Tobacco Chemistry. 41. Structure Determination and Synthesis of 5(13), 7E-Megastigmadien-6,9-diol, a New Constituent of Greek Tobacco

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A new C_{13} nor-carotenoid was isolated from Greek *Nicotiana tabacum* L. and shown by spectroscopic methods and synthesis to be 5(13),7E-megastigmadien-6,9-diol.* The additional compounds obtained in the reaction of singlet oxygen with (\pm) - β -ionol were also characterized.

A large number of C_{13} compounds obviously formed by initial oxidative cleavage of carotenoids and subsequent chemical alterations have been isolated from tobacco.² The present paper deals with the structure elucidation and biomimetic synthesis of a new representative of this group of compounds. An account of the additional products obtained in the reaction of singlet oxygen with β -ionol is also given.

RESULTS

The new tobacco constituent, 1, $C_{13}H_{22}O_2$, was isolated in minute quantity by repeated liquid chromatography of a polar, volatile fraction, B 8,³ obtained from an ether extract of sun-cured Greek tobacco. It contained a secondary hydroxyl group as shown by IR absorption at 3600 and 3550-3120 cm⁻¹ and a one-proton quintet at δ 4.39 in the ¹H NMR spectrum. This quintet was converted to a quartet on irradiation at the frequency of an olefinic doublet of doublets at δ 5.85 and to a doublet on irradiation at the frequency of a methyl doublet at δ 1.30. Since, conversely,

Moreover, it was clear from the presence of two three-proton singlets at δ 0.91 and one-proton signals at δ 4.86 and 4.91 in the ¹H NMR spectrum that 1 contains two methyl groups on fully substituted carbon(s) and an exocyclic methylene group (IR band at 1640 cm⁻¹). In view of these results and the elemental composition the nor-carotenoid structure 1 appeared highly probable.

Since the minute quantity available did not allow corroborative ¹³C NMR studies, the proposed structure of I was confirmed by synthesis. This was achieved by subjecting $(\pm) \cdot \beta$ -ionol (2) to photooxygenation sensitized by rose bengal followed by treatment of the reaction mixture with Na₂SO₃. The main product thus obtained, 5(13),7E-megastigmadien-6,9-diol, whose ¹³C NMR spectrum contained diagnostically important peaks at δ 151.31 (s), 108.86 (t) and at 79.25/79.17 (s) due to the >C=CH₂ and |C-OH groups, respectively (cf. Table 1), gave IR, ¹H NMR and mass spectra identical

irradiation at the frequency of the quintet at δ 4.39 converted the doublet of doublets at δ 5.85 to the A part of an AB system, δ _B 6.14, $J_{AB} = 15.5$ Hz, it follows that the new tobacco isolate incorporates partial structure A.

^{*} For nomenclature: see Ref. 1.

with those of the new tobacco constituent (1). This synthetic product (1) is evidently formed in an ene-reaction involving an attack of singlet oxygen on the tetra-substituted 5,6 double bond and loss of a hydrogen from C-13 in (\pm) - β -ionol (2). This attack should occur at either side of the molecule and should furnish two pairs of diastereoisomers, a view corroborated by the fact that the signals ascribed to C-4, and C-6 to C-10 are doublets in the proton noise-decoupled ¹⁸C NMR spectrum of the synthetic sample. The stereochemistry and absolute configuration of the natural product (1) could not be determined due to shortage of material.

In addition to the major product (1) from the reaction of singlet oxygen with (\pm) - β -ionol (2), a few further compounds were isolated and identified. Of these, 4,7-megastig-madien-6,9-diol (3) and 6,7-megastigmadien-5,9-diol (4) can be viewed as products of enereactions, whereas 5,8-epidioxy-6-megastigmen-9-ol (5), dihydroactinidiolide (6) and 5,6-7,8-diepoxymegastigman-9-ol (7) probably arise via a 1,4-addition reaction.

The presence of the 4,5 double bond in 4,7-megastigmadien-6,9-diol (3) was disclosed by the ¹H NMR spectrum, which contained a broad three-proton signal at δ 1.61 and a one-proton multiplet at δ 5.50 corresponding to a methyl group allylically coupled to an olefinic proton. The ¹⁸C NMR spectrum, which in addition to the C-7 and C-8 sp^2 carbon signals displayed resonances due to two sp^2 carbon atoms, a doublet at δ 122.09 and a singlet at δ 136.91, was in accordance with the structure proposed for 3. Moreover, the mass spectrum of 3 had a prominent peak at m/e 154 corresponding to a $C_8H_{14}O_2$ fragment, which arises by retro-Diels-

Fig. 1. The retro-Diels-Alder cleavage of the molecular ion.

Alder cleavage of the molecular ion induced by the 4,5 double bond (Fig. 1).

6,7-Megastigmadien-5,9-diol (4) gave a 13 C NMR spectrum having signals at δ 117.47, 196.66 and 99.59 corresponding to the carbon atoms of the trisubstituted allenic group and an IR spectrum having a characteristic allene band at 1965 cm⁻¹.

5,8-Epidioxy-6-megastigmen-9-ol (5) was isolated as a 1:1 mixture with dihydroactinidiolide (6, vide infra). Its structure was deduced from the ¹⁸C NMR spectrum of this mixture. Thus, subtraction of the signals due to dihydroactinidiolide (6) left thirteen signals to be assigned to 5. Eleven of these had chemical shift values close to those previously published for the C-1 to C-8 and C-16 to C-18 signals for compound 9.4 Since the remaining two signals were due to an oxygen-bearing sp² methine carbon and a methyl carbon it was reasonable to formulate 5 as 5,8-epidioxy-6-megastigmen-9-ol. The mass spectrum having the base peak at m/e 181, which corresponds to an ion formed by a favoured rupture of the 8,9 bond in the molecular ion, provides supporting evidence for the structure proposed.

(±)-Dihydroactinidiolide (6) obtained in a pure state from a different fraction, was identified by spectral comparison with an authentic sample.

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As shown by GC-MS analysis and by the ¹⁸C and ¹H NMR spectra, 5,6-7,8-diepoxymegastigman-9-ol (7) was obtained as a mixture of isomers. The presence of the 7,8-epoxide group adjacent to the secondary hydroxyl group at C-9 was disclosed by the ¹H NMR spectrum of this mixture, which exhibited a one-proton doublet at δ 3.38/3.42 (J=2.1 Hz), a oneproton doublet of doublets at δ 3.04/3.09 (J =2.1 and 3.3 Hz), a one-proton doublet of quartets at $\delta 3.76/3.97$ (J = 3.3 and 6.5 Hz), and a three-proton doublet at δ 1.27/1.30 (J = 6.5Hz). The 18C NMR spectrum not only confirmed the presence of the 7,8-epoxide group, δ 57.26/56.96 (d) and 60.24/59.48 (d), but also suggested that 7 incorporated a tetra-substituted epoxide group, δ 66.10 (s) and 63.80/ 63.59 (s). This must be attached to C-5 and C-6, a conclusion supported by the fact that the ¹H NMR spectrum contained a methyl singlet at δ 1.31/1.36.

Oxidation using Jones' reagent converted the alcohol (7) to 5,6-7,8-diepoxymegastigman-9-one (8), whose IR spectrum had an absorption band at 1710 cm⁻¹ and whose ¹H NMR spectrum contained a three proton singlet at δ 2.09 and an AB quartet centered at δ 3.47 corresponding to the protons of the methyl ketone and 7,8-epoxy groups, respectively. The facts that no doubling of peaks was observed in either the ¹H or the ¹⁸C NMR spectrum of the ketone (8) and that the sample was optically inactive support the view that the mixture of alcohols (7) is derived from the two pairs of diastereomeric 5,8-epidioxy-6-megastigmen-9-ols (5).

Sensitized photooxygenation of β -ionol (2), performed in the presence of catalytic amounts of alkali, has previously been studied by Isoe et al.⁵ They isolated dihydroactinidiolide (6)

as the major product and identified one of the minor components as 6,7-megastigmadien-5,9-diol (4). The latter compound, which has been patented as a flavour additive to tobacco, and the 7Z-derivative of 1 are major products of the photooxidation of cis- β -ionol. Compounds 10-15 have been obtained on photooxygenation of 3-methyl-1-(2,6,6-trimethylcyclohexen-1-yl)-1,3-butadiene (16), a result which is analogous to our findings for the reaction of β -ionol (2) with singlet oxygen.

EXPERIMENTAL

IR spectra were measured on Digilab FTS-14 and Perkin-Elmer 257 instruments and mass spectra on an LKB 2091 instrument. Accurate mass measurements were carried out on an Atlas SM 1 instrument at the Laboratory for Mass Spectrometry, the Karolinska Institute, Stockholm. Fourier transform ¹H NMR (100 MHz) and ¹³C NMR (25.16 MHz) spectra were obtained on a Varian XL-100-12 spectrometer equipped with S-124 FT and disk accessories and controlled by a Varian 620/L computer. Gas chromatography was performed on a Varian 1700 instrument using glass capillary columns (50 m \times 0.37 mm) coated with HB 5100. High performance liquid chromatography was carried out using a Waters 6000 A solvent delivery system, a U6K injector and an R-401 differential refractometer.

Isolation of 5(13), 7E-megastigmadien-6,9-diol (1) from tobacco. A volatile, neutral fraction (B8) of an extract obtained from 295 kg of sun-cured Greek Nicotiana tabacum L. was chromatographed over silica gel using a light petrol/diethyl ether gradient. One of the subfractions obtained was purified further by liquid chromatography using columns packed with Bondapak C₁₈/Porasil (Waters), μ-Bondapak C₁₈ (Waters) and μ-Porasil (Waters) to afford 3 mg of 1 as a colourless oil (Found: M 210.1620. Calc. for C₁₃H₂₂O₂: 210.1620), which had IR (CHCl₃) bands at 3600 (m), 3550-3120 (w) and 1640 (w) cm⁻¹; ¹H NMR (CDCl₃): δ

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0.91 (6 H, s), 0.93 (1 H, s), 1.30 (3 H, d, J=6Hz), 4.39 (1 H, quintet J = 6 Hz), 4.86 (1 H, m), 4.91 (1 H, m), 5.85 (1 H, dd, J = 15.5 and 6 Hz) and 6.14 (1 H, broad d, J = 15.5 Hz) (AB part of an ABX system); MS [m/e (com-(AB part of an ABA system; and [m/e] (composition, %)]: 210 ($C_{13}H_{22}O_2$, 2), 192 ($C_{13}H_{20}O_3$, 4), 177 ($C_{12}H_{17}O_3$, 3), 165 ($C_{11}H_{17}O_3$, 5), 152 ($C_{10}H_{16}O_3$, 68), 141 ($C_{5}H_{13}O_2$, 30), 123 ($C_{9}H_{16}$ and $C_{8}H_{11}O_3$, 22), 109 ($C_{8}H_{13}$ and $C_{7}H_{9}O_3$, 42), 97 ($C_{7}H_{13}$ and $C_{6}H_{9}O_3$, 38), 96 ($C_{7}H_{12}$ and $C_{6}H_{8}O_3$, 27), 95 ($C_{7}H_{11}$ and $C_{6}H_{7}O_3$, 43), 81 (29), and 43 (100) 69 (33), 55 (28) and 43 (100).

Photoxidation of β -ionol. A solution of 1.0 g of (\pm) - β -ionol and 0.1 g of rose bengal in 25 ml of methanol in a tube cooled by a water jacket was irradiated for 1 h with a 400 W sodium high pressure lamp placed outside the tube while oxygen was bubbled through the reaction mixture. A solution of 1.0 g of Na₂SO₃ in 20 ml of H₂O was added and the reaction mixture was stirred at room temperature for 17 h. Dilution with water, extraction with ether and chromatography on silica gel using a hexane/ethyl acetate gradient eluted in order compounds 5, 6, 7, 1, 3, and 4. Further purifica-tion was achieved by high performance liquid chromatography on columns packed with u-Porasil (Waters) using hexane and ethyl acetate

5(13),7E-megastigmadien-6,9-diol (1, 134 mg) gave IR, 'H NMR and mass spectra identical

to those of the natural product.

4,7-Megastigmadien-6,9-diol (3, 9 mg), which was recrystallized from diethyl ether – hexane (1:1) to give white needles, m.p. 108 - 109 °C. (Found: $[M - H_2O]^+$ 192.1518. Calc. for $C_{13}H_{20}O$: 192.1514). IR bands at 3600 (s), and 3550 – 3100 (m) cm⁻¹; ¹H NMR (CDCl₃): δ 0.90 (3 H, s), 0.98 (3 H, s), 1.30 (3 H, d, J = 6 Hz), 1.61 (3 H, m), 4.39 (1 H, m, $W_{1/2} = 14$ Hz), 5.50 (1 H, m, $W_{1/2} = 10$ Hz) and 5.6 – 5.8 (2 H, overlapping signals); MS [m/e (composition, %)]: The strain of t

6.7-Megastigmadien-5,9-diol (4, 5 mg), which was recrystallized from ethyl acetate to give white needles, m.p. 140-143 °C. (Found: $[M-H_2O]^+$ 192.1512. Calc. for $C_{13}H_{20}O$: 192.1514). IR (KBr) bands at 3650-3040 (s) and 1965 (m) cm⁻¹; ¹H NMR (CDCl₃): δ 1.06 (3 H, s), 1.26 (3 H, s), 1.32 (3 H, d, J =6 Hz), 1.36 (3 H, s), 4.76 (1 H, quintet, J=6 Hz) and 5.45 (1 H, d, J=6 Hz); MS [m/e] (com-12) and 3-13 (1 11, d, 9 = 0 112); his [m/e (colliposition, %)]: 210 (M, 1), 192 (C₁₃H₂₀O, 10), 177 (C₁₄H₁₇O, 28), 149 (C₁₁H₁₇ and C₁₀H₁₃O, 17), 133 (C₁₀H₁₃, 30), 119 (C₈H₁₁, 23), 107 (C₈H₁₁ and C₇H₇O, 100), 85 (24) and 43 (88). Acetylation of 4 with acetic anhydride and pyridine at room temperature for 30 min afforded 9-acetoxy-6,7-megastigmadien-5-ol as a colourless oil, which had IR (CHCl₃) bands at 3580 (m), 3560-3300 (m), 1955 (m) and

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Com- pound	C-1	G-2	C-3	C-4	G-5	G-6	C-7	C-8	C-9	C-10	C-11/C-12	C-13
I	39.14	37.24	22.66	33.04 32.98	161.31	79.25	131.39	134.21	68.59	23.64	24.16/22.88	108.86
2,	36.61	33.10	22.47	122.09	131.91	76.76	131.33	134.07	66.61	22.38	24.27/24.52	18.65
4 2	33.29	40.26	17.96	40.48	68.73	117.47	196.66	99.59	64.08	23.88	$31.87^d/30.83^d$	28.90 4
0 9	30.13	40.73	18.70	30.65	79.67	149.00	115.01	82.67	70.21	19.72	$25.49^{c}/30.48$ c	27.77 6
0 ;	30.32	40.07	19.03	41.62	86.80	182.26	112.18	172.23			24.05/29.70	24.30
	33.41	38.43	17.10	32.02	63.80	66.10	57.26	60.24	66.04	20.01	25.27/26.79	20.01
	33.36	38.37		31.90	63.29		56.96	59.48	65.82	19.40	-	
&	33.39	38.17	17.03	31.80	63.92	65.66	58.94 0	57.92°	*~	24.86	25.02/26.78	19.69
a ô-Values relative to TMS between the C-6 and C-8 sign was assigned to C-8 and the	a ô-Values relative to TMS; tween the C-6 and C-8 signs as assigned to C-8 and the	-:•_ag as	spectrum	recorded in e with the 182.26, brosa	CDCl ₃ . ^b S _l and of the dened due	oectrum rec proton-coup to several th	orded in DM oled 13C NMF oree bond C-	SO-d _e . c,d A spectrum; H couplings,	ssignment the sharp was assig	may be r doublet (2 ned to C-	; spectrum recorded in CDCl ₃ . ^b Spectrum recorded in DMSO- $d_{\rm e}$. $c_{\rm e}d$ Assignment may be reversed. ^e The distinction als was made with the aid of the proton-coupled ¹³ C NMR spectrum; the sharp doublet (2 JCH = 8.3 Hz) at 5 171.23 signal at 6 182.26, broadened due to several three bond C-H couplings, was assigned to C-6. 7 Not visible due to low	stinction δ 171.23 e to low

sample concentration

1725 cm⁻¹; ¹H NMR (CDCl₃): δ 1.04/1.05 (3 H, s), 1.22/1.24 (3 H, s), 1.32/1.34 (3 H, s), 1.37 (3 H, d, J=6 Hz), 2.04/2.05 (3 H, s) and 5.15 – 5.55 (2 H, overlapping signals). These data are somewhat at variance with previously published values.⁵

5,8-Epidioxy-6-megastigmen-9-ol (5, 7 mg), was present as a 1:1 mixture with dihydroactinidiolide (6). Subtraction of the peaks due to 6 from the 'H NMR spectrum (CDCl₃) of the mixture left the following signals for 5: δ 1.12 (3 H, s), 1.17 (3 H, s), 1.26 (3 H, d, J=6 Hz), 1.61 (3 H, s), 4.15 (2 H, overlapping signals) and 5.67 (1 H, d, J=3 Hz); MS [m/e (%)]: 208 (M-18, 1), 193 (2), 181 (100), 163 (13), 135 (22), 121 (18), 107 (18), 95 (18), 81 (11), 69 (17), 55 (13) and 43 (58).

(±)-Dihydroactinidiolide (6, 103 mg) obtained in a pure form from a more polar fraction, was identified by comparison of its IR, ¹H NMR and mass spectra with those of an

authentic sample.

5,6-7,8-Diepoxymegastigman-9-ol (7, 25 mg), which was recrystallized from hexane to give white needles, m.p. $91-93^{\circ}$ C. (Found: [M-45]+ 181.1221. Calc. for $C_{11}H_{17}O_{2}$: 181.1228. 14 NMR (CDCl₃): δ 1.09 (3 H, s), 1.27/1.30 (3 H, d, J=6.5 Hz), 1.28 (3 H, s), 1.31/1.36 (3 H, s), 3.04/3.09 (1 H, dd, J=3.3 and 2.1 Hz), 3.38/3.42 (1 H, d, J=2.1 Hz), 3.76/3.97 (1 H, dq, J=6.5 and 3.3 Hz); MS [m/e (composition, 9/e)]: 208 (M-18, 1), 193 (1), 181 ($C_{11}H_{17}O_{2}$, 13), 163 ($C_{11}H_{15}O$, 10), 145 ($C_{11}H_{18}$, 6), 123 ($C_{9}H_{15}$ and $C_{8}H_{10}O$, 39), 121 ($C_{9}H_{15}$ and $C_{8}H_{9}O$, 29), 109 ($C_{8}H_{13}$ and $C_{7}H_{9}O$, 17), 105 ($C_{8}H_{9}$, 15), 95 ($C_{7}H_{11}$ and $C_{8}H_{7}O$, 22), 85 ($C_{5}H_{9}O$ and $C_{4}H_{5}O_{2}$, 29), 71 (15), 69 (18), 67 (12), 55 (17) and 43 (100). Treatment of 7 with Jones' reagent for 1 h at room temperature afforded 5.6-7.8-diepoxymegastigman-9-one (8) which had an IR band at 1710 cm⁻¹; ^{1}H NMR (CDCl₃): δ 1.07 (3 H, s), 1.30 (3 H, s), 1.31 (3 H, s), 2.09 (3 H, s), 3.43 (1 H, d, J=2 Hz) and 3.51 (1 H, d, J=2 Hz); MS [m/e (9/e)]: 206 (M-18, 1), 181 (12), 163 (10), 145 (3), 137 (6), 123 (29), 121 (17), 105 (15), 95 (21), 85 (27), 69 (16), 55 (17) and 43 (100).

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