 and 15.6 Hz, H-6) and 5.45 (d, J 15.6 Hz, H-5); ¹³C NMR (CDCl₃): δ 18.9/20.8 (C-14/C-15), 23.8 (C-11), 26.6 (C-8), 28.8 (C-12), 31.9 (C-1), 31.9 (C-13), 37.2 (C-9), 49.1 (C-7), 53.0 (C-3), 64.6/64.6 (–OC H_2 C H_2 O–), 71.7 (C-4), 110.2 (C-10), 130.1 (C-6), 137.1 (C-5) and 210.4 (C-2).

(±)-(*E*)-10,10-Ethylenedioxy-7-isopropyl-2,4-dimethyl-undec-5-ene-2,4-diol (18) had IR (CCl₄): 3613 and 3364 cm⁻¹; ¹H NMR (CDCl₃): δ 0.83 (0.85) (d, *J* 7.0 Hz)/ 0.87 (0.88) (d, *J* 6.8 Hz) (H-15/H-16), 1.28 (s)/1.30 (s)/1.30 (1.31) (s)/1.32 (1.33) (s) (H-1/H-11/H-12/H-13), 1.77 (d, *J* -14.6 Hz, H-3a), 1.87 (d, *J* -14.6 Hz, H-3b), 3.92 (m, -OC H_2 C H_2 O-), 5.46 (5.49) (dd, *J* 8.6 and 15.6 Hz, H-6) and 5.58 (d, *J* 15.6 Hz, H-5); MS [m/z (%)]: 299 (0.3, M-15), 296 (0.5), 281 (0.3), 271 (0.8), 253 (0.8), 223 (2), 183 (3), 179 (3), 176 (4), 161 (3), 136 (17), 121 (14), 115 (10), 99 (13), 93 (19), 87 (99), 71 (12), 59 (23), 55 (13) and 43 (100).

Hydrolysis of (\pm) -(E)-8,8-ethylenedioxy-5-isopropyl-2-methylnon-3-en-2-ol (19). A solution of 4.1 mg of 19 in 2 ml of dioxane/aqueous (3%) H₂SO₄ (2:1) was stirred at room temperature and under nitrogen for 2 h. Work-up of the reaction mixture and separation of the crude product by HPLC (Spherisorb 5; hexane/EtOAc 60:40) gave 0.9 mg of a product which was indistinguishable from (E)-2-hydroxy-5-isopropyl-2-methylnon-3-en-8-one (20).

Hydrolysis of the (\pm) -(E)-10, 10-ethylenedioxy-4-hydroxy-7-isopropyl-4-methylundec-5-en-2-ones 16 and 17. A so-

lution of 5.7 mg of 16 in 1.5 ml of dioxane/aqueous (3%) H₂SO₄ (2:1) was stirred at room temperature and under nitrogen for 2 h. Work up and separation by HPLC (Spherisorb 5; hexane/EtOAc 40:60) yielded 3.8 mg of (±)-(E)-4-hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione (21), which was an oil and had IR (CCl₄): 3509, 1716. 1385, 1367 and 978 cm⁻¹; ¹H NMR (CDCl₃): δ 0.81 (d, J 6.8) Hz)/0.85 (d, J 6.6 Hz) (H-14/H-15), 1.28 (s, H-12), 1.42 (m, H-8a), 1.65 (m, H-13), 1.68 (m, H-7), 1.75 (m, H-8b), 2.12 (s, H-11), 2.16 (s, H-1), 2.30 (m, H-9a), 2.38 (m, H-9b), 2.62 (d, J - 17.1 Hz, H-3a), 2.78 (d, J - 17.1 Hz, H-3b),4.10 (br s, -OH), 5.32 (dd, J 9.0 and 15.5 Hz, H-6) and 5.44 (d, J 15.5 Hz, H-5); ¹³C NMR (CDCl₃): δ 19.1/20.7 (C-14/C-15), 26.2 (C-8), 29.0 (C-12), 30.0 (C-11), 31.9 (C-1), 32.0 (C-13), 41.9 (C-9), 48.6 (C-7), 52.7 (C-3), 71.6 (C-4), 129.7 (C-6), 137.6 (C-5), 209.3 (C-10) and 210.4 (C-2); MS [m/z (%)]: 236 (2, M-18), 218 (2), 196 (1), 178 (4), 163 (3), 151 (1), 135 (25), 121 (15), 109 (51), 95 (25), 81 (11), 71 (12), 55 (14) and 43 (100).

Using the method described above, 17 (7.1 mg) was converted into (\pm) -(E)-4-hydroxy-7-isopropyl-4-methylundec-5-ene-2,10-dione (4.9 mg), the IR, 1 H and 13 C NMR and mass spectra of which were identical with those of the naturally occurring 4.

Hydrolysis of (\pm) -(E)-methyl 9,9-ethylenedioxy-3-hydroxy-6-isopropyl-3-methyldec-4-enoate (14). A solution of 820 mg of 14 in 5 ml of dioxane/aqueous (3 %) H₂SO₄ (2:1) was stirred at room temperature and under nitrogen for 4 h. Work-up and flash chromatography over silica gel using hexane/EtOAc (70:30) as the eluent gave 602 mg of a mixture of (\pm) -(E)-methyl 3-hydroxy-6-isopropyl-3methyl-9-oxodec-4-enoates epimeric at C-3. Part of this mixture (10 mg) was separated by HPLC (Spherisorb 5; hexane/EtOAc 75:25); the least polar epimer (2.8 mg) had ¹H and ¹³C NMR spectra identical with those of the methyl ester (23) derived from the new (E)-3-hydroxy-6-isopropyl-3-methyl-9-oxodec-4-enoic acid (5). The most polar epimer (3.7 mg) had ¹H and ¹³C NMR spectra identical with those of the methyl ester (24) obtained from acid 22, previously reported as a tobacco constituent.8

Preparation of the (±)-(E)-1,3-dihydroxy-6-isopropyl-3-methyldec-4-en-9-ones epimeric at C-3 (25, 26). A solution of 110.6 mg of racemic methyl 9,9-ethylenedioxy-3-hydroxy-6-isopropyl-3-methyldec-4-enoate (14) in 5 ml of dry Et₂O was treated with an excess of LAH at room temperature for 4 h. The reaction mixture was worked up in the usual manner. Without further purification the crude product was dissolved in 1.5 ml of dioxane/aqueous H₂SO₄ (3%) (2:1) and kept at room temperature and under nitrogen for 3 h. Work-up and flash chromatography over silica gel using a hexane/EtOAc gradient as the eluent gave 48 mg of a mixture of C-3 diastereomers. Part of this mixture (16 mg) was separated by HPLC (Spherisorb 5; hexane/EtOAc 30:70) into 25 and 26.

The most polar of these (\pm) -(E)-1,3-dihydroxy-6-iso-

propyl-3-methyldec-4-en-9-ones epimeric at C-3 (5.8 mg) had IR, ¹H and ¹³C NMR and mass spectra identical with those of naturally occurring 25.⁹

The least polar epimer (26, 5.5 mg) had IR (CHCl₃): 3603, 3491, 1710 and 981 cm⁻¹; ^{1}H NMR (CDCl₃): δ 0.85 (d, J 6.8 Hz)/0.89 (d, J 6.7 Hz) (H-13/H-14), 1.32 (s, H-11), 1.70 (ddd, J 3.9, 5.5 and -14.7 Hz, H-2a), 1.87 (ddd, J 4.9, 8.2 and -14.7 Hz, H-2b), 2.12 (s, H-10), 2.33 (m, H-8a), 2.40 (m, H-8b), 3.82 (ddd, J 4.9, 5.5 and -10.9 Hz, H-1a), 3.86 (ddd, J 3.9, 8.2 and -10.9 Hz, H-1b), 5.43 (dd, J 8.3 and 15.5 Hz, H-5) and 5.45 (d, J 15.5 Hz, H-4); 13 C NMR (CDCl₃): δ 19.1/20.8 (C-13/C-14), 26.3 (C-7), 29.7 (C-11), 30.1 (C-10), 32.1 (C-12), 42.0 (C-8), 42.6 (C-2), 48.8 (C-6), 60.4 (C-1), 74.1 (C-3), 129.7 (C-5), 138.1 (C-4) and 209.4 (C-9); MS [m/z (%)]: 224 (2, M-18), 197 (2), 194 (3), 181 (1), 179 (2), 161 (5), 151 (3), 139 (6), 136 (9), 121 (26), 109 (13), 97 (21), 95 (13), 93 (18), 81 (16), 71 (26), 55 (20) and 43 (100).

Preparation of (\pm) -(2E,6E)-methyl 11,11-ethylenedioxy-5hydroxy-8-isopropyl-5-methyldodeca-2,6-dienoate (27). A mixture of 303 mg of (±)-norsolanadione monoacetal (13) and 139 mg of activated zinc powder in 12 ml of dry benzene was refluxed with 325 μ l of (E)-methyl 4-bromobut-2enoate under nitrogen for 2 h. Work-up and flash chromatography (silica gel; hexane/EtOAc gradient) gave 130 mg of starting material (13) and 97 mg of a 1:1 mixture of the C-5 epimers of racemic (2E,6E)-methyl 11,11-ethylenedioxy-5-hydroxy-8-isopropyl-5-methyldodeca-2,6-dienoate (27), which had IR (CCl₄): 3610, 3483, 1726, 1658 and 982 cm⁻¹; ¹H NMR (CDCl₃): δ 0.82 (d, J 6.8 Hz)/0.87 (d, J 6.8 Hz) (H-15/H-16), 1.30 (s)/1.32 (s) (H-12/H-13), 2.44 (dd, J 1.3 and 7.7 Hz, H-4a and H-4b), 3.72 (s, $-OCH_3$), 3.93 (m, $-OCH_2CH_2O_-$), 5.40 (dd, J 8.8 and 15.6 Hz, H-7), 5.48 (5.49) (d, J 15.6 Hz, H-6), 5.87 (5.88) (dt, J 1.3 and 15.7 Hz, H-2) and 6.96 (6.97) (dt, J 7.7 and 15.7 Hz, H-3); ¹³C NMR (CDCl₃): δ 18.9/20.8 (C-15/C-16), 23.8 (C-12), 26.5 (26.6) (C-9), 28.3 (28.6) (C-13), 31.9 (C-14), 37.3 (C-10), 45.5 (45.6) (C-4), 49.1 (C-8), 51.4 (-OCH₃), 64.6/64.6 $(-OCH_2CH_2O-)$, 72.5 (C-5), 110.2 (C-11), 124.1 (C-2), 130.6 (130.7) (C-7), 137.5 (C-6), 144.7 (144.8) (C-3) and 166.5 (C-1); MS [m/z (%)]: 325 (2, M-15), 241 (1), 223 (3), 183 (7) 179 (9) 161 (4), 139 (9), 121 (14), 115 (12), 109 (9), 99 (10), 87 (100), 81 (6), 71 (21), 59 (10), 55 (12) and 43 (93).

Hydrolysis of (\pm)-(2E,6E)-methyl 11,11-ethylenedioxy-5-hydroxy-8-isopropyl-5-methyldodeca-2,6-dienoate (27). A solution of 49 mg of 27 in 3 ml of ethanol and 0.1 ml of aqueous KOH (45%) was kept at 50 °C and under nitrogen for 2 h. Work-up gave 30 mg of a crude 1:1 mixture of the C-5 diastereoisomers of (\pm)-(2E,6E)-11,11-ethylenedioxy-5-hydroxy-8-isopropyl-5-methyldodeca-2,6-dienoic acid (28), which had ¹H NMR (CDCl₃): δ 0.82 (d, J 6.8 Hz)/0.87 (d, J 6.8 Hz) (H-15/H-16), 1.30 (s, H-12), 1.33 (s, H-13), 2.47 (dd, J 1.4 and 7.6 Hz, H-4a and H-4b), 3.93 (m, -OC H_2 C H_2 O-), 5.40 (dd, J 8.8 and 15.7 Hz, H-7), 5.49 (d,

J 15.7 Hz, H-6), 5.87 (dt, *J* 1.4 and 15.6 Hz, H-2) and 7.06 (7.07) (dt, *J* 7.6 and 15.6 Hz, H-3); MS [*m/z* (%)]: 311 (1, *M*-15), 225 (1), 223 (1), 183 (3), 179 (3), 161 (2), 139 (3), 121 (5), 115 (5), 99 (7), 87 (100), 81 (4), 71 (8), 59 (6), 55 (7) and 43 (69).

Treatment of (\pm) -(2E,6E)-11,11-ethylenedioxy-5-hydroxy-8-isopropyl-5-methyldodeca-2,6-dienoic acid (28) with methyllithium. To a solution of 30 mg of the crude 28 in 3 ml of dry THF was added 0.8 ml of a dry solution of methyllithium in $Et_2O(5\%)$. After being stirred under nitrogen at 0°C for 1 h, the reaction mixture was slowly poured into 50 ml of aqueous acetic acid (10%) kept at 0°C. Work-up of the reaction mixture and separation of the crude product by HPLC (Spherisorb 5; hexane/EtOAc 1:1) gave as the main products 6.0 mg of a 1:1 mixture of C-6 diastereoisomers of (\pm) -(3E,7E)-12,12-ethylenedioxy-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-dien-2-one (29) and 2.4 mg of a 1:1 mixture of C-6 diastereoisomers of (\pm) -(3E,7E)-2,6-dimethyl-12,12-ethylenedioxy-9-isopropyltrideca-3,7-diene-2,6-diol (30).

 (\pm) -(3E,7E)-12,12-Ethylenedioxy-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-dien-2-one (29) had ¹H NMR (CDCl₃): δ 0.83 (d, J 6.8 Hz)/0.87 (d, J 6.8 Hz) (H-16/ H-17), 1.30 (s, H-13), 1.33 (s, H-14), 2.25 (s, H-1), 2.46 (dd, J 1.3 and 7.5 Hz, H-5a and H-5b), 3.93 (m, -OCH₂CH₂O-), 5.41 (dd, J 8.7 and 15.6 Hz, H-8), 5.50 (5.51) (d, J 15.6 Hz, H-7), 6.10 (dt, J 1.3 and 16.0 Hz, H-3) and 6.81 (dt, J 7.5 and 16.0 Hz, H-4); 13 C NMR (CDCl₃): δ 18.9/20.8 (C-16/C-17), 23.8 (C-13), 26.5 (26.6) (C-10), 26.7 (26.8) (C-1), 28.6 (28.9) (C-14), 31.9 (C-15), 37.4 (37.5) (C-11), 45.7 (45.8) (C-5), 49.1 (C-9), 64.6/64.6 $(-OCH_2CH_2O-)$, 72.5 (72.6) (C-6), 110.1 (110.2) (C-12), 130.6 (130.7) (C-8), 134.2 (C-3), 137.5 (C-7), 143.6 (143.7) (C-4) and 198.4 (C-2); MS [m/z (%)]: 309 (0.1, M-15), 306 (0.1), 225 (1), 183 (1), 179 (1), 161 (1), 139 (2), 121 (3), 115 (3), 109 (2), 99 (7), 95 (4), 87 (100), 71 (5), 69 (7), 59 (8), 55 (6) and 43 (67).

(±)-(3*E*,7*E*)-12,12-ethylenedioxy-9-isopropyl-2,6-dimethyltrideca-3,7-diene-2,6-diol (**30**) had IR (CCl₄): 3610, 3430 and 978 cm⁻¹; ¹H NMR (CDCl₃): δ 0.83 (d, *J* 6.8 Hz)/0.88 (d, *J* 6.7 Hz) (H-17/H-18), 1.28 (s)/1.30 (s)/1.30 (1.31) (s)/1.31 (s) (H-1/H-13/H-14/H-15), 2.23 (2.25) [dd, *J* 7.3 (6.1) and -13.8 (-13.6) Hz, H-5a], 2.30 [dd, *J* 5.2 (6.1) and -13.8 (-13.6) Hz, H-5b], 3.93 (m, -OC H_2 C H_2 O-), 5.34 (dd, *J* 9.0 and 15.6 Hz, H-8), 5.46 (5.47) (d, *J* 15.6 Hz, H-7), 5.62 [ddd, *J* 5.2, 7.3 and 15.6 Hz (dt, *J* 6.1 and 15.6 Hz), H-4] and 5.71 (d, *J* 15.6 Hz, H-3);/MS [m/z (%)]: 325 (0.1, M-15), 304 (1), 241 (2), 223 (3), 202 (4), 183 (6), 179 (7), 159 (12), 121 (10), 115 (11), 99 (7), 93 (8), 87 (84), 82 (28), 71 (18), 55 (14) and 43 (100).

Hydrolysis of (\pm) -(3E,7E)-12,12-ethylenedioxy-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-dien-2-one (29). A solution of 6.0 mg of 29 in 1.5 ml of dioxane/aqueous (3 %) H_2SO_4 (2:1) was stirred at room temperature and under nitrogen for 2 h. Work-up of the reaction mixture and purification of

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the crude product by HPLC (Spherisorb 5; hexane/EtOAc 1:1) yielded 3.6 mg of a 1:1 mixture of the C-6 diastereoisomers of (\pm) -(3E,7E)-6-hydroxy-9-isopropyl-6-methyltrideca-3,7-diene-2,12-diones, the IR and mass spectra of which were identical with those of the naturally occurring 6. The ¹H and ¹³C NMR spectra of 6 were superimposable on those of the mixture. The NMR spectra also contained signals derived from the other C-6 diastereomer 31: ¹H NMR (CDCl₃): δ 0.83 (d, J 6.8 Hz)/0.88 (d, J 6.7 Hz) (H-16/H-17), 1.34 (s, H-14), 2.13 (s, H-13), 2.25 (s, H-1), 2.35 (m, H-11a and H-11b), 2.46 (dd, J 1.4 and 7.5 Hz, H-5a and H-5b), 5.37 (dd, J 8.9 and 15.6 Hz, H-8), 5.49 (d, J 15.6 Hz, H-7), 6.12 (dt, J 1.4 and 16.0 Hz, H-3) and 6.82 (dt, J7.5 and 16.0 Hz, H-4); 13 C NMR (CDCl₃): δ 19.1/20.7 (C-16/C-17), 26.1 (C-10), 27.0 (C-1), 28.6 (C-14), 30.0 (C-13), 32.0 (C-15), 42.0 (C-11), 45.7 (C-5), 48.7 (C-9), 72.5 (C-6), 130.3 (C-8), 134.0 (C-3), 137.8 (C-7), 143.6 (C-4), 198.4 (C-2) and 209.3 (C-12).

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