The Formation of Bile Acids from 7 a-Hydroxycholesterol in the Rat *

Bile Acids and Steroids 47

SVEN LINDSTEDT

Department of Physiological Chemistry, University of Lund, Lund, Sweden

Tritium labelled 7a-hydroxycholesterol has been prepared. It has been injected intraperitoneally into bile fistula rats and cholic and chenodeoxycholic acid identified as acid metabolities. The mechanism of the degradation of cholesterol is discussed.

In the autoxidation of cholesterol the two epimeric 7-hydroxycholesterols are formed ^{2,3}. They have also been reported to occur in small amounts in the unsaponifiable fraction in various organ lipids 4-7. The possibility of their formation by autoxidation during the isolation procedure has, however, not been rigorously excluded.

It has been suggested that a 7-hydroxycholesterol may appear as an intermediate in the in vivo degradation of cholesterol 38.

In this work the formation of bile acids from 7α-hydroxycholesterol is demonstrated.

EXPERIMENTAL

7a-Hydroxycholesterol was prepared essentially according to Henbest et al. 9,10. 2 g of sodium formiate dissolved in 20 ml of formic acid was added to a solution of 2 g of 7a-bromocholesterylbenzoate in 20 ml of ether. After 24 h water was added and the steroid extracted with ether. The material was dissolved in 2 ml of benzene and 15 ml of methanol added. After 8 h at room temperature 1.35 g of crystals melting at $115-144^\circ$ were obtained. This material was brought onto a 20 g alumina column and eluted with increasing concentration of benzene in light petroleum. The material eluted with 0-30~%of benzene in light petroleum was crystallized from benzene/methanol and yielded 0.91 g of 7a-formoxycholesterolbenzoate m. p. 150-152°. Saponification with alcoholic potassium hydroxide gave 7a-hydroxycholesterol m. p. 150°.

Tritium-labelled 7a-hydroxycholesterol. Attempts to label 7a-hydroxycholesterol by platinum catalyzed exchange in tritiated water/acetic acid or tritiated water-ethanol/

^{*} For a discussion of the steric configuration at the 7-position see Ref.¹. A preliminary report of this work was read at the meeting of the Norwegian Biochemical Society, June 1956 20.

sodium hydroxide were not successful and led to extensive destruction. The labelled 7a-hydroxycholesterol was therefore synthesized by the above procedure from tritium-labelled cholesterol prepared according to Bloch ¹¹. To ensure the absence of radioactive contaminants it was chromatographed according to Danielsson ¹² on hydrophobic kieselguhr with 50 % (v/v) aqueous isopropanol as moving phase and 20 % (v/v) chloroform in heptane as stationary phase. The tritium labelled 7a-hydroxycholesterol had a specific activity of 4.3×10^{5} c.p.m. when counted in an "infinitely thin" layer in a gas-flow counter.

Chromatographic separations of the bile acids were performed according to Sjövall and Norman ^{13,14}. The following two systems were used:

Phase system	moving phase	stationary phase
$C(Ref.^{14})$	methanol/water 50/50	isooctanol/chloroform 1/1
$F(Ref.^{13})$	methanol/water 55/10	chloroform/heptane 9/1

Each fraction was titrated with 0.02 N sodium hydroxide and an aliquot plated on a copper planchet for counting in a gas-flow counter. Accurate determinations of specific activity were performed by the gas-phase counting technique of Glascock ¹⁵.

RESULTS

2—3 mg of tritium-labelled 7α -hydroxycholesterol was dissolved in 0.2 ml of ethanol and brought into a colloidal suspension by the addition of 2 ml of 0.9 % sodium chloride containing 2 mg of sodium oleate. It was injected intraperitoneally into white rats with a bile fistula, and the bile collected.

After hydrolysis in 2 N sodium hydroxide in a sealed glass tube at 120° for 8 h the bile acids were extracted and isolated by chromatography with phase system C.

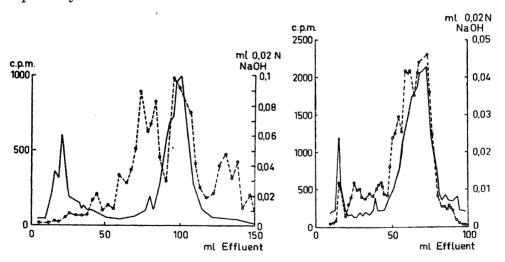


Fig. 1. Chromatogram of hydrolyzed bile after intraperitoneal injection of tritium-labelled 7a-hydroxycholesterol. Phase system C. Broken line: isotope determination. Solid line: titration values.

Fig. 2. Chromatogram of dihydroxy-acids after intraperitoneal administration of tritium-labelled 7a-hydroxycholesterol. Phase system F. Broken line: isotope determination, Solid line: titration values.

Fig. 1 shows a chromatogram where the titration peak of cholic acid appears at 80—120 ml. At the same position a peak of radioactivity is seen. This material was collected, rerun with the same chromatographic system, diluted with carrier cholic acid and recrystallized from different solvents. From each crystallization material was taken for determination of the specific activity by the gas-phase counting method ¹⁵. It remained constant throughout five recrystallizations (EtOH, EtOAc, acetone/water) and the identity of the material with cholic acid seems established.

The material remaining in the column was eluted and rechromatographed on system F suitable for the separation of dihydroxycholanic acids. The chromatogram which is seen in Fig. 2 shows a titration peak at 50—80 ml caused by chenodeoxycholic acid in the bile. The identity of the labelled compound eluted at this position with chenodeoxycholic acid was established by recrystallization with carrier and determination of specific activity as for cholic acid.

In Fig. 1 labelled material is eluted before and after the cholic acid peak. When labelled chenodeoxycholic acid is administered to rats labelled metabolites appear at these locations ^{16,21} and in this experiment they probably also represent metabolites of the chenodeoxycholic acid.

DISCUSSION

In the rat cholesterol (I) gives rise to cholic acid (III) and chenodeoxycholic acid (IV) 16,17. Work by Bergström and Sjövall 16 has, however, established that cholic acid is not formed by a 12a-hydroxylation of chenodeoxycholic acid. This type of reaction has been shown to take place when $3\alpha,7\alpha$ -dihydroxycoprostane is administered leading to the formation of cholic acid as well as of chenodeoxycholic acid 18. This has brought further support to the view that in the formation of bile acids from cholesterol hydroxylations in the steroid nucleus precede the total degradation of the side chain 19.

Although the in vivo formation of 7α-hydroxycholesterol (II) from cholesterol has never been demonstrated, the results suggest the possibility that one of the primary steps in the degradation of cholesterol in the rat is a 7-hydroxylation.

A summary of some of the reactions studied in the rat is given in Fig. 3.

REFERENCES

- 1. Heymann, H. and Fieser, L. F. Helv. Chim. Acta 35 (1952) 631.
- 2. Bergström, S. and Wintersteiner, O. J. Biol. Chem. 141 (1951) 597.

3. Bergström, S. Arkiv Kemi Mineral. Geol. 16 A (1943) No. 10.

- Haslewood, G. A. D. Biochem. J. London 33 (1939) 709.
 Wintersteiner, O. and Ritzmann, J. R. J. Biol. Chem. 136 (1940) 697.
 Hardegger, E., Ruzicka, L. and Tagmann, E. Helv. Chim. Acta 26 (1943) 2205.

- Haldegger, B., Markett, E. and Taginani, E. Hete. Chim. Acta 26 (1943) 2222.
 Prelog, V., Ruzicka, L. and Stein, P. Helv. Chim. Acta 26 (1943) 2222.
 Haslewood, G. A. D. Biochem. J. London 36 (1942) 389.
 Bide, A. E., Henbest, H. B., Jones, E. R. H., Peevers, R. W. and Wilkinson, P. A. J. Chem. Soc. 1948 1783.
- 10. Henbest, H. B. and Jones, E. R. H. J. Chem. Soc. 1948 1792.
- 11. Danielsson, H. Acta Chem. Scand. In press.
- Bloch, K. and Rittenberg, D. J. Biol. Chem. 149 (1943) 505.
 Sjövall, J. Acta Physiol. Scand. 29 (1953) 232.
 Norman, A. Acta Chem. Scand. 7 (1953) 1413.

- 15. Glascock, R. F. Isotopic Gas Analysis for Biochemists, Academic Press Inc., New York
- 16. Bergström, S. and Sjövall, J. Acta Chem. Scand. 8 (1954) 611.
- 17. Bergström, S. Proc. Roy. Physiograph. Soc. Lund. 22 (1952) No. 16.
- 18. Bergström, S. and Lindstedt, S. Biochim. et Biophys. Acta 19 (1956) 556. 19. Bergström, S. Record Chem. Progr. Kresge-Hooker Sci. Lib. 16 (1955) 63.

- Bergström, S. and Lindstedt, S. Acta Chem. Scand. 10 (1956) 1044.
 Mahowald, T. A., Elliott, W. W. and Thayer, S. A. Federation Proc. 14 (1950) 250.

Received February 2, 1957.