Tobacco Chemistry. 60.* Five New Hydroperoxycembratrienediols from Tobacco

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Five new diterpenoids of the cembrane class have been isolated from flowers of Greek tobacco and shown to be (1S,2E,4S,6R,7E,11S)- and (1S,2E,4R,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol (1,2) and (1S,2E,4S,6R,7E,10E,12S)-, (1S,2E,4S,6R,7E,10E,12R)- and (1S,2E,4R,6R,7E,10E,12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (3-5) by spectral and chemical methods. The biogenesis of these compounds is discussed.

The cembranic diterpenoids, which are present in the gummy exudate of the leaf and flower of most tobacco varieties, include as the major components the (1S,2E,4S,6R,7E,11E)- and (1S,2E,4R,6R,7E,11E)-2,7,11-cembratriene-4,6-diols (6,7). These two compounds are the postulated precursors of the majority of the other tobacco cembranoids. We now report the isolation and biomimetic synthesis of five new cembranoids, which are likely to arise in tobacco by oxidation of the 4S,6R- and 4R,6R-diols (6,7).

Results. The first two compounds (1, 2) gave rise to ¹H and ¹³C NMR spectra consistent with the presence of an isopropyl group, a methyl group linked to a fully substituted, oxygencarrying carbon atom, a vinylic methyl group and three double bonds. Of these, one is \bar{E} -disubstituted, one is trisubstituted and one extends to an exocyclic methylene group. Two protons, resonating at δ 4.28 and 4.66 for 1 and at δ 4.30 and 4.68 for 2, are evidently attached to oxygenbearing carbon atoms. These results suggest that the new compounds (1, 2) are structurally related the 2,7,12(20)-cembratriene-4,6,11-triols, to which have previously been isolated from tobacco.1,3

A comparison of the ¹³C NMR data confirmed this and revealed that the C-1 to C-9 and C-14 to C-19 signals are present at virtually invariant positions in the spectra of *1* and (1*S*,2*E*,4*S*,6*R*,

Table 1. 13 C NMR chemical shifts and assignments for compounds $I-5$ and $8-12.^a$	_	
	C-18	33.1. 33.1. 33.1. 33.1. 33.1. 33.1. 31.6
	C-17	20.5 20.5 20.5 20.6 20.6 20.6 21.8 21.6 21.6
	C-16	19.2 18.8 17.9 17.9 17.9 19.2 18.7 17.8 17.8
	C-15	32.2 30.2 30.2 30.2 30.2 29.8 31.9 30.1 29.8
	C-14	29.0 28.6 27.4 25.9 29.3 30.3 26.5 28.0 28.0
	C-13	29.6 33.3 35.8 35.9 34.6 40.1 39.7
	C-12	147.1 148.6 85.0 86.4 85.0 151.6 152.4 74.0 ⁶ 73.9
	C-11	88.1 86.2 133.9 134.0 133.6 74.8 73.4 138.9 138.1
	C-10	34.6 35.0 126.9 130.0 128.0 32.9 32.5 124.6 126.1
	C-9	30.2 31.3 41.0 41.1 41.7 29.8 30.3 40.7 40.0
	C-8	137.5 138.7 135.2 135.4 138.3 137.8 139.1 134.5 135.7
	C-7	128.5 129.8 127.9 127.6 129.4 128.5 129.6 128.4 128.4
	C-6	68.5 64.7 64.9 64.9 66.2 66.2 66.2
	C-5	46.2 52.1 52.1 47.0 52.1 46.5 51.8 47.2 52.3
	C-4	74.2 72.3 73.9 72.0 72.1 72.1 74.3 74.3 74.3
	C-3	139.7 138.4 138.5 136.7 139.8 137.1 138.3 138.4
	C-2	127.9 129.8 127.9 128.1 129.4 127.4 127.4 127.3 127.3
	C-1	48.7 50.7 50.8 48.3 49.0 47.6 49.9 49.3
Table	Com- pound	12 10 10 11 12

[&]amp;Values in CDCl₁ relative to TMS. ^b Assignment may be reversed.

^{*} For part 59 see Ref. 1.

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^{0302-4369/83 \$2.50}

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7E.11S)-2.7.12(20)-cembratriene-4.6.11-triol (8.3) cf. Table 1). However, since the shieldings of C-11 were found to be markedly different. δ 88.1 for 1 as against δ 74.8 for 8, and since the ¹H NMR spectrum of 1 exhibits a broad singlet at δ $8.05,^4$ \dot{l} was provisionally formulated as a (1S, 2E, 4S, 6R, 7E)-11-hydroperoxy-2, 7, 12(20)-cembratriene-4,6-diol. Analogous findings obtained by a comparison of the ¹³C NMR spectra of 2 and (1S, 2E, 4R, 6R, 7E, 11S)-2, 7, 12(20)-cembratriene-4.6.11-triol (9)1 allowed the identification of 2 as a $(1S, 2E, 4R, 6R, 7E, 11\xi)$ -11-hydroperoxy-2,7,12 (20)-cembratriene-4,6-diol. These assignments were in harmony with the mass spectra of 1 and 2, which displayed a peak at m/z 320 (M-18) due to a $C_{20}H_{32}O_3$ ion.

The remaining three new compounds (3-5) were also hydroperoxides [broad singlets at δ 8.22, 7.33 and 7.43 in the ¹H NMR spectra of 3-5, respectively; ¹³C NMR signals at δ 85.0 (3, s), δ 86.4 (4, s) and δ 85.0 (5, s)]. In contrast to hydroperoxides I and 2, however, compounds 3-5 contain two E-disubstituted and one trisubstituted double bond as well as three methyl groups, of which one is vinylic and two attached to (a) fully substituted carbon atom(s).

These results and a comparison of the 13 C NMR spectra with those of the (1S, 2E, 4S, 6R, 7E, 10E, 12S)-, (1S, 2E, 4S, 6R, 7E, 10E, 12R)- and (1S, 2E, 4R, 6R, 7E, 10E, 12S)-2,7,10-cembratriene-4,6, 12-triols $(10-12)^{1,3}$ led to a tentative formulation of 3 and 4 as (1S, 2E, 4S, 6R, 7E, 10E)-12-hydroperoxy-2,7,10-cembratriene-4,6-diols and of 5 as a $(1S, 2E, 4R, 6R, 7E, 10E, 12\xi)$ -12-hydroperoxy-2,7,10-cembratriene-4,6-diol.

Conclusive evidence of the structures of hydroperoxides I-5 was obtained via biomimetic syntheses. These involved sensitized photo-oxygenation of the 4S,6R- and 4R,6R-diols (6,7) and resulted, in each case, in the formation of a mixture of ene products, which was subjected to repeated HPLC. The three major products of the 4S-series, which were identified as (1S,2E,4S,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol, (1S,2E,4S,6R,7E,10E,12S)- and 1S,2E,4S,6R,77E,10E,12R)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol by reduction to the known triols 8,10 and $11^{1,3}$ using

from hydroperoxides 1, 3 and 4, respectively. The two major photo-oxygenation products of the 4R-series, which were correlated with triols 9 and 12 by reduction, were found to be identical to hydroperoxides 2 and 5, respectively. The latter are hence fully characterized as (1S, 2E, 4R, 6R, 7E, 11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol (2) and (1S, 2E, 4R, 6R, 7E, 10E, 12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-

triethyl phosphite, proved to be indistinguishable

diol (5).

With the structures of these new compounds at hand, it can be inferred from the ¹H and ¹³C NMR results that the conformational difference with respect to the orientation about the 5,6 bond, previously noted between triols of the 4S-and the 4R-series (e.g. 8, 10 and 11 vs. 9 and 12), ¹ is retained among the hydroperoxides.

Discussion. The present results provide experimental support for the view that the 4,6,11-and 4,6,12-triols, which are fairly abundant in the cuticular wax of the tobacco leaf and flower, are formed from the 4,6-diols (6-7) through the intermediacy of hydroperoxides. They do not, however, imply the exclusive operation of singlet oxygen reactions, since enzyme-catalyzed oxidations would also be expected to proceed via hydroperoxide intermediates.

These new compounds (1-5), which are the only hydroperoxides found in tobacco so far,* add to the small but growing group of terpenic hydroperoxides. Among these are neoconcinndiol hydroperoxide from Laurencia snyderiæ,⁴ crispolide from Tanacetum vulgare⁵ as well as 4(15),5,10(14)-germacratriene-1β-hydroperoxide from Senecio glanduloso-pilosus.⁶

Experimental. With the exception of accurate mass measurements, which were carried out on a Kratos MS 50 Stereo DS 55 SM/DS 55 S mass spectrometer-computer system and the NMR spectra, which were recorded on a Varian XL-200 spectrometer, the instruments specified in Ref. 7 were used.

An extract (83 g) obtained by immersing flowers of Greek *Nicotiana tabacum* (Basma Drama) in chloroform, was initially separated into five fractions, A (12.7 g), B (4.7 g), C (8.0 g), D (30 g) and E (3.6 g) by flash chromatography using a column packed with silica gel and gradients of hexane-ethyl acetate-methanol as eluent.

Fraction C was further separated by flash chromatography over silica gel into five fractions C1 (250 mg), C2 (485 mg), C3 (3.0 g), C4 (3.5 g) and C5 (465 mg). Repeated HPLC of fraction C4 using columns packed with Spherisorb and Spherisorb 5 Nitrile led to the isolation of 21 mg

^{*} An analysis by TLC using ferrous thiocyanate as the spraying reagent of an extract, obtained by immersing fresh green leaves of Burley tobacco in cold chloroform (0 °C), revealed the presence of a peroxide-positive zone, whose R_F value agreed with that of hydroperoxides I-5. Moreover, since the 4,6-diols (6,7) fail to produce hydroperoxides even on prolonged exposure to triplet oxygen, it can be concluded that hydroperoxides I-5, now isolated from flowers of Greek tobacco, are not artefacts but genuine tobacco constituents.

of (1S, 2E, 4S, 6R, 7E, 11S)-11-hydroperoxy-2,7, 12(20)-cembratriene-4,6-diol (1), 28 mg of (1S, 2E, 4S, 6R, 7E, 10E, 12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (3) and 5 mg of (1S, 2E, 4S, 6R, 7E, 10E, 12R)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (4).

Fraction C5 was further separated by HPLC using columns packed with Spherisorb and Spherisorb 5 Nitrile to give 7 mg of (1S,2E,4R,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol (2) and 5 mg of (1S,2E,4R,6R,7E,10E,12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (5).

(1S, 2E, 4S, 6R, 7E, 11S)-11-Hydroperoxy-2,7,12 (20)-cembratriene-4,6-diol (1) had m.p. 132-134 °C, $[\alpha]_D$ +53° (c 0.47, CHCl₃) (Found: [M-18]⁺ 320.2337. Calc. for $C_{20}H_{32}O_3$: 320.2352); IR (CHCl₃) bands at: 3603, 3464, 3080, 1385 and 1370 cm⁻¹; ¹H NMR (CDCl₃): δ 0.87 (d, J=6.7 Hz)/0.89 (d, J=6.7 Hz) (H-16/H-17), 1.28 (s, H-18), 1.66 (d, J=1.4 Hz, H-19), 1.77 (dd, J=2.4and -14.4 Hz, H-5a), 2.23 (dd, J=5.6 and -14.7Hz, H-5b), 4.28 (dd, J=4.5 and 9.1 Hz, H-11), 4.66 (ddd, J=2.4, 5.6 and 9.3 Hz, H-6), 5.12 (q,J=1.6 Hz, H-20a), 5.15 (broad s, H-20b), 5.46 (d, J=15.6 Hz, H-3), 5.54 (dd, J=8.3 and 15.6 Hz, H-2), 5.61 (broad d, J=9.3 Hz, H-7) and 8.05 (broad s, OOH); MS [m/z (%)]: 320 (0.2), 302 (0.3), 259 (0.8), 243 (1), 217 (0.9), 203 (1), 135 (8), 121 (13), 109 (16), 95 (20), 81 (20), 69 (21), 55 (25) and 43 (100).

(1S,2E,4R,6R,7E,11S)-11-Hydroperoxy-2,7,12 (20)-cembratriene-4,6-diol (2) was an oil, which had $[a]_D + 34^\circ$ (c 0.54, CHCl₃) (Found: [M-18]⁺ 320.2358. Calc. for C₂₀H₃₂O₃: 320.2352); IR (CHCl₃) bands at: 3601, 3401, 3080, 1384 and 1370 cm⁻¹; ¹H NMR (CDCl₃): δ 0.82 (d, J=6.4 Hz)/0.85 (d, J=6.2 Hz) (H-16/H-17), 1.36 (s, H-18), 1.72 (d, J=1.3 Hz, H-19), 1.96 (dd, J=7.7 and -14.1 Hz, H-5a), 2.05 (dd, J=2.8 and -14.1

Hz, H-5b), 4.30 (t, J=6 Hz, H-11), 4.68 (ddd, J=2.8, 7.7 and 9.0 Hz, H-6), 5.06 (q, J=1.4 Hz, H-20a), 5.13 (broad s, H-20b), 5.28 (broad d, J=9.0 Hz, H-7), 5.37 (dd, J=8.3 and 15.8 Hz, H-2), 5.47 (d, J=15.8 Hz, H-3) and 7.90 (broad s, OOH); MS [m/z (%)]: 320 (0.2), 302 (0.5), 259 (0.8), 243 (1), 215 (1), 203 (2), 135 (8), 121 (14), 109 (16), 95 (24), 81 (24), 69 (24), 55 (27) and 43 (100).

(1S, 2E, 4S, 6R, 7E, 10E, 12S)-12-Hydroperoxy-2,7,10-cembratriene-4,6-diol (3) was an oil, which had $[\alpha]_D$ +91° (c 0.50, CHCl₃) (Found: $[M-18]^{+}$ 320.2345. C₂₀H₃₂O₃: Calc. for 320.2352); IR (CHCl₃) bands at: 3602, 3384, 1664, 1384 and 1369 cm⁻¹; ¹H NMR (CDCl₃): δ 0.82 (d, J=6.7 Hz)/(0.86 (d, J=6.6 Hz) (H-16/H-17), 1.21 (s)/1.37 (s) (H-18/H-20), 1.67 (broad s, H-19), 1.73 (dd, J=1.8 and -14.4 Hz, H-5a), 2.20 (dd, J=6.2 and -14.4 Hz, H-5b), 2.65 (ddd, J=0.6, 7.0 and -17.6 Hz, H-9a), 2.75 (ddd, J=0.6, 7.0 and -17.6 Hz, H-9b), 4.76 (ddd, J=1.8, 6.2 and 8.4 Hz, H-6), 5.39 (dt, J=0.6 and 16.1 Hz, H-11), 5.41 (d, J=15.4 Hz, H-3), 5.56 (broad d, J=8.4 Hz, H-7), 5.57 (dd, J=8.5 and 15.4 Hz, H-2), 5.63 (dt, J=7.0) and 16.1 Hz, H-10) and 8.22 (broad s, OOH); MS [m/z (%)]: 320 (0.1), 302 (0.2), 287 (0.4), 259 (0.4), 243 (0.8), 219 (0.8), 203 (1), 135 (8), 121 (10), 109 (16), 95 (18), 81 (17), 71 (21), 55 (21) and 43 (100)

(1S,2E,4S,6R,7E,10E,12R)-12-Hydroperoxy-2,7,10-cembratriene-4,6-diol (4) was an oil, which had $[\alpha]_D$ +124° (c 0.23, CHCl₃) (Found: [M-36]⁺ 302.2270. Calc. for C₂₀H₃₀O₂: 302.2246); IR (CHCl₃) bands at: 3602, 3472, 1667, 1385 and 1370 cm⁻¹; ¹H NMR (CDCl₃): δ 0.81 (d, J=6.7 Hz)/0.86 (d, J=6.7 Hz) (H-16/H-17), 1.23 (s, H-20), 1.35 (s, H-18), 1.68 (broad s, H-19), 1.74 (dd, J=2.2 and -14.3 Hz, H-5a), 2.20 (dd, J=6.4 and -14.3 Hz, H-5b), 2.75 (dd, J=0.8

and 6.8 Hz, H-9a and H-9b), 4.79 (ddd, J=2.2, 6.4 and 8.6 Hz, H-6), 5.44 (d, J=15.4 Hz, H-3), 5.45 (dt, 0.8 and 16.0 Hz, H-11), 5.48 (broad d, J=8.6 Hz, H-7), 5.55 (dd, J=8.3 and 15.4 Hz, H-2), 5.67 (dt, J=6.8 and 16.0 Hz, H-10) and 7.33 (broad s, OOH); MS [m/z (%)]: 302 (0.1), 287 (0.6), 259 (0.5), 243 (0.8), 217 (1), 203 (1), 135 (5), 121 (9), 109 (18), 95 (18), 81 (17), 69 (18), 55 (20) and 43 (100).

(1S, 2E, 4R, 6R, 7E, 10E, 12S)-12-Hydroperoxy-2,7,10-cembratriene-4,6-diol (5) was an oil, which had $[\alpha]_D$ +64° (c 0.09, CHCl₃) (Found: $[M-18]^{+}$ 320.2343. Calc. C₂₀H₃₂O₃: for 320.2352); IR (CHCl₃) bands at 3602, 3381, 1662, 1385 and 1370 cm⁻¹; ¹H NMR (CDCl₃): δ 0.76 (d, J=6.6 Hz)/0.85 (d, J=6.5 Hz) (H-16/H-17), 1.36 (s)/1.37 (s) (H-18/H-20), 1.79 (d, J=1.3Hz, H-19), 1.83 (dd, J=9.3 and -13.8 Hz, H-5a), 2.05 (dd, J=2.9 and -13.8 Hz, H-5b), 2.61 (dd, J=2.9 and -13.8 Hz, H-5b)J=7.5 and -14.9 Hz, H-9a), 2.83 (dd, J=6.7 and -14.9 Hz, H-9b), 4.69 (ddd, J=2.9, 8.8 and 9.3 Hz, H-6), 5.27 (broad d, J=8.8 Hz, H-7), 5.35 (dd, J=8.3 and 15.8 Hz, H-2), 5.45 (d, J=15.6Hz, H-11), 5.45 (d, J=15.8 Hz, H-3), 5.72 (ddd, J=6.7, 7.5 and 15.6 Hz, H-10) and 7.43 (broad s, OOH); MS [m/z (%)]: 320 (0.1), 302 (0.2), 287 (0.2), 259 (0.4), 243 (0.7), 221 (1), 203 (1), 136 (6), 121 (11), 109 (15), 95 (20), 81 (19), 69 (21), 55 (23) and 43 (100).

Sensitized photo-oxygenation of (1S, 2E, 4S, 6R, 7E,11E)- and (1S,2E,4R,6R,7E,11E)-2,7,11cembratriene-4,6-diol (6, 7). A solution of 1.1 g of 6 and 34 mg of Rose Bengal in 20 ml of methanol, in a tube cooled by a water jacket, was irradiated with a 400 W sodium high pressure lamp placed outside the tube, while oxygen was bubbled through the reaction mixture. After 1.5 h the mixture was taken to dryness and the residue filtered through alumina. Repeated HPLC using a column packed with Spherisorb 5 Nitrile allowed the isolation of 210 mg of (1S, 2E,4S,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol (1), 140 mg of (1S,2E,4S,6R,7E,10E,12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (3) and 12 mg of (1S,2E,4S,6R,7E,10E,12R)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (4).

A solution of 200 mg of 7 and 10 mg of Rose Bengal in 25 ml of methanol was treated with singlet oxygen for 1 h using the apparatus described above. Separation by repeated HPLC using a column packed with Spherisorb 5 Nitrile afforded 80 mg of (1S,2E,4R,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-cembratriene-4,6-diol (2) and 40 mg of (1S,2E,4R,6R,7E,10E,12S)-12-hydroperoxy-2,7,10-cembratriene-4,6-diol (5).

Of these, the (1S,2E,4S,6R,7E,11S)- and (1S, 2E,4R,6R,7E,11S)-11-hydroperoxy-2,7,12(20)-

cembratriene-4,6-diols were identical (optical rotation, IR, ¹H NMR and MS) to compounds *I* and 2 and the (1*S*,2*E*,4*S*,6*R*,7*E*,10*E*,12*S*)-, (1*S*, 2*E*,4*S*,6*R*,7*E*,10*E*,12*R*)- and (1*S*,2*E*,4*R*,6*R*,7*E*, 10*E*,12*S*)-12-hydroperoxy-2,7,10-cembratriene-4,6-diols to compounds 3, 4 and 5, respectively.

Conversion of hydroperoxides 1–5 to corresponding triols (8–12). A solution of 6.9 mg of (1S,2E,4S,6R,7E,11S)-11-hydroperoxy-2,7,12 (20)-cembratriene-4,6-diol (1) and 10 μ l of triethyl phosphite in 1 ml of methanol was kept at room temperature for 5 min. Removal of the solvent and separation by HPLC using a column packed with Spherisorb and hexane—ethyl acetate (20:80) as an eluent afforded 6.0 mg of a product, whose IR, 1 H NMR and mass spectra were identical to those of (1S,2E,4S,6R,7E,11S)-2,7, 12(20)-cembratriene-4,6.11-triol (8).

Using the same method, $(1\hat{S},2\hat{E},4R,6R,7E,11S)$ -11-hydroperoxy-2,7,12(20)-cembratriene-4, 6-diol (2), $(1\hat{S},2\hat{E},4\hat{S},6R,7\hat{E},10\hat{E},12\hat{S})$ -, $(1\hat{S},2\hat{E},4\hat{S},6R,7\hat{E},10\hat{E},12\hat{S})$ -, $(1\hat{S},2\hat{E},4\hat{S},6R,7\hat{E},10\hat{E},12\hat{S})$ -, and $(1\hat{S},2\hat{E},4R,6R,7\hat{E},10\hat{E},12\hat{S})$ -12-hydroperoxy-2,7,10-cembratriene-4,6-diol (3-5) were converted to compounds, which were indistinguishable from $(1\hat{S},2\hat{E},4R,6R,7\hat{E},11\hat{S})$ -2,7,12(20)-cembratriene-4,6,11-triol (9), $(1\hat{S},2\hat{E},4S,6R,7\hat{E},10\hat{E},12\hat{S})$ -, $(1\hat{S},2\hat{E},4S,6R,7\hat{E},10\hat{E},12\hat{S})$ -2,7, 10-cembratriene-4,6,12-triol (10-12), 1,3 respectively.

Acknowledgement. We are grateful to Dr. Petra Ossowski, Mr. Jacek Bielawski and Mr. Ole Homestad for recording the mass spectra.

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Received June 29, 1983.