

Studies on Orchidaceae Glycosides. 2.* The Structures of Loroglossine and Militarine, two Glucosides from *Orchis militaris* L.

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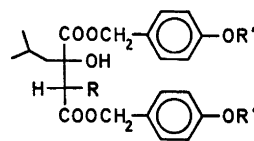
Two glucosides, loroglossine (I) and militarine (II), have been isolated from *Orchis militaris* L. Loroglossine (I) is shown to be bis[4-(β -D-glucopyranosyloxy)benzyl] *erythro*-isobutyltartrate and militarine (II) bis[4-(β -D-glucopyranosyloxy)benzyl] (*R*)-2-isobutylmalate. Syntheses of the (\pm)-*erythro* and (\pm)-*threo* isomers of dimethyl isobutyltartrate are described.

In 1919 Bourquelot and Bridel² isolated a glucoside from *Loroglossum hircinum* (L.) L.C. Rich. for which they proposed the name loroglossine. This glucoside was later found in *Orchis militaris* L. and twenty-one other Orchidaceae species.³⁻⁷

In this communication we report the structure of loroglossine (I) and another glucoside, for which we propose the name militarine (II), both of which were isolated from *O. militaris*.

The two glucosides, which differ by only one hydroxyl group, have the molecular weights 742 and 726, respectively. In spite of the small differences in size and structure, I and II were separated by gel permeation on Sephadex G-15.

Loroglossine (I). On the basis of elemental analysis and molecular weight determinations, Delauney⁸ suggested the molecular formula $C_{30}H_{42}O_{18}$ for loroglossine (I). Later Karrer and Matter⁹ prepared some derivatives of I and from the results of the elemental analysis of these, they considered loroglossine (I) to be a mono-glucoside with the composition $C_{14}H_{20}O_8$.



I: R=OH, R'= β -D-glucopyranosyl

II: R=H, R'= β -D-glucopyranosyl

In the present study, spectrochemical and elemental analyses indicated that loroglossine (I) has the molecular formula $C_{34}H_{46}O_{18}$. Sugar and methylation analyses¹⁰ showed I to be a glucopyranoside. Catalytic hydrogenation of I produced *p*-cresyl β -D-glucopyranoside¹¹ and isobutyltartric acid. As there are no signals due to aromatic methyl groups present in the NMR spectrum of I, loroglossine (I) must have suffered hydrogenolysis during the reduction. This evidence together with the NMR spectrum of I, which shows *inter alia* eight aromatic protons, indicates the presence of two 4-(β -D-glucopyranosyloxy)benzyl residues in loroglossine (I). Since loroglossine (I) is a neutral compound it must be a diester and hence have the structure I.

The relative configuration of the isobutyltartric acid was established by synthesis. Hydroxylation of isobutylfumaric acid¹² and isobutylmaleic anhydride¹² with osmium tetroxide gave, after methylation (\pm)-*threo* and (\pm)-*erythro* dimethyl isobutyltartrate, respectively. The *erythro* isomer was found to be identical, except for the optical rotation, with the dimethyl ester of the natural acid.

* For paper 1 in this series, see Ref. 1.

Attempts to determine the absolute configuration of the isobutyltartaric acid are in progress.

Militarine (II). The molecular formula for militarine (II) was found to be $C_{34}H_{46}O_{17}$, i.e. one oxygen atom less than loroglossine (I). Catalytic hydrogenation of II gave *p*-cresyl β -D-glucopyranoside¹¹ and (*R*)-2-isobutylmalic acid.^{13,14} From the spectral similarities of loroglossine (I) and militarine, it is evident that the latter has the structure II.

p-Hydroxybenzyl alcohol has been isolated from *O. militaris* infected by the fungus *Rhizoctonia repens* Bern.¹⁵⁻¹⁷ Since this alcohol is normally absent from the plant,¹⁵⁻¹⁷ it seems probable that it has been released from loroglossine (I) and/or militarine (II) by enzymatic hydrolysis during the period of infection.

EXPERIMENTAL

All melting points are corrected. Mass spectra were measured on a Perkin-Elmer 270 or a Varian MAT 311 spectrometer, optical rotations on a Perkin-Elmer 141 polarimeter, IR spectra on a Perkin-Elmer 257 instrument, UV spectra on a Beckman DK2 instrument, and NMR spectra on a Varian XL 100 spectrometer. Concentrations were performed under diminished pressure, at bath temperatures not exceeding 40°C. Elemental analyses were carried out at Alfred Bernhardt, Mikroanalytisches Laboratorium, Elbach über Engelskirchen, Germany and Mikroanalytlaboratoriet Lantbrukshögskolan, Uppsala, Sweden.

Isolation and characterisation of I and II. Fresh plants of *O. militaris* (5.5 kg) were extracted with methanol (15 l). The filtrate was concentrated to 0.7 l, washed with chloroform (3 × 0.25 l) and extracted with butanol (13 × 0.15 l). The butanolic phase was washed with water (0.15 l) and evaporated to dryness. Half of the residue (14 g) was filtered through a column of Sephadex LH 20 (8 × 60 cm) with water-ethanol (1:1, 1 ml/min) as eluent. The fraction (4.6 g) containing the glucosides was chromatographed on the same column giving a mixture of I and II (3.4 g). Crystallisation from water gave I (0.9 g) as needles, m.p. 153–157°C (decomp). $[\alpha]_D^{25} - 34^\circ$ (c 0.36, methanol). (Lit.⁸ m.p. 149–150°C, $[\alpha]_D - 36.28^\circ$, methanol. Lit.⁹ m.p. 149–151°C). (Found: C 55.1; H 6.36; O 38.7. Calc. for $C_{34}H_{46}O_{15}$: C 55.0; H 6.24; O 38.8). IR: ν_{\max} (KBr) 3600–3000(s), 1735(s), 1615(m), 1515(s) cm^{-1} . NMR (pyridine- d_5): δ 0.85 (d, 3 H, *J* 6 Hz), 0.97 (d, 3 H, *J* 6 Hz), 1.8–2.2 (m, 2 H), 2.2–2.5 (m, 1 H), 3.8–4.5 (m, 12 H), 4.98–5.16 (m, 4 H), 5.21 (m, 2 H), 5.6–6.8 (broad s, 10 H), 7.25 (m, 8 H).

UV, nm (ϵ): λ_{\max} (methanol) 277 (1340), 270.5 (1590), 223.5 (25 400).

Part of the mother liquor (100 mg) from the crystallisation of I was chromatographed on Sephadex G 15 (1.6 × 70 cm) with water as eluent, giving I (50 mg) and a mixture of I and II (20 mg). Further chromatography in the same system, followed by evaporation gave pure II as a colourless amorphous solid, $[\alpha]_{578}^{25} - 47^\circ$ (c 0.78, methanol). (Found: C 56.2; H 6.60; O 37.4. Calc. for $C_{34}H_{46}O_{17}$: C 56.2; H 6.38; O 37.4). IR: ν_{\max} (KBr) 3700–3000(s), 1735(s), 1615(m), 1515(s) cm^{-1} . NMR (pyridine- d_5): δ 0.90 (d, 3 H, *J* 6 Hz), 0.97 (d, 3 H, *J* 6 Hz), 1.7–1.94 (m, 2 H), 1.94–2.3 (m, 1 H), 2.99 (d, 1 H, *J* 15 Hz) and 3.19 (d, 1 H, *J* 15 Hz, AB system), 3.9–4.6 (m, 12 H), 5.08 (s, 2 H), 5.17 (s, 2 H), 5.38–5.64 (m, 2 H), 5.8–7.2 (broad s, 9 H), 7.22–7.45 (m, 8 H). UV, nm (ϵ): λ_{\max} (methanol) 277 (1430), 270.5 (1730), 223 (24 900).

Hydrogenation of I and II. A solution of I (140 mg) in methanol (10 ml) was hydrogenated over palladium (20 mg, 10% on carbon) at room temperature and atmospheric pressure. After 2.5 h the catalyst was filtered off and the solution was evaporated to dryness. The residue was purified by preparative TLC on silica gel using chloroform-methanol (1:1) as eluent. Crystallisation from a mixture of propan-2-ol and water (3:1) gave *p*-cresyl β -D-glucoside (67 mg) m.p. 178–181°C (Lit.¹¹ m.p. 178–179.5°C), $[\alpha]_D^{25} - 66.5^\circ$ (c 2.26, water) (Lit.¹¹ $[\alpha]_D^{20} - 67.7^\circ$, water). IR: ν_{\max} (KBr) 3490(s), 3420(s), 3380(s), 1610(m), 1510(s) cm^{-1} . NMR (pyridine- d_5): δ 2.14 (s, 3 H), 3.9–4.6 (m, 6 H), 5.4–5.6 (m, 1 H), 6.6–7.3 (m, 8 H). UV, nm (ϵ): λ_{\max} (methanol) 281 (1090), 275 (1370), 220 (11 200). The same compound (m.p., specific rotation, IR, NMR, and UV) was obtained when II was hydrogenated under the same conditions.

Dimethyl isobutyltartrate. I (620 mg) was hydrogenated as described above. Evaporation of the solvent gave a residue which was purified by preparative TLC on silica gel using methanol-butanol-chloroform (1:3:6) as eluent. The eluate from the plates was filtered through Dowex 50 W-X8 (H^+ , 1 × 5 cm) using water as eluent. The filtrate was evaporated to dryness giving isobutyltartaric acid, which was esterified with diazomethane. The crude ester was purified by sublimation (0.1 Torr, 45°C) giving dimethyl isobutyltartrate (67 mg) as white needles, m.p. 57–59°C, $[\alpha]_{578}^{25} + 31^\circ$ (c 0.43, methanol). (Found: C 51.4; H 7.88; O 41.1. Calc. for $C_{10}H_{18}O_6$: C 51.3; H 7.75; O 41.0.) IR: ν_{\max} (CHCl₃) 3530(m), 1740(s) cm^{-1} . NMR (CDCl₃+D₂O): δ 0.85 (d, 3 H, *J* 6.5 Hz), 0.95 (d, 3 H, *J* 6.5 Hz) 1.33–2.14 (m, 3 H), 3.73 (s, 3 H), 3.80 (s, 3 H), 4.33 (s, 1 H). MS, *m/e* (rel. intensity): M^+ 234 (absent), 175 (7), 157 (4), 145 (4), 129 (4), 115 (7), 103 (5), 101 (18), 90 (100), 85 (38), 73 (9), 59 (10), 57 (42), 43(33).

Dimethyl (R)-2-isobutylmalate. II (150 mg) was hydrogenated as described above for I.

The catalyst was filtered off and an excess of diazomethane in ether was added. After 1 h, the reaction mixture was evaporated to dryness and the residue was filtered through a column of silica gel (2.5 × 7 cm) using chloroform-methanol (9:1) as eluent. The crude dimethyl ester was purified by preparative GLC (5 % SE-52 on Chromosorb W, 3 mm × 1.8 m, 130 °C, flow rate 40 ml/min, retention time 4 min) giving dimethyl (*R*)-2-isobutylmalate (20 mg) as an oil $[\alpha]_{D}^{25} -0.55^\circ$, $[\alpha]_{436}^{25} +0.30^\circ$, $[\alpha]_{578}^{25} +5.3^\circ$ (c 2.0, ethanol) [Lit.^{13,14} $[\alpha]_{578}^{24} -0.64^\circ$, $[\alpha]_{436}^{24} +0.16^\circ$, $[\alpha]_{565}^{24} +3.9^\circ$ (c 1.0, ethanol)]. IR: ν_{\max} (CHCl₃) 1740 cm⁻¹. NMR (CDCl₃): δ 0.85 (d, 3 H, *J* 6 Hz), 0.95 (d, 3 H, *J* 6 Hz), 1.55–1.75 (m, 3 H), 2.73 (d, 1 H, *J* 16 Hz) and 2.89 (d, 1 H, *J* 16 Hz, AB spectrum), 3.67 (s, 3 H); 3.79 (s, 3 H), 3.2–4.0 (1 H, exchangeable in D₂O).

Dimethyl (±)-erythro-isobutyltartrate. Isobutylmaleic anhydride¹² (191 mg) was dissolved in pyridine (15 ml) and osmium tetroxide (333 mg) was added. The mixture was stirred at room temperature for 35 min after which sodium hydrogen sulfite (0.6 g), pyridine (7 ml), and water (10 ml) were added. After 30 min at room temperature the solution was heated to 60 °C for 15 min. The reaction mixture was evaporated to dryness and the residue was filtered through Dowex 50W–X8 (H⁺, 2.5 × 8 cm) using water as eluent. The dark eluate (180 ml) was extracted with ethyl acetate (6 × 50 ml) and the organic phase was dried and evaporated to dryness. The residue (175 mg) was treated with diazomethane giving the crude ester which was purified by chromatography on silica gel (2.5 × 5.5 cm) using chloroform as eluent. The fraction containing the dimethyl ester was purified by sublimation (0.02 Torr, 45 °C) giving dimethyl (±)-erythro-isobutyltartrate (114 mg), m.p. 80–82 °C, indistinguishable (IR, NMR, MS, and GLC) from the natural product.

Dimethyl (±)-threo-isobutyltartrate. Isobutylfumaric acid¹² (220 mg) was reacted as described above for isobutylmaleic anhydride, except that the heating was omitted. Sublimation (0.02 Torr, 50 °C) gave dimethyl (±)-threo-isobutyltartrate (112 mg) m.p. 50–52 °C. IR: ν_{\max} (CHCl₃) 3530(m), 1740(s) cm⁻¹. NMR (CDCl₃): δ 0.83 (d, 3 H, *J* 6 Hz), 0.94 (d, 3 H, *J* 6 Hz), 1.63–1.92 (m, 3 H), 3.83 (s, 6 H), 4.28 (s, 1 H).

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REFERENCES

- Behr, D., Berg, J.-E., Karlsson, B., Leander, K., Pilotti, A.-M. and Wiehager, A.-C. *Acta Chem. Scand. B* 29 (1975) 401.

- Bourquelot, E. and Bridel, M. *C. R. Acad. Sci.* 168 (1919) 701.
- Delauney, P. *C. R. Acad. Sci.* 171 (1920) 435.
- Delauney, P. *J. Pharm. Chim.* 23 (1921) 265.
- Delauney, P. *C. R. Acad. Sci.* 176 (1923) 598.
- Delauney, P. *C. R. Acad. Sci. Ser. C.* 180 (1925) 224.
- Charaux, C. and Delauney, P. *C. R. Acad. Sci.* 180 (1925) 1770.
- Delauney, P. *Contribution à l'étude des glucosides de la famille des Orchidées*, Diss., Univ. Paris 1924.
- Karrer, P. and Matter, E. *Helv. Chim. Acta* 30 (1947) 2096.
- Björndal, H., Hellerqvist, C. G., Lindberg, B. and Svensson, S. *Angew. Chem. Int. Ed. Engl.* 9 (1970) 610.
- Semke, L. K., Thompson, N. S. and Williams, D. G. *J. Org. Chem.* 29 (1964) 1041.
- Vaughan, W. R. and Andersen, K. S. *J. Amer. Chem. Soc.* 77 (1955) 6702.
- Brandänge, S., Luning, B., Moberg, C. and Sjöstrand, E. *Acta Chem. Scand.* 25 (1971) 349.
- Brandänge, S., Josephson, S. and Vallén, S. *Acta Chem. Scand. B* 28 (1974) 153.
- Gäumann, E. and Kern, H. *Phytopathol. Z.* 35 (1959) 347.
- Gäumann, E. and Kern, H. *Phytopathol. Z.* 36 (1959) 1.
- Hardegger, E., Schellenbaum, M. and Corrodi, H. *Helv. Chim. Acta* 46 (1963) 1171.

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