A Convenient Preparation of *N*-Protected Nucleosides with the 2,2,2-Trichloro-*t*-butyloxycarbonyl (TCBOC) Group. Structural Assignment of *N*,*N*-bis-TCBOC Guanoside and Its 2'-Deoxy Analogue

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TCBOC protected derivatives of nucleosides have been prepared using a "one-pot" procedure involving protection of the hydroxyl functions with trimethylsilyl chloride. Spectroscopic studies showed that the TCBOC groups of the guanine moiety of bis-TCBOC-guanosine and its 2'-deoxy analogue are at N^1 and at the N^2 positions respectively; while in the mono-TCBOC derivatives, the TCBOC group is, as expected, at the N^2 position.

The 2,2,2-trichloro-t-butyloxycarbonyl- (TCBOC) group was proposed ¹ for the protection of the exocyclic amino function of adenosine and for the protection of the urethane function of uridine. ² Subsequently, the TCBOC-protected derivatives of cytidine and guanosine and their 2'-deoxy analogues have also been prepared. ³ Both groups employed a two-step procedure for the preparation of these base-protected nucleosides which made the employment of TCBOC group in oligoribo- and deoxyribonucleotide synthesis time consuming. It may be noted that uridine, cytidine and adenosine, both in deoxyribo and ribo series, gave only mono-TCBOC derivatives; while guanosine and 2'-deoxyguanosine ³ gave bis-TCBOC derivatives. The structures of these bis-TCBOC derivatives of guanosine and its 2'-deoxy analogue were not clearly elucidated; nevertheless, evidence was advanced to support that the second TCBOC group could be at the N¹ and not at the O⁶ position. We herein describe a simple and general procedure for the "one-step" preparation of TCBOC protected nucleosides with free sugar hydroxyl groups and subsequently, we present direct spectroscopic arguments for the chemical structure of bis-TCBOC-guanosine and its 2'-deoxy analogue.

The general procedure for the "one-pot" synthesis of the TCBOC-protected nucleosides, I to I0, involved trimethylsilylation of a nucleoside in dry pyridine solution at 20 °C, which is followed by the addition of TCBOC-Cl in situ and then hydrolysis. Standard work-up and crystallization from aqueous ethanol gave compound I to I0 in 60-95 % yield respectively (experimental section). Element analysis and spectroscopic properties of these newly synthesized compounds were identical to the authentic ones. I^{-3} We believe that such a

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"one-step" preparation of TCBOC-protected nucleosides, 1 to 10, would promote their applications in the areas of DNA and tRNA synthesis.

We then elucidated the position of the second TCBOC group in 9 and 10, in comparison with the corresponding mono-TCBOC derivatives 7 and 8, respectively. A perusal of their ultraviolet absorption spectra at pH 2, 7 and 13 show that the compounds 7 and 8 absorb at 257 and 278 nm(sh). While the former band has a bathochromic shift (ca. 12 nm) in the alkaline pH, the latter undergoes a hypsochromic shift. Compounds 9 and 10, on the other hand, have absorptions at 255 and 245 nm(sh) at pH 2 and 7. These bands undergo a much smaller bathochromic shift in alkaline pH (4 to 5 nm). A comparison of infrared spectra of compounds 9 and 10 with 7 and 8 respectively, shows that the stretching frequencies of the conjugated carbonyl groups originate, in the latter, in the region of 1710-1690 cm⁻¹, while in the former, one observes a non-conjugated carbonyl stretching frequency at 1820 cm⁻¹ beside other conjugated carbonyl frequencies (1720-1680 cm⁻¹). It may be added that such a small high frequency shift of carbonyl frequency has been also observed 5 upon the protonation of the N^7 -nitrogen of the guanine residue. A comparison of ¹³C chemical shifts of compounds 7 to 10 with guanosine and deoxyguanosine (Table 1) show that the chemical shifts of the C^6 -carbon atoms were very similar, suggesting that the nature of the sp² hybridized carbon in these compounds have remained unchanged. However, a comparison of the chemical shifts of the C^2 -carbons in guanosine and its mono- and bis-TCBOC derivatives has revealed that the C^2 -carbon is more shielded by 1.9 and 10.5 ppm in 7 and 9, respectively, in comparison with its chemical shift in guanosine. A similar correlation is also observed for the chemical shift of the C^2 -carbon in comparison with the 13 C spectra of 2'-deoxyguanosine and its mono- and bis-TCBOC derivatives. It is clear that the attachment of an acyl group to the ring nitrogen would appear to favour localization of ring π -electrons in the various double bonds within the ring which is very similar to a situation that exists in the case of a protonation. Such a localization effect, due to N^1 -acylation, would be expected, in an analogy with the " α -protonation effect", to be most pronounced on the C^2 -carbon as seen in its shielding (ca. 10.5 ppm) in the derivatives 9 and 10 as compared to 7 and 8 respectively; while the small change of the chemical shifts of the C⁶-carbon can be attributed to its overall non-conjugated nature from the rest of ring π -electrons.

These spectroscopic properties, along with the observed stability towards alkali, of the bis-TCBOC derivatives of guanosine and deoxyguanosine, 9 and 10, are consistent with their 1,2-bis-N-TCBOC structures, 11 and 12, respectively.

Further indirect evidence for the bis-TCBOC structures, 11 and 12, has emerged by the 1H NMR study of the exchangeable protons of the aglycones. Such a study has shown that the N^1 and N^2 protons of 7 and 8, in anhydrous deuterated dimethylsulfoxide absorb at δ 11.48 and 11.38 respectively. These protons are shifted to δ 11.22 and 10.5 respectively upon warming the solutions to 70 °C; these chemical shifts remained unchanged upon returning to

the ambient temperature. In contrast, the N-H proton of the bis-TCBOC derivatives, 9 and 10, absorbed at δ 13.0 which, upon warming at 70 °C, shifted to the high field region where all other exchangeable sugar protons absorb (above δ 6.0): Upon cooling to the ambient temperature, the N-H proton reverted to the original chemical shift. This suggests that there is very little hydrogen bonding between N^1 H and N^2 -acyl function in 7 and 8; on the other hand, a strong hydrogen bonded structure for the bis-TCBOC derivatives, like 13, seems to be possible.

EXPERIMENTAL

 1 H NMR spectra were measured (δ scale) at 60 MHz with a Perkin-Elmer R 600 and at 90 MHz with a Jeol FX 90Q spectrometer using tetramethylsilane as an internal standard. 13 C NMR spectra were recorded at 23.7 MHz in the same solvent mixture. UV absorption spectra were recorded with a Cary 2200 spectrophotometer in pure methanol. Reactions were monitored by using Merck pre-coated silica gel 60 F₂₅₄ plates. 2,2,2-Trichloro-tert.-butyloxychloroformate (TCBOC-Cl) was prepared using a literature procedure. 1,3

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Table 1. ¹³C-absorptions from the base residues (δ scale).

	C-6	C-2	C-4	C-8	C-5
Guanosine	157.12	153.7	151.4 (4.5 Hz)	136.0 (214 Hz)	116.6 (11.2 Hz)
(7)	155.2	151.8	148.9 (2 Hz)	137.9 (215.6 Hz)	120.0 (11.2 Hz)
(9)	156.7	143.2	147.1 (1.5 Hz)	139.6 (212.3 Hz)	123.5 (11.2 Hz)
2'-Deoxyguanosine	157.4	153.8	151.1 (1 Hz)	136.0 (214.6 Hz)	116.6 (11.2 Hz)
(8)	155.2	151.8	148.4 (1.5 Hz)	137.7 (211.2 Hz)	120.0 (12.3 Hz)
(10)	156.8	143.0	146.8 (2.5 Hz)	139.5 (213.0 Hz)	123.7 11.2 Hz)

Preparation of 6-N-(2,2,2-trichloro-t-butyloxycarbonyl)adenosine (1) A general procedure: Adenosine (1.34g, 5 mmol) was dried by co-evaporation with dry pyridine ($2 \times ca$. 10 ml) and was then re-dissolved in the same solvent (50 ml). To this solution was added trimethylsilyl chloride (10 eq. per mmol of Nucleoside) under an atmosphere of argon at 20 °C. The reaction was monitored by TLC using 30 % ethanol-chloroform mixture as an eluent. When TLC showed the formation of a single higher R_f product, TCBOC-Cl (1.25 g, 5.2 mmol) was added. After 3h at 20 °C, TLC (5 % ethanol-chloroform mixture) showed a complete conversion to a higher R_f product. Water (2.0 ml) was added. After about 30 min, a TLC analysis (20 % ethanol-chloroform mixture) revealed a mixture of two compounds in the reaction mixture. Aqueous ammonia (d 0.9) was added to adjust the pH to 9-10. Water (20 ml) was added after 30 min stirring; the reaction mixture was subsequently extracted with chloroform (3 \times 20 ml). The organic layers were pooled, dried (MgSO₄) and concentrated on a rotavapor to a gum. The residue was first dissolved in a volume of pyridine (ca. 0.2 ml) and then taken up in chloroform (ca. 1.5 ml). This solution was subsequently precipitated from light petroleum. The precipitate was subsequently crystallized from 20 % ethanol-water mixture. Yield 2.14 g (91.2 %); mp. 162 °C; UV: λ_{max} 267 nm (ε 17000) (pH 7); 268 nm (ε 13500) (pH 2); 294 nm (ε 17500) (pH 13). IR (nujol): 1750 cm⁻¹; ¹H NMR (CDCl₃): 8.67 (ε , 1H); 8.2 (ε , 1H); 5.92 (d, 7 Hz, 1H); 4.85 (m, 1H); 4.38 (m,

2H); 3.88 (m, 2H), 2.03 (s, 6H).

Compound (2). This was prepared using essentially similar procedures described for the compound 1. A precipitate of 2 was crystallized from ca. 15 % aqueous ethanol in 82.3 % yield. mp. 168 °C.

UV: λ_{max} 267 nm (ε 17500) (pH 7); 268 nm (ε 14000) (pH 2); 294 nm (ε 18000) (pH 13); IR (nujol): 1755 cm⁻¹; ¹H NMR (CDCl₃): 8.73 (s, 1H); 8.20 (s, 1H); 6.42 (*dd*, 6 Hz, 1H); 4.7 (m, 1H); 4.19 (m, 1H); 3.88 (m, 2H); 2.68 (m, 2H); 2.05 (s, 6H).

Compound (3). Essentially, a similar reaction condition as for 1 was used for the preparation of 3; however the solubility of 3 allowed it to be dissolved in a small volume of

preparation of 3; nowever the solubility of 3 allowed it to be dissolved in a small volume of chloroform for precipitation in light petroleum. The precipitate was crystallized from ca. 5 % ethanol-chloroform mixture in 95.5 % yield. mp. 125 °C.

UV: λ_{max} 295 nm (ε 8000), 241 nm (ε 15500) (pH 7); 295 nm (ε 7200), 241 nm (ε 12000) (pH 2); 293 nm (ε 17000) (pH 13). IR (nujol): 1755 cm⁻¹. ¹H NMR (CDCl₃): 8.49 (d, 8 Hz, 1H); 7.25 (d, 8 Hz, 1H); 5.75 (s, 1H); 4.21 (bs, 3H); 3.91 (m, 2H); 1.98 (s, 6H).

Compound: (4). This was also prepared using a reaction confidence of the solubility proportion approach it to be dissolved in a small volume of

however, the solubility properties permitted it to be dissolved in a small volume of chloroform for precipitation in light petroleum. The precipitate was subsequently crystallized from ca. 5 % ethanol-chloroform mixture in 93 % yield. mp. 155 °C.

UV: λ_{max} 294 nm (ε 7000), 240 nm (ε 13500) (pH 7); 295 nm (ε 6300), 240 nm (ε 10500) (pH 2); 293 nm (ε 14500) (pH 13). IR (nujol): 1750 cm⁻¹. ¹H NMR (CDCl₃): 8.29 (d, 8.2 Hz, 1H); 7.16 (d, 8.2 Hz, 1H); 6.12 (dd, 6 Hz, 1H); 4.36 (m, 1H); 4.30 (m, 1H); 3.84 (m, 2H); 2.50 (m, 2H); 1.94 (s, 6H).

Compound: (5). This has been prepared using a very similar condition as reported for the preparation of I except that the TCBOC-Cl reaction time is around 24 h. The precipitate has been crystallized from ca. 5% ethanol-chloroform mixture in 89% yield. mp. 136°C. UV: λ_{max} 263 nm (ε 8500), (pH 7), 262 nm (ε 8000) (pH 2); 265 nm (ε 5000) (pH 13). IR (nujol): 1785, 1720 and 1680 cm⁻¹. HNMR (CDCl₃): 8.17 (d, 8.2 Hz, 1H); 6.0 (d, 8.2 Hz,

1H); 6.0 (d, 4Hz, 1H); 4.44 (m, 3H); 4.09 (m, 2H); 2.31 (s, 6H).

Compound: (6). This has been prepared using a very similar condition as reported for the preparation of l except that TCBOC-Cl reaction time is around 24 h. The precipitate has been crystallized from ca.5% ethanol-chloroform mixture in 88 % yield, mp. 132 °C. UV: λ_{max} 270 nm (ε 9500), (pH 7); 270 nm (ε 8000) (pH 2): 270 nm (ε 8000) (pH 13). IR (nujol): 1795, 1715 and 1660 cm⁻¹. ¹H NMR (CDCl₃): 7.61 (bs, 1H); 6.17 (dd, 6.9 Hz, 1H); 4.37 (m, 1H); 3.80 (m, 3H); 2.25 (m, 2H); 2.07 (s, 6H); 1.93 (bs, 3H).

Compound (7). Same procedure as for the preparation of I except that 1.05 equiv. of TCBOC-Cl was used for the reaction for 3 h. The precipitate was crystallized from ca. 10 % ethanol-water mixture in 72 % yield. mp. 224 °C. UV: λ_{max} 257 nm (ε 15000), 278 nm (ε 10300) (pH 7); 257 nm (ε 1400), 278 nm (ε 9600) (pH 2); 268 nm (12800) (pH 13). IR (nujol): 1710 and 1695 cm⁻¹. H NMR (DMSO- d_6): 8.26 (s, 1H); 5.83 (d, 6.1 Hz, 1H); 4.50 (m, 1H); 4.05 (m, 4H); 1.95 (s, 6H).

Compound (8). Same procedure as for compound 7 was used. The precipitate was crystallized from ca. 10 % ethanol—water mixture in 60 % yield. mp. 214 °C. UV: λ_{max} 257 nm (ε 16000), 278 nm (ε 11300) (pH 7); 257 nm (ε 12300), 278 nm (ε 9800) (pH 2); 268 nm (ε 14600) (pH 13). IR (nujol): 1705 and 1690 cm⁻¹. ¹H NMR (DMSO- d_6): 8.22 (s, 1H); 6.22 (s, 6.2 Hz, 1H); 4.46 (s, 1H); 4.0 (s, 3H); 2.52 (s, 2H); 1.94 (s, 6H).

Compound (9). Same procedure as for the preparation of 1 was used except that 2.5 equiv. of TCBOC-Cl was used for reaction for ca. 16 h and then a hydrolysis step with water. The precipitate was crystallized from ca. 25 % ethanol-water mixture in 90 % yield. mp. 252 °C. UV: λ_{max} 252 nm (ε 7700), 244 nm (sh) (pH 7); 252 nm (ε 6500), 244 nm (sh) (pH 2); 256 nm (ε 7500) (pH 13). IR (nujol): 1820, 1705 and 1695 cm⁻¹. ¹H NMR (DMSO-d₆): 8.37 (s, 1H); 5.78 (d, 6 Hz, 1H); 4.44 (dd, 6 Hz, 1H); 3.96 (m, 2H); 3.66 (m, 2H); 1,90 (s, 12H).

Compound (10). Same procedure as for compound 9 was used. The precipitate was

crystallized from ca. 25 % ethanol-water mixture in 88 % yield. mp. 234 °C.

UV: λ_{max} 252 nm (\$\varepsilon\$7600), 245 nm (sh) (pH 7); 252 nm (\$\varepsilon\$6400), 244 nm (sh) (pH 2); 257 nm (\$\varepsilon\$7400) (pH 13). IR (nujol): 1820, 1720 and 1705 cm⁻¹. ¹H NMR (DMSO- d_6): 8.30 (\$\varepsilon\$, 1H); 6.22 (\$\varepsilon\$d, 7.2 Hz, 1H); 4.36 (\$\varepsilon\$, 1H); 3.78 (\$\varepsilon\$, 3H); 2.52 (\$\varepsilon\$, 2H); 1.88 (\$\varepsilon\$, 12H).

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