The Structure of Caperatic Acid SVANTE BRANDÄNGE, LARS MÖRCH and STAFFAN VALLÉN

Department of Organic Chemistry, Arrhenius Laboratory, University of Stockholm, S-104 05 Stockholm, Sweden

The lichen compound caperatic acid (1) was first isolated from Parmelia caperata in 1897,¹ and has subsequently been found in many other lichens.²-⁵ Zopf ⁵ isolated an acid believed to be I from Cetraria glauca and Mycoblastus sanguinarius, and we have now confirmed that I can be isolated from the former species. Asano and Ohta found that the acid is a monomethyl ester of 2-hydroxy-1,2,3-heptadecanetricarboxylic acid,⁵ but the position of the methyl ester group has hitherto been unknown. We now report that caperatic acid has the structure I.

This result is based upon a comparison of the mass spectra of the trimethyl ester 2 and its corresponding d_4 -ester 3 (Scheme 1), and

m/e = 273 (69) m/e = 339 (100) m/e = 342 (79)
[2: 270 (44)] [2: 339 (100)]

Scheme 1.

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also upon a chemical degradation of 1. In the upper parts of the mass spectra of 2 and 3 there are, respectively, three and four prominent peaks, and from the m/e values of these peaks, the structure of caperatic acid was deduced (Scheme 1). Support for the degradations shown in Scheme 1 was obtained from high resolution mass spectrometry of 2 and from the existence of analogous prominent peaks in the mass spectra of homologous trimethyl alkylcitrates.

In a corroborative chemical degradation, 1 was oxidized with sodium bismuthate, a reagent which oxidizes α -hydroxy carboxylic acids to aldehydes or ketones. An NMR spectrum of the crude reaction product suggested that its major component was methyl 3-oxo-octadecanoate, the decarboxylation product of the initially formed β -keto carboxylic acid.

A CĎ investigation has shown that caperatic acid forms a complex with molybdate (VI) at pH 5.7.7 This demonstrates that 1 contains a free α-hydroxy carboxylic acid group.9

free α-hydroxy carboxylic acid group. Experimental. Melting points are corrected. The NMR spectrum was recorded on a Varian XL-100 spectrometer and the mass spectra on a Varian MAT 311 instrument using the direct

inlet system.

Isolation of 1 from Cetraria glauca. From 300 g of fresh lichen, collected 30 km northeast of Stockholm, 1.6 g of 1 was isolated. After four recrystallisations from ethanol the product melted at $132.5-134\,^{\circ}\mathrm{C}$ (lit.6 m.p. $132-133.5\,^{\circ}\mathrm{C}$) $[\alpha]_{\mathrm{D}^{23}}-15.5\,^{\circ}$ (c=0.9, dioxane). An authentic sample of 1, isolated from Parmelia caperata, was obtained from L. Tibell, University of Uppsala, and this sample showed $[\alpha]_{\mathrm{D}^{23}}-15.7\,^{\circ}$ (c=0.6, dioxane). The optical rotations of 1 have previously been measured in chloroform,6,10 but we found that all samples of 1 were only slightly soluble in ordinary ethanol-stabilized chloroform (c<0.1). Reaction with diazomethane in ether solution afforded the trimethyl ester 2, m.p. $55-56\,^{\circ}\mathrm{C}$ (from ethanol), lit.6 m.p. $56.5-57.5\,^{\circ}\mathrm{C}$. MS: m/e (rel. intensity) 371 (33), 339 (100), 270 (44), 101 (48), 87 (49), 43 (37).

The d₄-ester 3 was prepared by reaction of 1-d₃ with diazomethane-d₂. The former reactant was obtained by dissolving 1 in a mixture of CH₃OD (99 % isotopic purity) and dry ether, followed by evaporation of the solvent. Diazomethane-d₂ was prepared largely after Hecht and Kozarich,¹¹ but the N-nitrosomethylurea was replaced with N-methyl-N-nitroso-p-toluene-sulfonamide and, instead of using the dimethoxyethane solution, dry ether was added and the diazomethane-d₂ was distilled together with ether into a flask containing a little D₂O (99.7 % isotopic purity). The water in the receiving flask was frozen out and the ethereal layer was then added to 1-d₃. The resulting ether solution was washed with water, dried (Na₂SO₄) and concentrated. The product 3 was crystallised from ethanol, m.p. 55.5—

56.5°C. MS: 374(71), 342(79), 339(100), 273(69), 101(33), 90(42).

Bismuthate oxidation of 1. A mixture of 1 (55 mg), sodium bismuthate (56 mg), and glacial acetic acid (2 ml) was warmed (35 $^{\circ}$ C, 80 min) with stirring in a stoppered flask. The resulting solution was poured into a mixture of cold water (100 ml) and ether (50 ml). The ether layer was repeatedly washed with sodium hydrogen carbonate solution and then twice with water. After drying (Na₂SO₄) and concentration of the ether layer, and finally drying in a vacuum desiccator over night, a residue (27 mg) was obtained. The NMR spectrum of this residue (CDCl₃) displayed singlets at $\delta = 3.66$ and 3.38, and triplet at $\delta = 2.46$ (integrals approximately 35, 18 and 21 mm, respectively), which indicate the partial structure $-CH_2-CH_2-CO-CH_2-COOCH_3$. No peaks from an isomeric β -keto ester could be detected.

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