Chemical Studies on Bryophytes. 17. A New Luteolin Tetraglycoside from *Hedwigia ciliata*

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The moss H. ciliata contains twelve flavonoid compounds. One of the flavonoids has been assigned the structure luteolin-7,4'-di-O-(2-O- α -L-rhamnopyranosyl- β -D-glucopyranoside) (1) by a combination of spectroscopic methods and a sugar sequence analysis by GLC-MS of the methylated alditol acetates.

In this series of chemical studies of liverworts and mosses, the common moss *Hedwigia ciliata* was investigated. This communication describes the isolation and separation of the flavonoids from this moss, and the structural determination of one luteolin tetraglycoside. The structures of the other flavonoids will be reported later.

The ethanolic extract from 4.8 kg of the moss *H. ciliata* after gel filtration, paper and column chromatography, contains twelve flavonoid compounds (Table 1). Four of these flavonoids (1, 3, 10, and 11) were obtained in larger quantities than the others. The yields of these main components were each larger than 0.5 g. One of the minor flavonoids (12) is probably identical with the biluteolin isolated from *Dicranum scoparium*.¹

One of the main components (1) was purified by gel filtration. By R_F values and UV spectral studies of the products obtained after acid hydrolysis of 1 the aglycone was identified as luteolin and the sugars as glucose and rhamnose. The sugars are linked to luteolin at the 7- and the 4'-position according to UV spectral data of 1 using diagnostic shift reagents.²

Partial hydrolysis of 1 gave two intermediate luteolin glycosides, showing that 1 contains at least three monosaccaride molecules. The

Table 1. R_F values of the isolated flavonoids from $H.\ ciliata$ on 0.1 mm pre-coated cellulose TLC plates.

Com-	Solvent			
pound	TBA	BAW	15 % HOAc	
1	0.09	0.15	0.75	
2	0.04	0.07	0.66	
3	0.04	0.07	0.63	
4	0.16	0.28	0.62	
5	0.11	0.24	0.52	
6	0.10	0.24	0.45	
7	0.13	0.20	0.34	
8	0.54	0.50	0.32	
9	0.30	0.37	0.18	
10	0.15	0.22	0.40	
11	0.07	0.11	0.25	
12	0.92	0.94	0.02	

osmometrically determined molecular weight of 1 (mol. wt. 885) indicates that 1 is probably a tetraglycoside of luteolin. Only one luteolin glycoside with more than two monosaccaride molecules has been reported earlier. The latter is a luteolin-triglycoside with sugars attached in the 7- and the 4'-positions.

The ¹H NMR spectrum of the TMS ether of I (Fig. 1) shows four sugar C-1 proton signals confirming that I is a tetraglycoside. The two doublets at 5.14 and 5.45 must be due to two glucosyl C-1 protons, since the coupling constants of about 7 Hz agree with a diaxial coupling between the C-1 and the C-2 protons in a β -linked glucose. 2,5 The two signals at 4.76 and 4.86 correspond to two rhamnosyl C-1 protons. 2 Since the two C-1 proton signals at the lowest field are due to glucose, this

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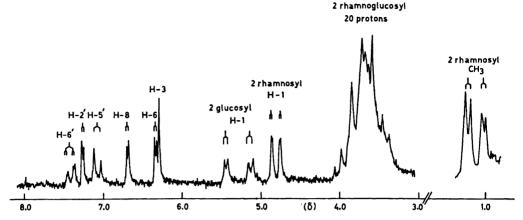


Fig. 1. ¹H NMR spectrum of the TMS ether of luteolin-7,4'-di-O-(2-O-α-L-rhamnopyranosyl- β -D-glucopyranoside).

indicates that I has two β -glucose molecules eties in flavonoids.¹¹⁻¹³ The luteolin tetraglyco-attached to luteolin and that the two rhamnose units are linked to the glucose. Treatment of I methylated sugars reduced and acetylated to with β -glucosidase (emulsin) gave no hydrolysis, showing that there are no terminal β -glucose Results obtained by GLC-MS analysis of the units.

Mass spectroscopy of the permethylated 1 gave no molecular peak and only a weak peak for the aglycone (m/e 314). A base peak at m/e 189 and peaks at m/e 157 (189-CH₃OH) and 125 (157-CH₃OH) indicated a terminal rhamnose unit. No fragments from a terminal glucose could be detected. The peak at m/e 393 indicated a rhamnoglucoside, and the peak at m/e 361 that the rhamnose unit is probably linked to the glucose at the 2-position.

To establish the sugar sequence, a method used for the analysis of polysaccharides was applied.⁷⁻¹⁰ This method has also been used for structural determinations of the sugar moi-

side was permethylated, hydrolyzed and the methylated sugars reduced and acetylated to give partially methylated alditol acetates. Results obtained by GLC-MS analysis of the alditol acetate mixture are given in Table 2. The ratio of the two GLC-peak areas was 1:1 proving that, after methylation and hydrolysis, there are only two different sugar units. The peak with a T value of 0.45 corresponds 1,5-di-O-acetyl-2,3,4-tri-O-methyl-L-rhamnitol showing that the rhamnose units are terminal. The peak with a T value of 1.97 corresponds to both 1,2,5-tri-O-acetyl-3,4,6tri-O-methyl-D-glucitol and 1,3,5-tri-O-acetyl-2,4,6-tri-O-methyl-D-glucitol, but the MS data eliminate the second alternative.10 This result shows that the two rhamnose units are linked to the two glucose units at the 2-position.

The aglycone isolated after permethylation

Table 2. GLC-MS analysis data of the methylated alditol acetates. T values are relative to 1,5-di-O-acetyl-2,3,4,6-tetra-O-methyl-D-glucitol.

Alditol acetate	T	T 10	Prominent fragments m/e
1,5-di-O-acetyl-2,3,4- tri-O-methyl-1,-rhamnitol	0.45	0.46	175, 161, 131, 117, 115, 101, 89, 72, 45, 43
1,2,5-tri-O-acetyl-3,4,6- tri-O-methyl-D-glucitol	1.97	1.98	205, 189, 161, 145, 129, 101, 99, 87, 71, 45, 43

and hydrolysis was identified as 5,3'-di-Omethylluteolin by means of MS and UV spectral data. Therefore 1 is a luteolin-7,4'-di-O-(2-Orhamnopyranosyl-glucopyranoside). The optical rotation of synthetic luteolin-7-O-(2-O-α-Lrhamnopyranosyl- β -D-glucopyranoside is $[\alpha]_D^{22}$ -97.55° (pyridine).14 To explain the large negative rotation, $[\alpha]_D^{26} - 192^{\circ}$ (pyridine), of the isolated luteolin 7,4'-di-O-rhamnoglucoside, 1 must contain two units of \$\beta\$-D-glucose and two units of α-L-rhamnose. Taking these data together, the structure of 1 is proposed to be luteolin-7,4'-di-O-(2-O- α -L-rhamnopyranosyl- β -D-glucopyranoside) (1).

EXPERIMENTAL

NMR spectra were recorded on a Varian A 60-D instrument, UV-visible spectra on a Bausch & Lomb Spectronic 505 spectrophotometer and on a Zeiss spektralphotometer DMR 10. MS were recorded on a LKB 9000 and an AEI MS 30 instrument. GLC were performed on a Perkin Elmer 990 Gas Chromatograph as described earlier. Gel filtrations were performed on Sephadex G-25 columns with EtOH-H₂O (1:1) as eluent. Molecular weight determination was made with a Knauer vapour pressure osmometer. Solvent systems: BuOH – HOAc – H₂O, 6:1:2 (BAW), t-BuOH – HOAc – H₂O, 3:1:1 (TBA).

Isolation. The moss H. ciliata was collected in the surroundings of Enköping and Uppsala. Foreign materials were carefully removed and the air-dried moss (4.8 kg) was pulverized and first extracted with CHCl₃, air-dried and then extracted with 80 % aqueous EtOH at room temperature. The ethanolic extract was evaporated in vacuum at room temperature and the residue suspended in H₂O and continually extracted with ether. The water phase was concentrated to a small volume and, after gel filtration, gave four flavonoid fractions. The first fraction consisted of nine flavone glyco-

sides (1-9), the second and third fractions gave two flavone C-glycosides (10 and 11) and the fourth fraction gave one biflavone (12). PC on Whatman 3 MM paper with BAW and CC on cellulose with 5 % HOAc of the first fraction led to complete separation of the nine flavone glycosides. R_F values, see Table 1.

Luteolin-7,4'-di-O-rhamnoglucoside (1). The crude fraction 1 was further purified by gel filtration. Drying at 100 °C/0.1 mmHg over filtration. Drying at 100 °C/0.1 mmHg over P_2O_5 gave 0.5 g of a pale yellow compound, m.p. > 360 °C, $[\alpha]_D^{26} - 192^\circ$ (c 0.27, pyridine). Found: C 48.24; H 5.95. Calc. for $C_{39}H_{50}O_{24}$: C 48.05; H 6.00. Mol. weight, obs. 885, calc. for $C_{39}H_{50}O_{24}$: 902.8. UV (99.9 % CH₃OH): 272, 340; (+AlCl₃): 279, 295sh, 354, 380sh; (+AlCl₃/HCl): 281, 293sh, 352, 379sh; (+McON₂): 270, 271, (+McOA₂): 271, 338. (+MeONa): 270, 371; (+NaOAc): 271, 338; (+NaOAc/H₂BO₃): 271, 342 nm.

Acid hydrolysis of 1 with 6 % HCl at 100 °C gave luteolin, glucose and rhamnose. Luteolin was identified by MS data, R_F values and UV data. Glucose and rhamnose were identified by PC and TLC by comparison with authentic

Partial hydrolysis of 1 with 10 % HCl at room temperature for 140 h gave two intermediates; 1a and 1b. 1a gave on complete hydrolysis luteolin, glucose and rhamnose, partial hydrolysis with 10 % HCl at room temperature gave 1b. 1b gave on complete hydrolysis luteolin, rhamnose and glucose. nyurolysis intenin, rnamnose and glucose. 1a: R_F values 0.48 (15 % HOAc) and 0.15 (TBA). UV (99.9 % CH₃OH): 269, 336; (+AlCl₃): 279, 294sh, 351, 392sh; (+MeONa): 276, 380; (+NaOAc): 269, 313sh, 330; (+NaOAc/H₃BO₃): 271, 340 nm. This indicated a luterial of charging of 21, 415 for H3b03). This indicated a factor of the control of glycoside.

The acetate was prepared with Ac₂O in pyridine. M.P. 144-146°C, Found: C 52.13; H 5.53. Calc. for C₆₇H₈₄O₄₁: C 52.07; H 5.48. ¹H NMR (100 MHz, CDCl₃): 5 7.73 (H6'), 7.64 (H2'), 7.31 (H5'), 7.03 (H8), 6.72 (H6), 6.58 (H3), 5.4-4.9 (sugar H), 4.3-3.7 (sugar H), 2.41 (aromatic acetyl H), 2.25-1.85 (sugar acetyl H), 1.22 and 1.12 (rhamnose CH₃).

The TMS ether was prepared according to standard procedures.² ¹H NMR (100 MHz, CCl₄): δ 7.42 (H6'), 7.26 (H2'), 7.08 (H5'), 6.70 (H8), 6.35 (H6), 6.30 (H3), 5.45 (glucose H1, J 6 Hz), 5.14 (glucose H1, J 7 Hz), 4.86 (rhamnose H1, J 2 Hz), 4.76 (rhamnose H1, J 2 Hz), 4.10-3.30 (sugar H), 1.22 and 1.02(rhamnose CH₃).

The permethyl ether was prepared with NaH, DMSO and CH₃I using Hakomori's procedure. 15 The permethyl ether was purified by CC on silica gel with acetone as eluent. 1H NMR

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(100 MHz, CDCl₃): δ 7.44 (H6'), 7.32 (H2'), 7.10 (H5'), 6.70 (H8), 6.60 (H3), 6.53 (H6), 5.42 (glucose H1), 5.33 (glucose H1), 5.00 (rhamnose H1), 4.93 (rhamnose H1), 4.0-3.0(sugar H and OCH₂), 1.33 and 1.25 (rhamnose CH₃). MS [70 eV; m/e (% rel. int.)]: 394 (0.5), 393 (1), 391 (0.6), 390 (0.4), 379 (0.4), 363 (0.3), 362 (1), 361 (5), 360 (4), 347 (0.7), 346 (0.4), 332 (0.4), 331 (0.3), 329 (0.3), 328 (0.4), 314 (0.3), 204 (13), 190 (11), 189 (100), 188 (23), 187 (7), 175 (8), 173 (5), 157 (33), 145 (13), (3), 131 (6), 129 (10), 125 (8), 113 (7), 101 (35), 99 (33), 89 (10), 88 (22), 75 (18), 71 (10), 59 (20). Only peaks larger than 5 % (0.3 % m/e 300 - 500) of the base peak are given.

Sequence analysis of sugar. The permethylated glycoside was hydrolyzed with 8 % H_2SO_4 at 100 °C for 3 h. 5,3 Di-O-methylluteolin deposited on cooling. The acid solution was neutralized with the anion exchanger IRA-400 in carbonate form and concentrated to 4 ml in vacuum at room temperature. Reduction with NaBH, and acetylation with Ac2O in pyridine was performed as described earlier.7 Retention times and prominent fragments in GLC-MS analysis are given in Table 2.

5,3'-Di-O-methylluteolin. M.p. 273-276 °C. 37. -17-0-methylutteothn. M.p. 273-276 °C. UV (99.9 % CH₃OH): 241, 265, 337; (+AlCl₃): 241, 265, 337, 412; (+AlCl₃/HCl): 241, 265, 301, 340, 414; (+MeONa): 261, 270sh, 317sh, 392; (+NaOAc): 263sh, 270, 313, 384; (+NaOAc/H₃BO₃): 264, 337 nm. MS [70 eV; m/e; (% rel. int.)]: 315 (21), 314 (100), 313 (50), 300 (16), 298 (10), 297 (14), 286 (6), 285 (31), 284 (26), 283 (14), 270 (11), 269 (9), 268 (36), 256 (6), 255 (6), 253 (6), 242 (5), 241 (6), 238 (5), 213 (6), 167 (4), 151 (4), 149 (4), 148 (19), 268 (10) 143 (9), 138 (4), 137 (21), 136 (5), 133 (14), 128 (6), 123 (4), 118 (4), 108 (5), 105 (10). Only peaks larger than 4 % of the base peak are given.

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REFERENCES

1. Lindberg, G., Österdahl, B.-G. and Nils-

son, E. *Chem. Scr.* 5 (1974) 140. Mabry, T. J., Markham, K. R. and Thomas, M. B. The Systematic Identification of Flavonoids, Springer, Berlin 1970.

- 3. Harborne, J. B., Mabry, T. J. and Mabry, H. The Flavonoids, Chapman & Hall, London 1975, Chapter 8.
- Dranik, L. I. and Chernobai, V. T. Khim. Prirodn. Soedin. 2 (1966) 16; see Chem. Abstr. 65 (1966) 3947f.
- 5. Kamerling, J. P., De Bie, M. J. A. and Vliegenthart, J. F. G. Tetrahedron 28 (1972)
- 6. Schmid, R. D. Tetrahedron 28 (1972) 3259:
- 7. Björndal, H., Lindberg, B. and Svensson, S. Acta Chem. Scand. 21 (1967) 1801.
- 8. Björndal, H., Lindberg, B. and Svensson, S. Carbohydr. Res. 5 (1967) 433.
 9. Hellerqvist, C. G., Lindberg, B. and Svens-
- son, S. Carbohydr. Res. 8 (1968) 43.
- 10. Björndal, H., Hellerqvist, C. G., Lindberg, B. and Svensson, S. Angew. Chem. 16 (1970)
- 11. Nilsson, E., Lindberg, G. and Österdahl, B.-G. Chem. Scr. 4 (1973) 66.
- Schmid, R. D., Varenne, P. and Paris, R. Tetrahedron 28 (1972) 5037.
 Schmid, R. D. and Harborne, J. B. Phy-
- tochemistry 12 (1973) 2269. 14. Inonye, H., Aoki, Y., Wagner, H., Hör-
- hammer, L., Aurnhammer, G. and Budweg, W. Chem. Ber. 102 (1969) 3009.
- 15. Hakomori, S. J. Biochem. (Tokyo) 55 (1964) 205.

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