exactly a third of the total radioactivity of the malonate which is only compatible with the formation of malonate exclusively by oxidative α-decarboxylation, i.e. according to route 1 or 2. In the second experiment using uniformly labelled aspartate a slightly higher radioactivity was obtained in the methylene group as compared to the radioactivity of each carboxyl group which might indicate some contribution of route 4 to the malonate formation. Calculations from the figures in the table show that 77 % of the labelling in the malonate is derived from route 1 or 2 and 23 % from route 4.

The experiments demonstrate thus that there is a metabolic pathway in *P. islandicum* from oxalacetate to malonate in which oxidative \( \alpha\)-decarboxylation of oxalacetate is involved. The question whether this reaction is related to the oxidative decarboxylation of pyruvate or is of peroxidase character can only be settled when the reaction is studied in a cell free system.

- Gibson, D. M., Titchener, E. B. and Wakil, S. J. Biochem. Biophys. Acta 30 (1958) 376.
- Hatch, M. D. and Stumpf, P. K. J. Biol. Chem. 236 (1961) 2879.
- de Vellis, J., Shannon, L. M. and Lew, J. Y. Plant Physiol. 38 (1963) 686.
- Shannon, L. M., de Vellis, J. and Lew, J. Y. Plant Physiol. 38 (1963) 691.
- Ebert, E. and Zenk, M. H. Phytochemistry. 6 (1967) 309.
- Gatenbeck, S. Acta Chem. Scand. 16 (1962) 1053.
- Van Slyke, D. D. and Folch, J. J. Biol. Chem. 136 (1940) 509.
- Phares, E. F. Arch. Biochem. Biophys. 33 (1951) 173.

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## Chemical Studies on Lichens

## 12.\* A New Lichen Xanthone from Lecanora reuteri JOHAN SANTESSON

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A rapidly growing number of xanthones, most of which chlorinated, are known from lichens. Some examples are lichexanthone (II),¹ thiophanic acid (III),²-⁴ arthothelin (IV),⁵-² and thuringion (V),²-8 All these are derived from the same parent compound norlichexanthone (I, 1,3,6-tri-hydroxy-8-methylxanthene-9H-9-one).

During a screening of Lecanora species by "lichen mass spectrometry" it was found that the spectrum of L. reuteri Schaer. (Fig. 1) exhibited a strong peak at m/e 258 which suggested the presence of norlichexanthone (M=258). A peak at m/e 360 with satellite peaks at m/e 362, 364, and 366 indicated the presence of the corresponding trichlorinated xanthone arthothelin (IV).

I R=H II R=CH3

III R=H R'=CI

IV R=R1=H

V R=CH3 R'=H

<sup>\*</sup> Part 11. Arkiv Kemi. In press.

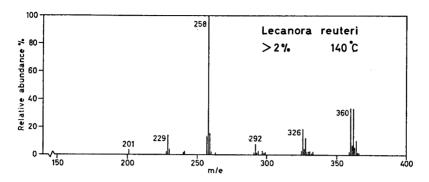


Fig. 1.

These tentative identifications were verified by isolation of the compounds and comparison with authentic specimens.

Norlichexanthone has previously not been found in Nature. It has been prepared as an intermediate in the synthesis of lichexanthone 1 and thiophanic acid.3

Arthothelin is known from Arthothelium pacificum Follm., Lecidea quernea (Dicks.) Arn., and some Lecanora species. 3-7

Experimental. Lichen samples: Lecanora reuteri (A) from Unterwalden, Switzerland, reference designation R. Sant. 1946; (B) Jura, Switzerland, Frey 1929. Specimens are to be found in the herbarium of Uppsala Botanical Museum (UPS).

Dry and ground lichen (sample A, 1.2 g) was extracted with acetone (20 ml, 12 h), the solution concentrated to 5 ml, and ether (20 ml) added. This solution was extracted with sodium hydrogen carbonate solution (5 %,  $2 \times 5$  ml) and then with sodium carbonate solution (10 %,  $2 \times 5$  ml).

Acidification of the NaHCO<sub>3</sub> solution (2 N hydrochloric acid), ether extraction, and preparative TLC (silica gel G, dichloromethane-acetone 4:1) afforded arthothelin (IV, 1.1 mg), identified by comparison with an authentic sample (IR spectra, co-chromatography).

Acidification of the Na<sub>2</sub>CO<sub>3</sub> solution (2 N hydrochloric acid), ether extraction, and recrystallisation from ethanol-water of the residue obtained after evaporation of the

ether, yielded norlichexanthone (I, 3.0 mg), m.p. 272-74°, identified by comparison with

a synthetic <sup>3</sup> sample (IR, mixed m.p.'s). In an acetone extract of the lichen (sample B), I and IV were identified by co-chromatography (TLC according to Ref. 9) with authentic samples.

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- Asahina, Y. and Nogami, H. Bull. Chem. Soc. Japan. 17 (1942) 202.
- 2. Huneck, S. Tetrahedron Letters 1966 3547.
- Santesson, J. and Sundholm, G. Arkiv Kemi. In press.
- 4. Arshad, M. and Ollis, W. D. To be published. 5. Huneck, S. and Follmann, G. Z. Natur-
- Huneck, S. and Follmann, G. Z. Naturforsch. 22b (1967) 461.
- 6. Santesson, J. To be published.
- 7. Huneck, S. and Santesson, J. To be published.
- 8. Santesson, J. Arkiv Kemi. In press.
- Santesson, J. Acta Chem. Scand. 21 (1967) 1162.

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