## Preparation of Ursodeoxycholic Acid and $3\alpha, 7\beta, 12\alpha$ Trihydroxycholanic Acid

Bile Acids and Steroids 94

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The compounds have been prepared through reduction of the corresponding 7-ketoacids with sodium in n-propanol. Reversed phase partition chromatography of the reduction products revealed that the epimeric  $7\beta$ - and 7a-hydroxycompounds are formed in the proportion of about 3:1.

Ursodeoxycholic acid (3  $\alpha$ ,  $7\beta$ -dihydroxycholanic acid) is found in the bile of certain bear species 1 and in the coypu 1. Traces of this acid has recently also been reported to occur in the rat 2 and man 3. Urxodeoxycholic acid has previously been prepared synthetically through reduction of 7-ketolithocholic acid (3a-hydroxy-7-ketocholanic acid) with sodium ethoxide and through catalytic hydrogenation in acidic medium. The yield of the  $7\beta$ -epimer was only about 10 % 4. Sodium borohydride reduction of 7-ketocholanic acids and derivatives produces specifically the corresponding  $7\alpha$ -hydroxycompounds  $^{5-8}$ . The 7-ketone in the bile acid series has been reported to be almost resistent to reduction with aluminium isopropoxide 8,9. In one case, however, a successful reduction, which gave predominately the axial epimer  $(7\alpha)$ , has been described 10. These authors also reported the preparation of ursodeoxycholic acid in 95 % yield by reduction of 7-ketolithocholic acid with sodium in npropanol. When this method was used in the present investigation and the reaction product separated by reversed phase partition chromatography, it appeared that the reduction gives a mixture consisting of the equatorial  $7\beta$ - and axial  $7\alpha$ -hydroxyepimers in the approximate proportion of 3:1.

 $3\alpha,7\beta,12\alpha$ -trihydroxycholanic acid has never been synthesized or isolated before. Recently, however, an acid with identical chromatographic properties and sulfuric acid spectrum has been isolated as a metabolite of cholic acid-24-14C in the rat 11 and has been detected in samples of human bile 3.

 $3\alpha,7\beta,12\alpha$ -trihydroxycholanic acid was obtained by reduction of 7-ketode-oxycholic acid  $(3\alpha,12\alpha$ -dihydroxy-7-ketocholanic acid) by the same method

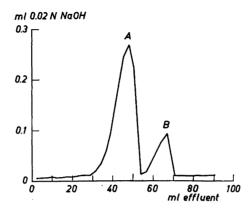


Fig. 1. Chromatography of an aliquot of the reaction product obtained by reduction of 7-ketolithocholic acid with sodium in n-propanol. Column: 4.5 g of hydrophobic SuperCel. Moving phase: 55 % (v/v) aqueous methanol. Stationary phase: 10 % (v/v)-heptane in chloroform. A: Ursodeoxycholic acid. B: Chenodeoxycholic acid.

as above. The two epimers, separated chromatographically, were found in the same proportion as in the reduction of 7-ketolithocholic acid. The methyl ester of  $3\alpha,7\beta,12\alpha$ -trihydroxycholanic acid yielded a tri-p-nitrobenzoate on acylation with p-nitrobenzoyl chloride in pyridine. Under similar conditions methyl cholate only gives the  $3\alpha$ -mono-p-nitrobenzoate <sup>12</sup>.

## **EXPERIMENTAL**

Ursodeoxycholic acid. 7-Ketolithocholic acid (m. p. 200-202°) was prepared through sodium dichromate oxidation of methyl 3a-cathyloxy-7a-hydroxycholanate followed by hydrolysis <sup>13,14</sup>. A solution of 0.5 g of the ketoacid in 10 ml of anhydrous n-propanol was refluxed with 1 g of sodium for 3 h. The solution was diluted with water, acidified with hydrochloric acid and extracted with ethylacetate. After evaporation to dryness in vacuo the residue was chromatographed on hydrophobic Super Cel with 55 % (v/v) methanol as moving phase and 10 % (v/v) heptane in chloroform as stationary phase <sup>16</sup> (Fig. 1).

No unchanged ketoacid could be detected and of the two peaks which appeared, the first one (77 %) was crystallized from ethylacetate/light petroleum. Yield: 0.32 g, m. p. 201-202°. There was no depression of the m. p. on admixture with authentic ursodeoxycholic acid that was kindly supplied by Prof. G. A. D. Haslewood.  $[a]_{15}^{15} = +57 \pm 2^{\circ}$ 

(c 1.02, dioxane). (Found: C 73.1; H 10.3. Calc. for C<sub>24</sub>H<sub>40</sub>O<sub>4</sub>: C 73.4; H 10.3). The material in the second peak (23 %) with an elution volume characteristic of chenodeoxycholic acid was crystallized from ethylacetate/light petroleum. M. p. 141-

142°, undepressed by authentic chenodeoxycholic acid.

3a, 7β, 12a-trihydroxycholanic acid. Methylcholate was oxidized with N-bromosuccinimide 16 to methyl-7-ketodeoxycholate, purified via the diacetate 17 (m. p. 118—118.5°) and saponified. 7-Ketodeoxycholic acid was crystallized according to the directions given by Hoehn and Linsk 16; m. p. 198—199°. The ketoacid was reduced as described according to the directions given by Hoehn and Linsk 16; m. p. 198—199°. cribed above and the reaction product was separated by reversed phase partition chromatography on hydrophobic Super Cel. Aqueous methanol (50 % (v/v)) was used as the moving phase and isocctanol-chloroform (1:1) as stationary phase <sup>10</sup>. (Fig. 2.) The more hydrophilic compound (72 %),  $3a,7\beta,12a$ -trihydroxycholanic acid (0.2 g), was dissolved in 3 ml methanol, neutralized with 0.1 N NaOH and added slowly to 100 ml of cold

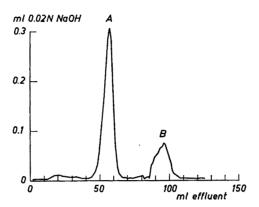


Fig. 2. Chromatography of an aliquot of the reaction product obtained by reduction of 7-ketodeoxycholic acid with sodium in n-propanol. Column: 4.5 g of hydrophobic Super Cel. Moving phase: 50 % (v/v) aqueous methanol. Stationary phase: isocctanol / chloroform (1:1). A:  $3a,7\beta,12a$ -trihydroxycholanic acid. B: Cholic acid.

0.05 N HCl, with vigorous stirring. The precipitated acid crystallized after standing 12 h at  $+4^{\circ}$ . 0.155 g (56 % yield calculated on 7-ketodeoxycholic acid) was obtained after filtration, washing with water and drying; m. p. 127-129°. The m. p. was not changed after repeated recrystallizations by this method. [a]<sub>5</sub><sup>25</sup> =  $+62 \pm 2^{\circ}$  (c 1.13, dioxane). (Found C 70.3; H 10.0. Calc. for C<sub>24</sub>H<sub>40</sub>O<sub>5</sub>: C 70.5; H 9.9.)

A sample of this acid was crystallized from ethylacetate/heptane and dried at  $130^{\circ}$  at 0.001 mm Hg for 24 h; m. p.  $157-158^{\circ}$  (decomp.). [a] $_{10}^{25}=+57\pm2^{\circ}$  (c 1.06, dioxane). The elemental analysis indicated that the acid had formed a complex with heptane. (Found C 72.7; H 10.4. Calc. for  $C_{24}H_{40}O_5+C_7H_{16}$ : C 73.2; H 11.1). Several other organic solvents were tried for recrystallization without success.

Methyl  $3a,7\beta,12a$ -trihydroxycholanate-tri-p-nitrobenzoate. The method of Borsche <sup>12</sup> was followed. A solution of 75 mg of methyl  $3a,7\beta,12a$ -trihydroxycholanate, prepared with diazomethane, was dissolved in 2 ml of dry pyridine and cooled to  $0^\circ$ . 180 mg of p-nitrobenzoylchloride in 2 ml of pyridine was added. After standing at room temperature for 12 h the reaction mixture was transferred to a mixture of ether, ethylacetate, ice and 2 N HCl. The ether layer was washed successively with 2 N HCl, 5 % aqueous sodium carbonate and water. After evaporation to dryness in vacuo the residue was crystallized from ethylacetate/ethanol to yield 121 mg of fine needles, m. p. 188—189°. The concentrated mother liquor gave an additional 24 mg of crystals with the same m. p. Total yield: 94% [a] $^{13}_{10} = +140 \pm 3^\circ$  (c 1.19, dioxane). (Found C 63.5; H 6.0; N 4.8. Calc. for  $C_{48}H_{51}O_{14}N_5$ : C 63.5; H 5.9; N 4.8).

The second compound (B in Fig. 2) comprising about 28 % of the two epimers, was eluted at the place of cholic acid. It was crystallized from ethylacetate, m. p. 197—198°, undepressed by authentic cholic acid.

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## REFERENCES

- 1. Haslewood, G. A. D. Physiol. Rev. 35 (1955) 178.
- 2. Mahowald, T. A., Yin, M. W., Matschiner, J. T., Hsia, S. L., Doisy, E. A. Jr., Elliott, W. H. and Doisy, E. A. J. Biol. Chem. 230 (1958) 581.

  3. Sjövall, J. Acta Chem. Scand. 13 (1959) 711.

  4. Miyazi, S. Z. Physiol. Chem. Hoppe-Seyler 250 (1937) 31.

  5. Mosbach, E. H., Meyer, W. and Kendall, F. E. J. Am. Chem. Soc. 76 (1954) 5799.

- Mossach, E. H., Meyer, W. and Kendan, F. E. J. Am. Chem. Soc. 16 (1954) 5799.
   Samuelsson, B. Acta Chem. Scand. 13 (1959). 976.
   Bergström, S., Lindstedt, S. and Samuelsson, B. J. Biol. Chem. 234 (1959) 2022.
   Hsia, S. L., Matschiner, J. T., Mahowald, T. A., Elliott, W. H., Doisy, E. A. Jr., Thayer, S. A. and Doisy, E. A. J. Biol. Chem. 225 (1957) 811.
   Tukamoto, M. J. Biochem. (Japan) 32 (1940) 461.
   Kanazawa, T., Shimazaki, A. Sato, T. and Hoshino, T. Proc. Japan Acad. 30 (1954)
- 391.

- Norman, A. and Sjövall, J. J. Biol. Chem. 233 (1958) 872.
   Borsche, W. Ber. 57 (1924) 1620.
   Fieser, L. F., Herz, J. E. Klohs, M. W., Romero, M. A. and Utne, T. J. Am. Chem. Soc. 74 (1952) 3309.

- 14. Samuelsson, B. Acta Chem. Scand. 13 (1959) 236.
  15. Bergström, S. and Sjövall, J. Acta Chem. Scand. 5 (1951) 1267.
  16. Fieser, L. F. and Rajagopalan, S. J. Am. Chem. Soc. 71 (1949) 3935.
  17. Corey, E. J. J. Am. Chem. Soc. 76 (1954) 175.
  18. Hoehn, W. M. and Linsk, J. J. Am. Chem. Soc. 67 (1945) 312.
- 19. Norman, A. Acta Chem. Scand. 7 (1953) 1413.

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